

# **FROM PETRI NETS TO PARTIAL DIFFERENTIAL EQUATIONS**

**- SPATIAL MODELLING IN SYSTEMS BIOLOGY -**

**Monika Heiner**

**Brandenburg University of Technology  
Computer Science Institute**

## □ FRAMEWORK

- > unifying paradigms:  
(coloured) QPN - SPN - CPN - HPN
- > (bio) processes evolving in time and space
- > *How to encode space ?*

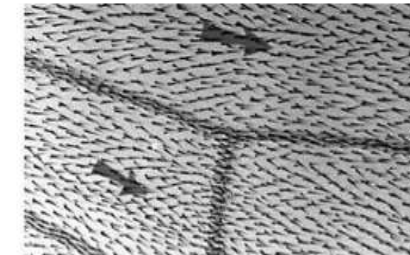
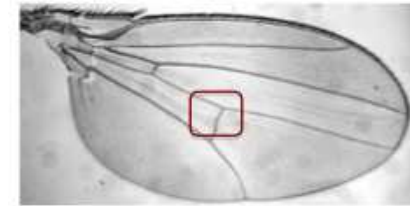
## □ COLOURING SPACE (VERSION 1)

- > diffusion in space
- > Turing patterns
- > phase variation in multistrain bacterial colonies
- > planar cell polarity in fly wing

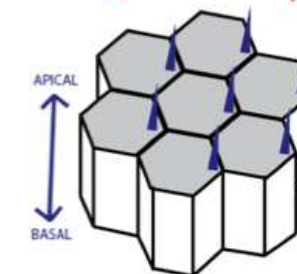
## □ LIMITATION -> VERSION 2

## □ SUMMARY & OUTLOOK

- > next steps

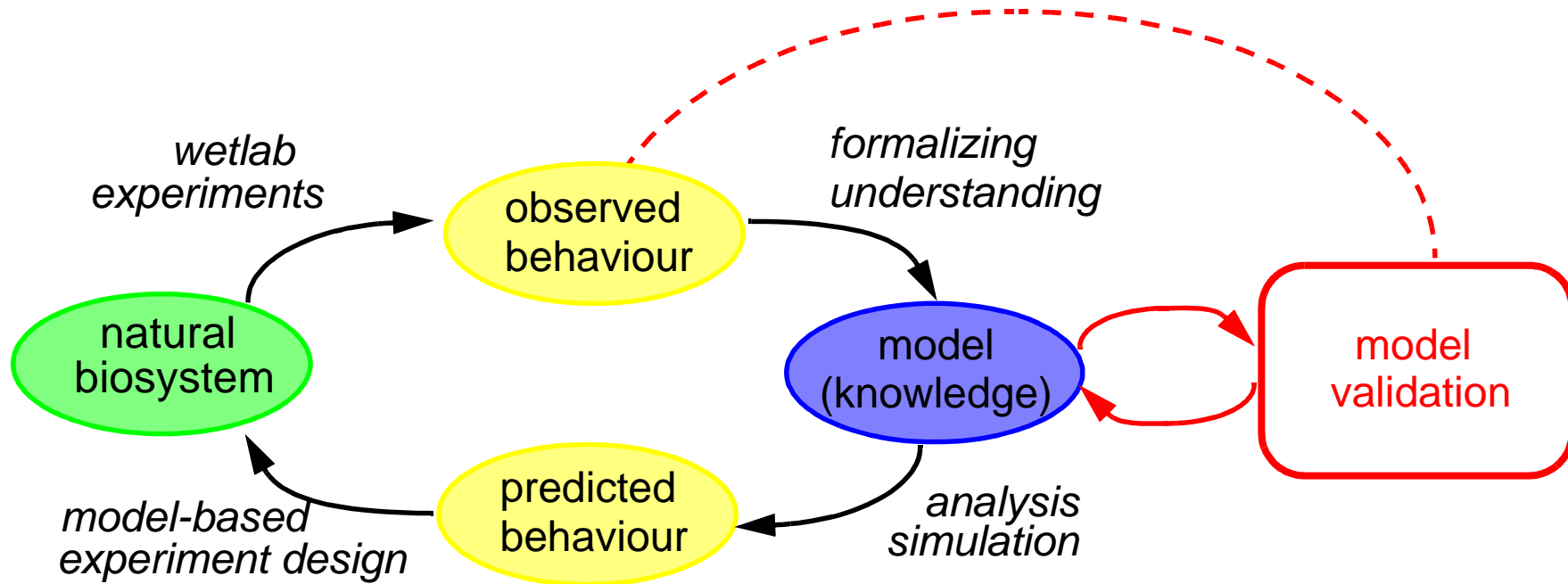


PROXIMAL ← → DISTAL



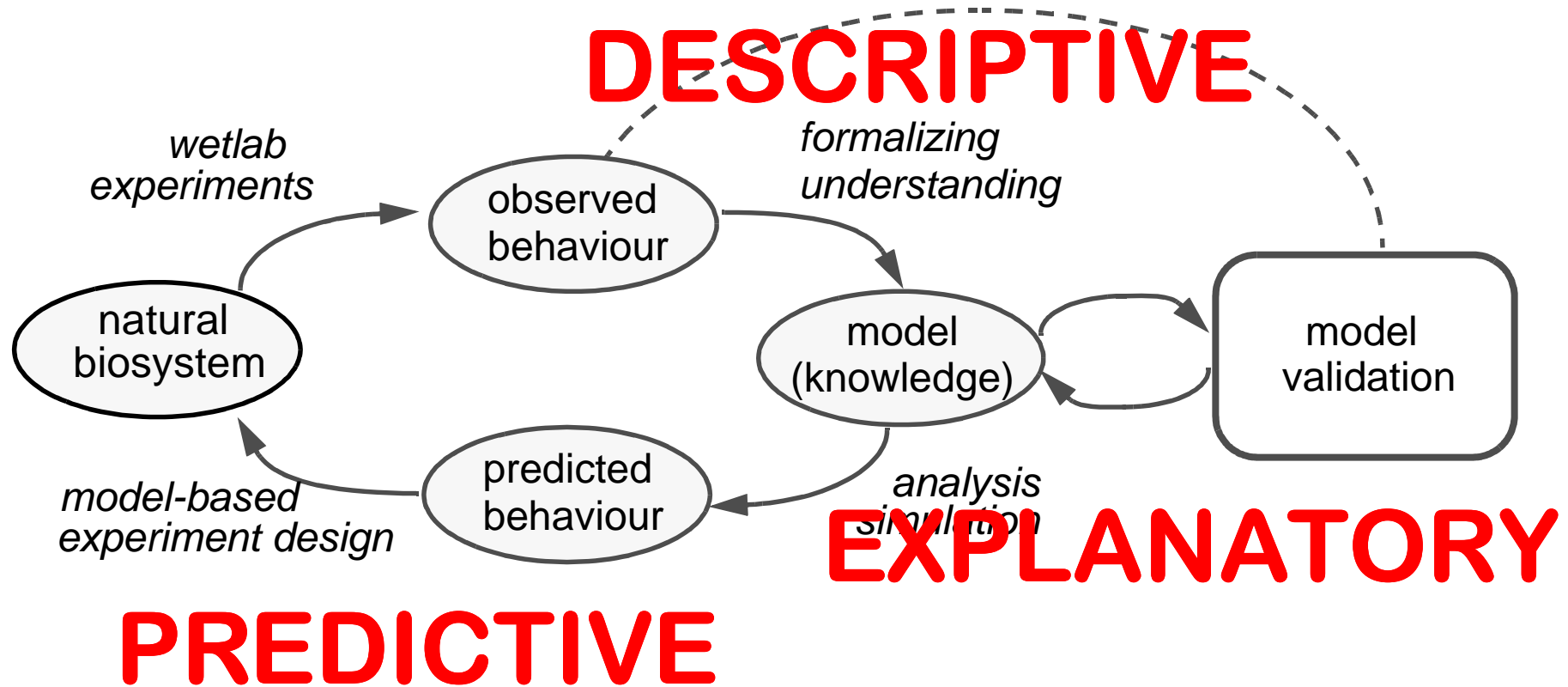
# THE FRAMEWORK

**MODELLING = FORMAL KNOWLEDGE REPRESENTATION**

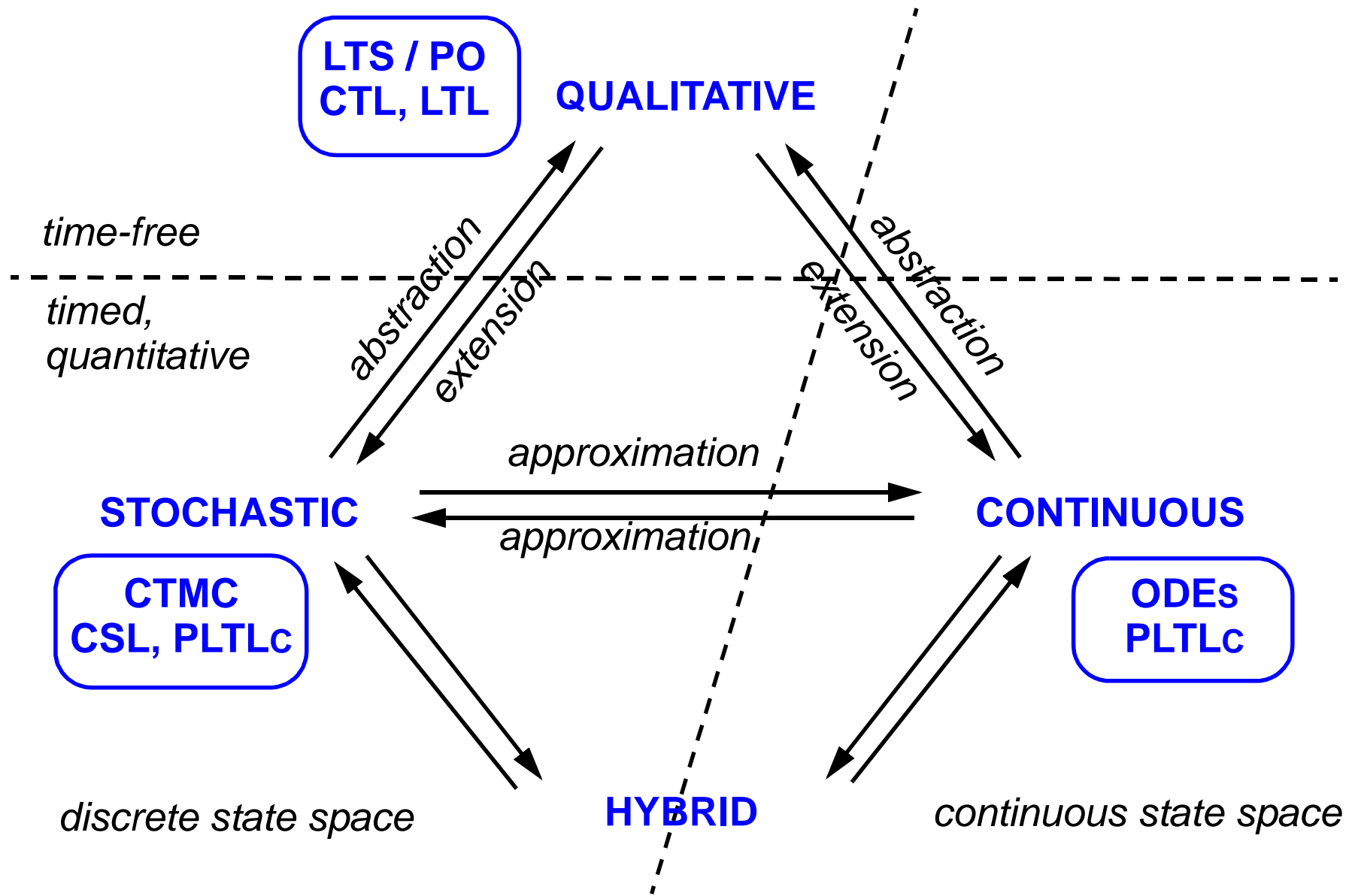


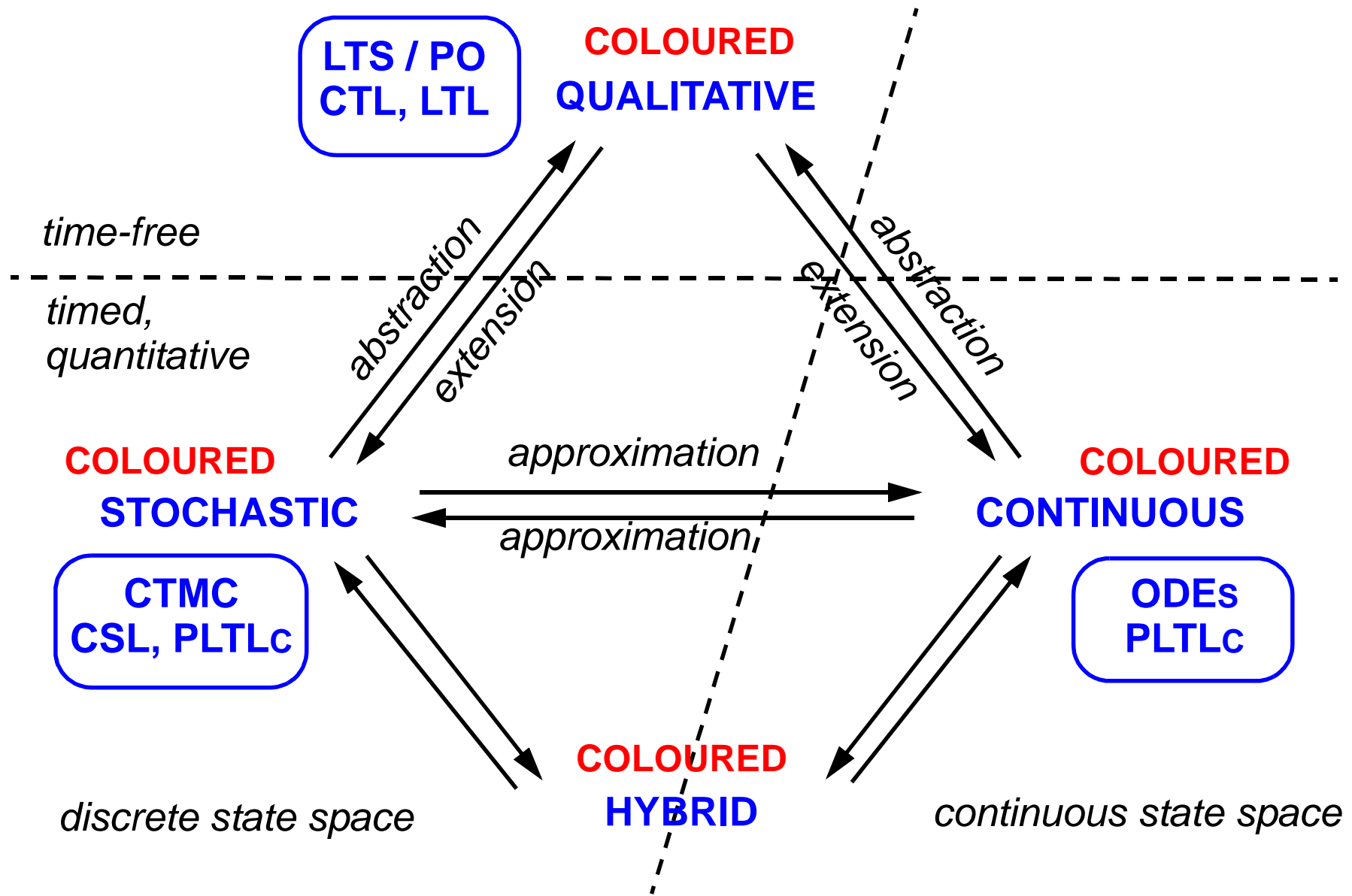
**MODEL VALIDATION = CONFIDENCE INCREASE**

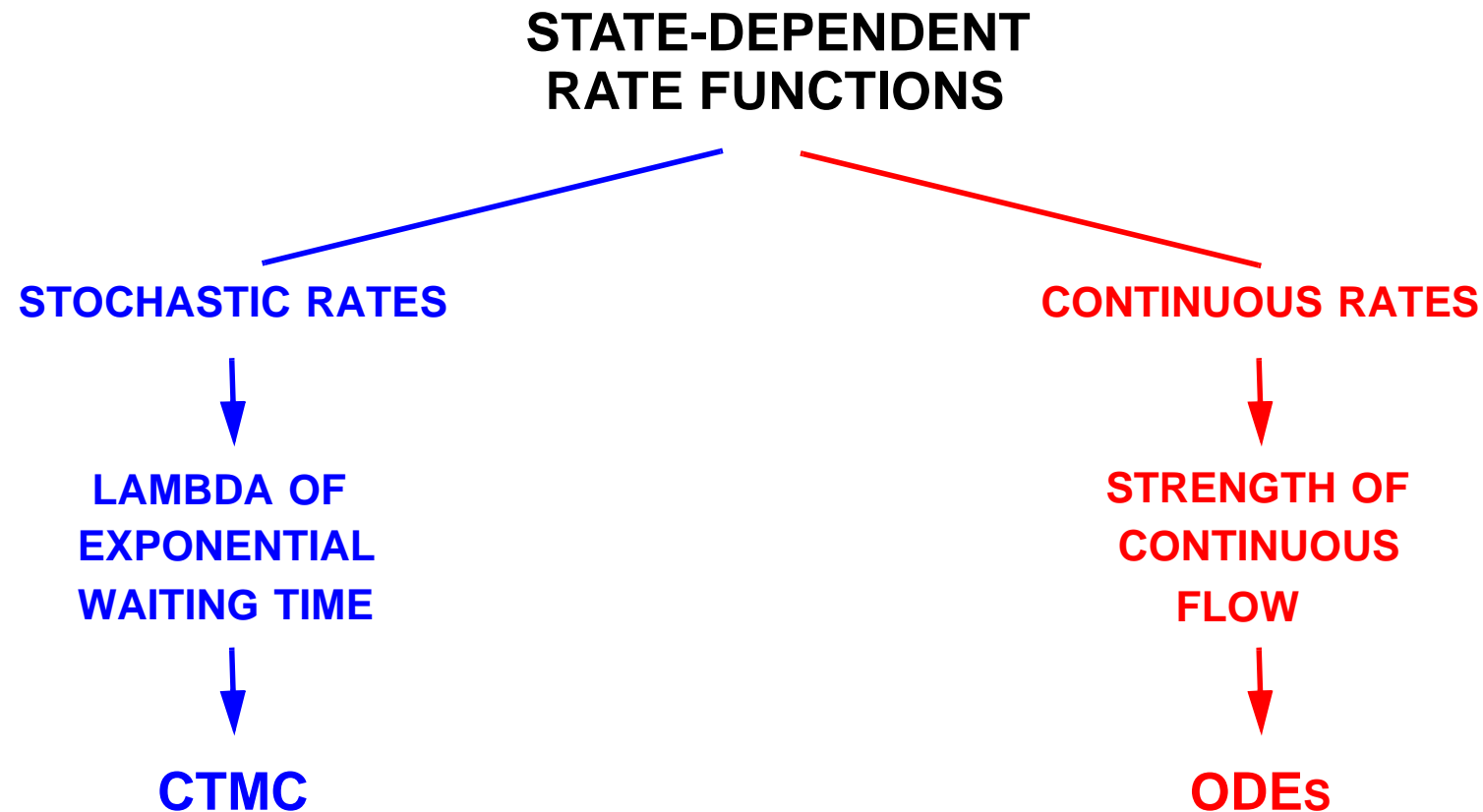
MODELLING = FORMAL KNOWLEDGE REPRESENTATION



MODEL VALIDATION = CONFIDENCE INCREASE





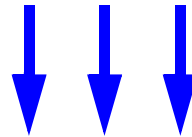


-> supported by, e.g., COPASI, Dizzy, ..., Snoopy



# 4x2

MODELS SHARING STRUCTURE

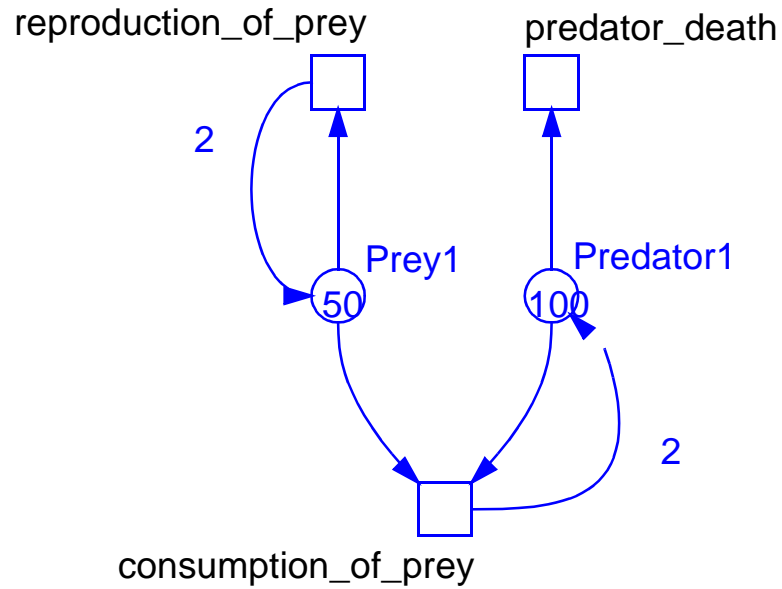


**QUANTITATIVE MODEL = QUALITATIVE MODEL  
+  
RATE FUNCTIONS  
(KINETICS)**

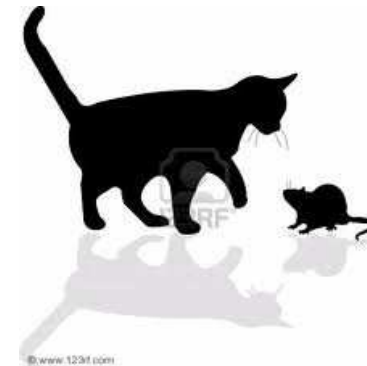
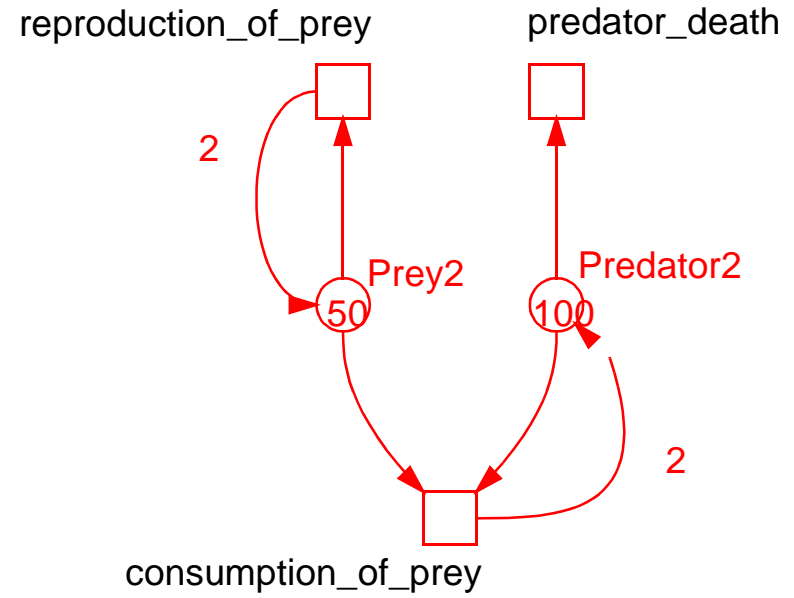
# COLOUR - WHAT FOR ?

# EX: PREY - PREDATOR

sub-system1



sub-system2



❑ **definitions**

*colourset* CS = 1-2;

*var* x : CS;

❑ **better:**

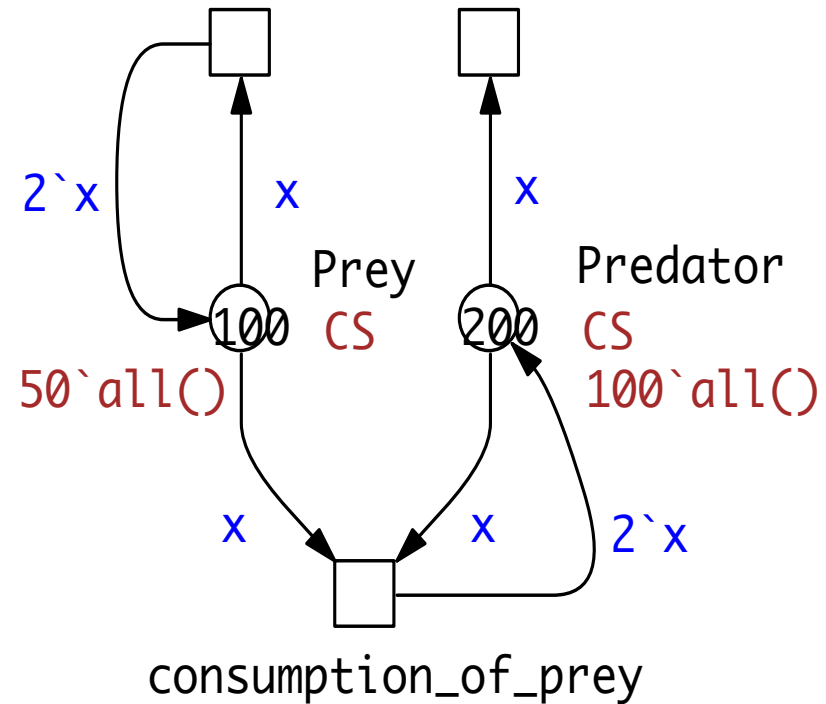
*const* SIZE = 2;

*colourset* CS = 1-SIZE;

*var* x : CS;



reproduction\_of\_prey predator\_death



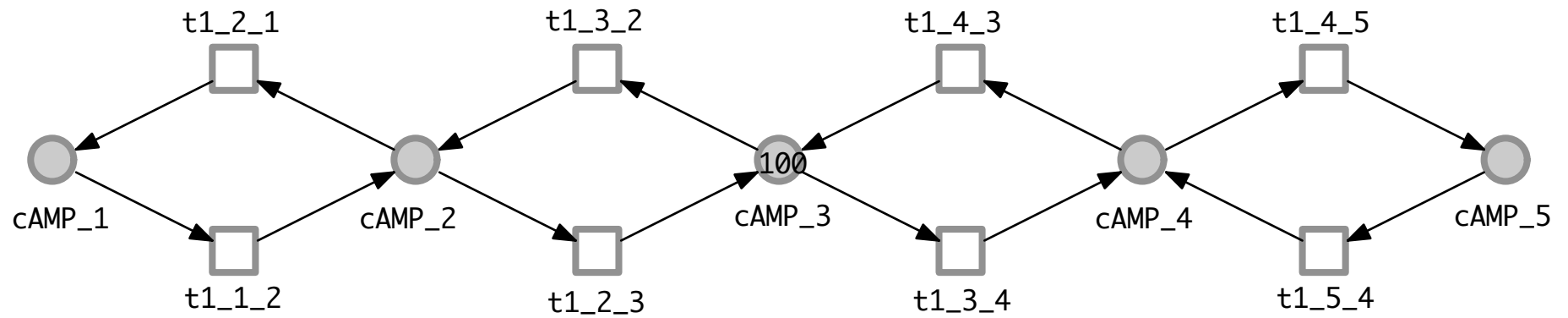
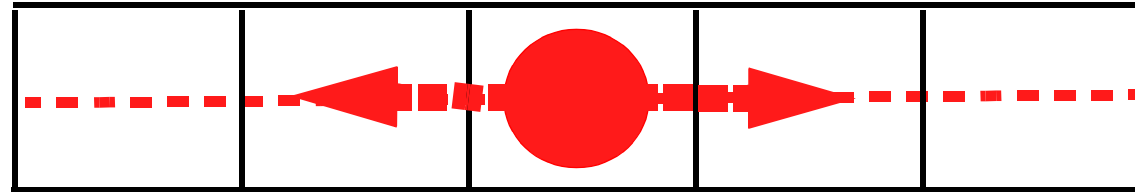
❑ **changing SIZE adapts the model to various scenarios**



*Richmond, 13/09/2011*

# EXAMPLE 1: DIFFUSION IN SPACE

# Ex1: DIFFUSION - 1D



## □ definitions

```
const D1 = 5;           // grid size
```

```
const MIDDLE = D1/2;
```

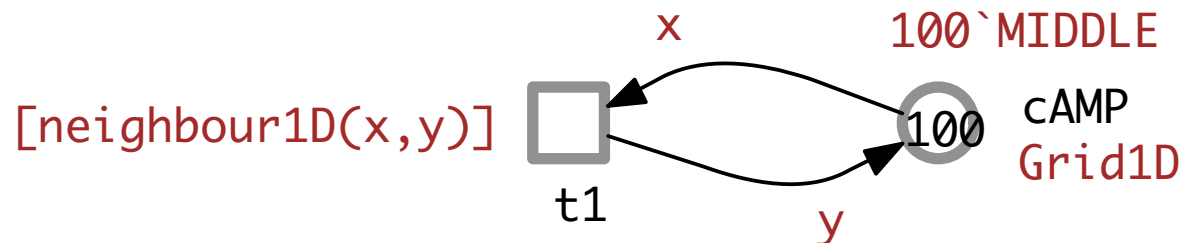
```
colorset CS = 1-D1;    // grid positions
```

```
var x,y : CS;
```

```
function neighbour1D (CS x,a) bool:
```

```
  // a is neighbour of x
```

```
  ( a=x-1 | a=x+1 ) & ( 1<=a ) & ( a<=D1 );
```



## □ movement = changing colour



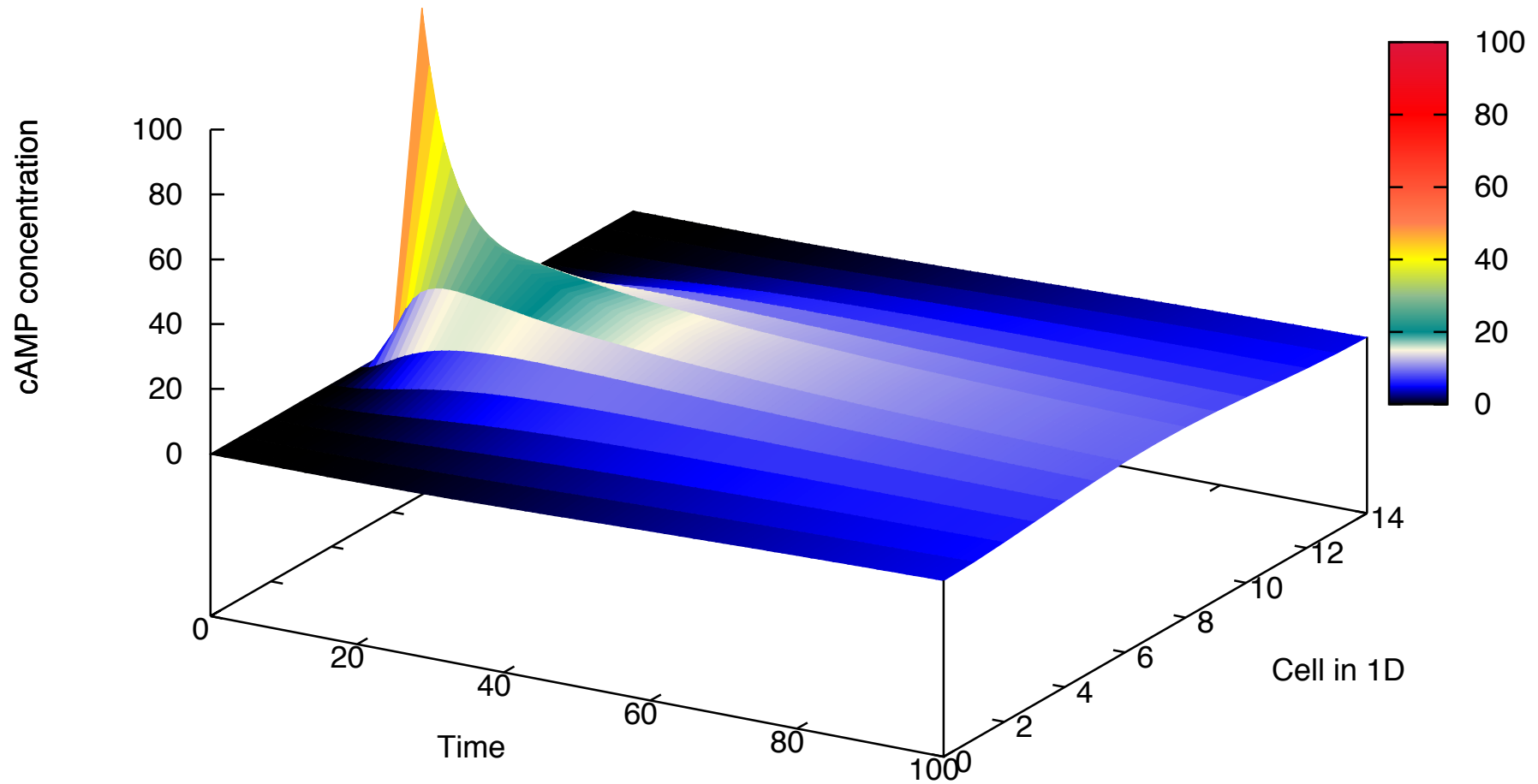
$$\frac{dc_1}{dt} = k \cdot c_2 - k \cdot c_1$$

$$\frac{dc_2}{dt} = k \cdot c_1 + k \cdot c_3 - 2 \cdot k \cdot c_2$$

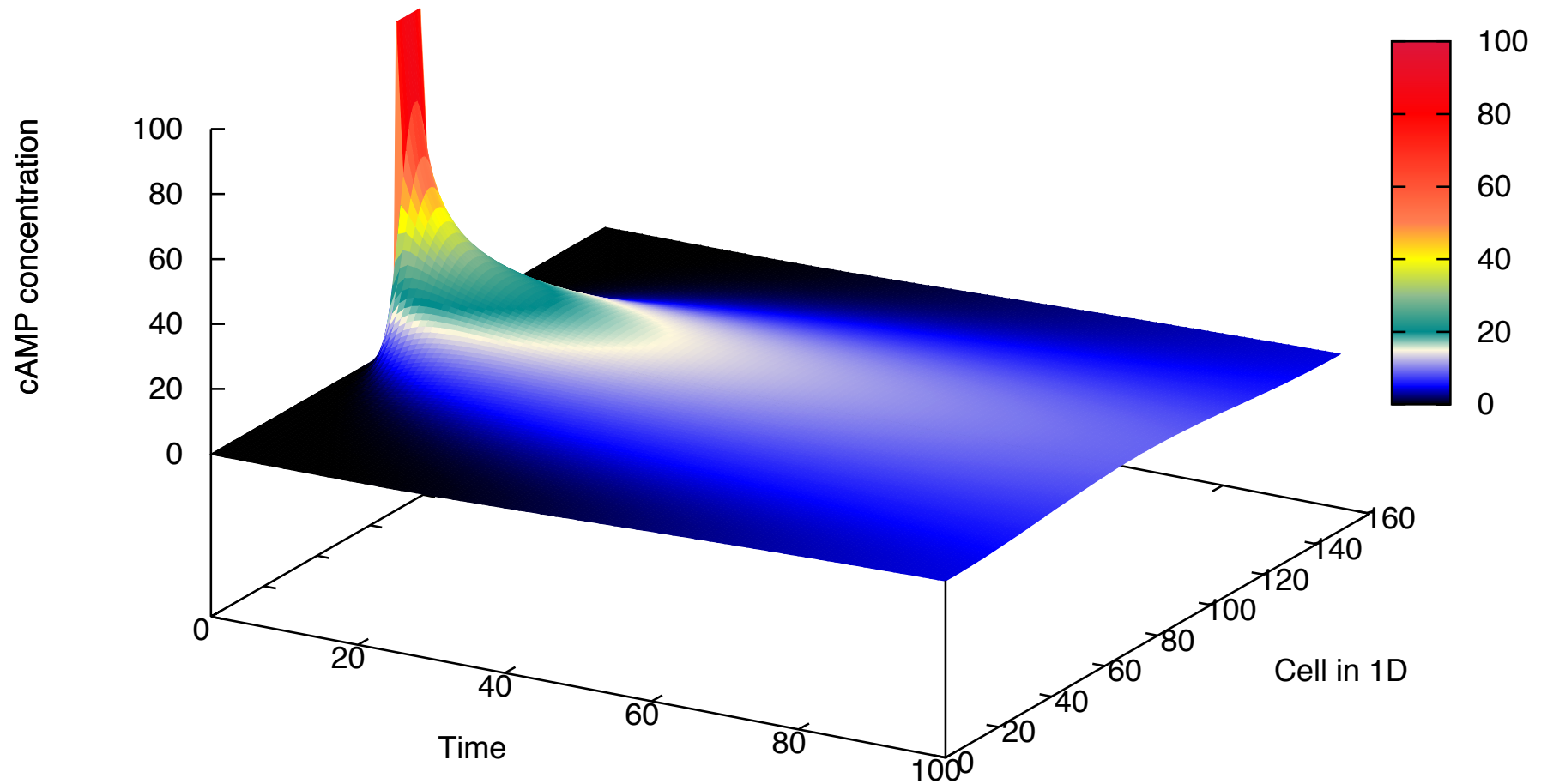
$$\frac{dc_3}{dt} = k \cdot c_2 + k \cdot c_4 - 2 \cdot k \cdot c_3$$

$$\frac{dc_4}{dt} = k \cdot c_3 + k \cdot c_5 - 2 \cdot k \cdot c_4$$

$$\frac{dc_5}{dt} = k \cdot c_4 - k \cdot c_5$$

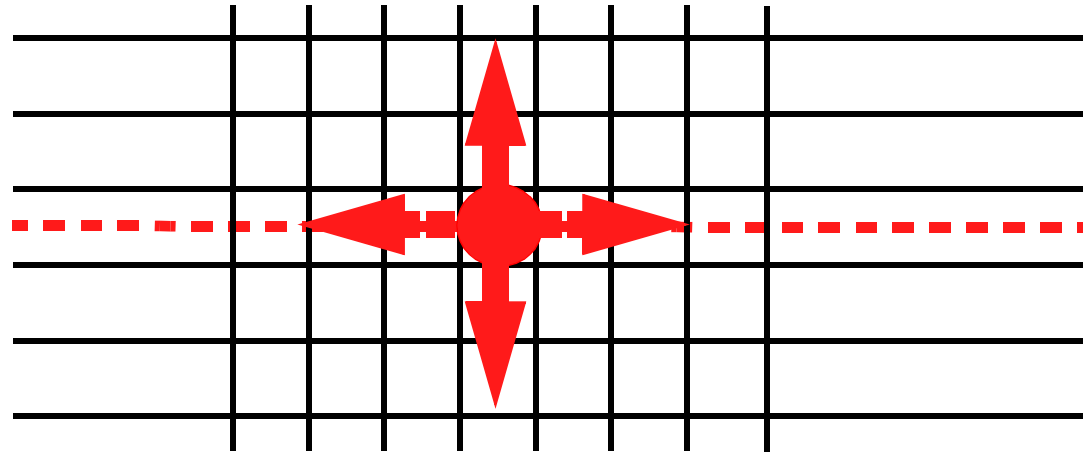


## 15 GRID POSITIONS

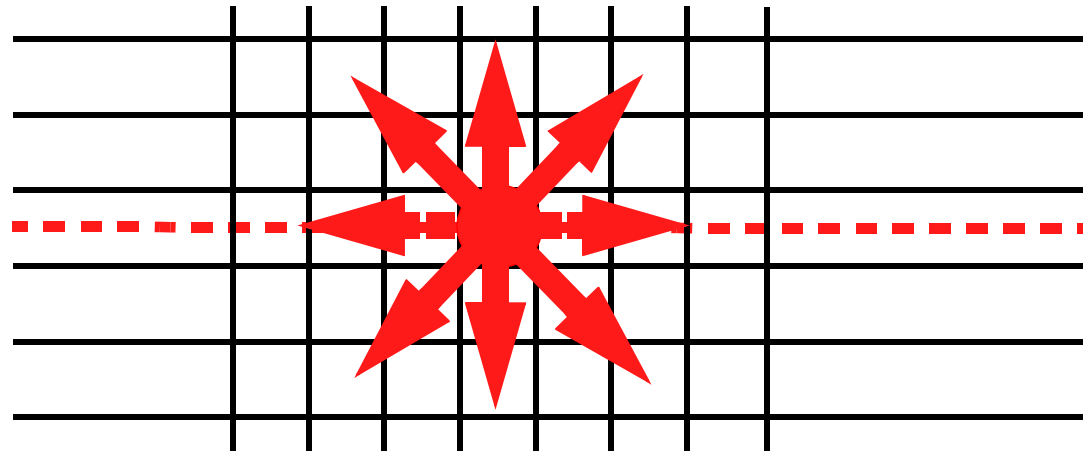


## 150 GRID POSITIONS, SCALING OF INITIAL MARKING AND RATES

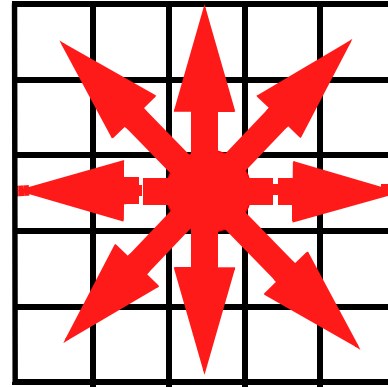
## □ SCHEME



## □ SCHEME



## ❑ SCHEME



## ❑ definitions

```
const D1 = 5;           // grid size first dimension
const D2 = D1;         // grid size second dimension
const MIDDLE = D1/2;

colorset CD1 = 1-D1;   // row index
colorset CD2 = 1-D2;   // column index
colorset Grid2D = CD1 x CD2; // 2D grid

var x, a : CD1;
var y, b : CD2;
```

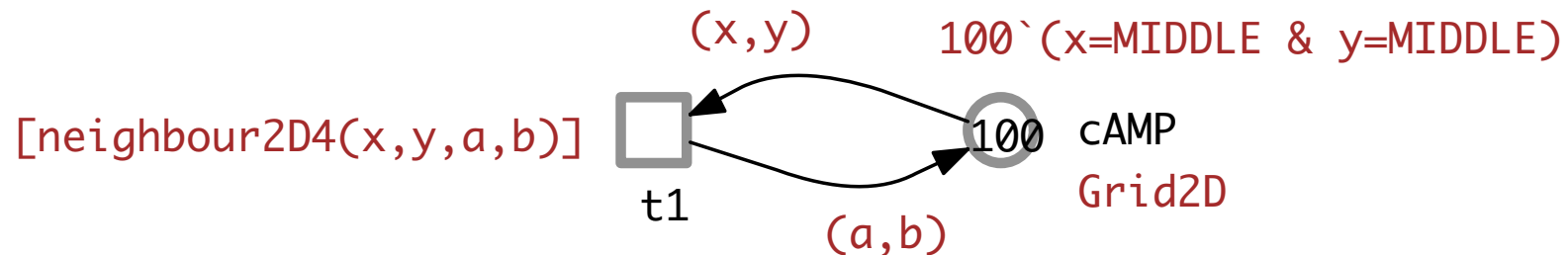
## □ four neighbours

**function** neighbour2D4 (CD1 x, CD2 y, CD1 a, CD2 b) **bool**:

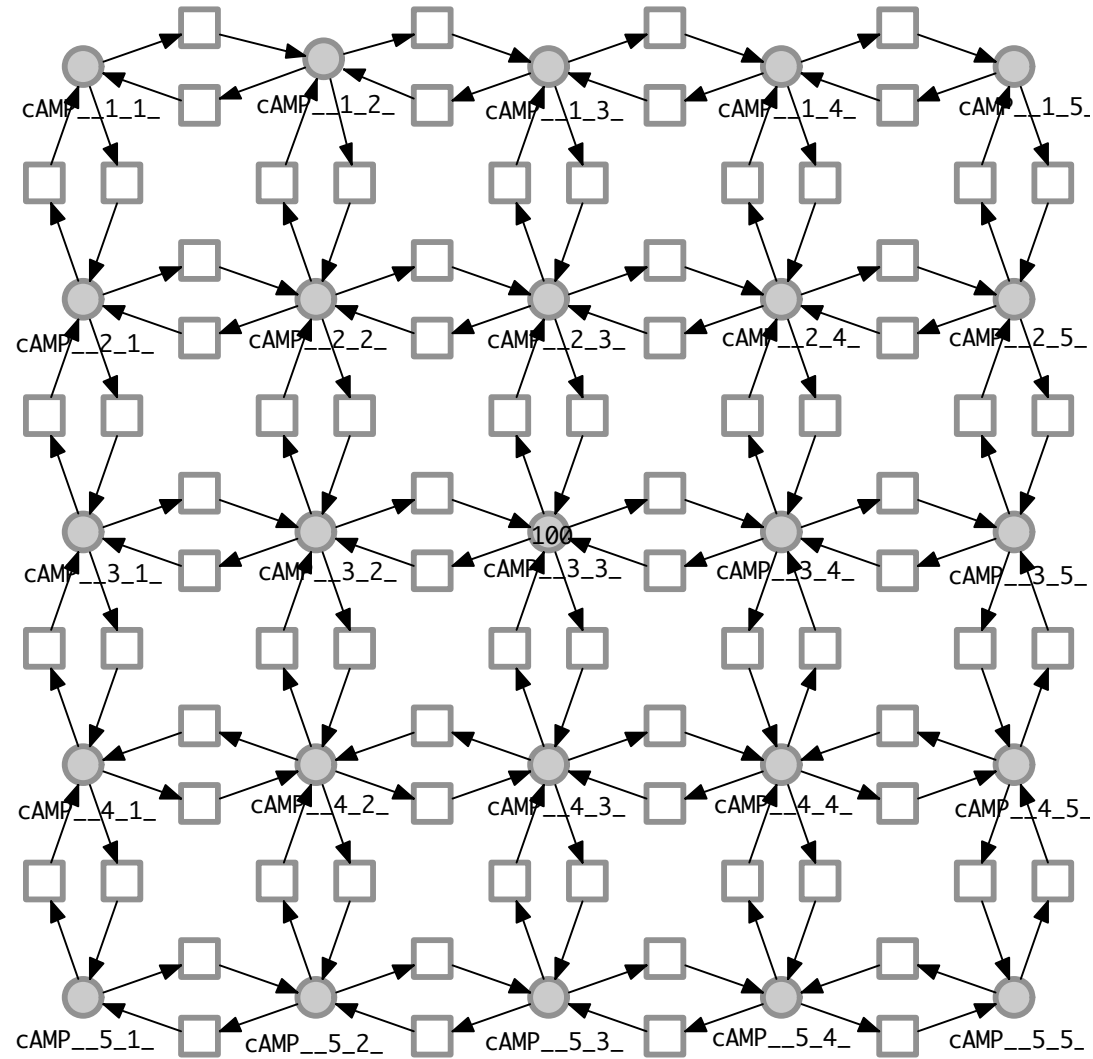
// (a,b) is one of the up to four neighbours of (x,y)

(a=x & b=y-1) | (a=x & b=y+1)

| (b=y & a=x-1) | (b=y & a=x+1);



# Ex1: DIFFUSION - 2D4 NEIGHBOURHOOD





## □ eight neighbours

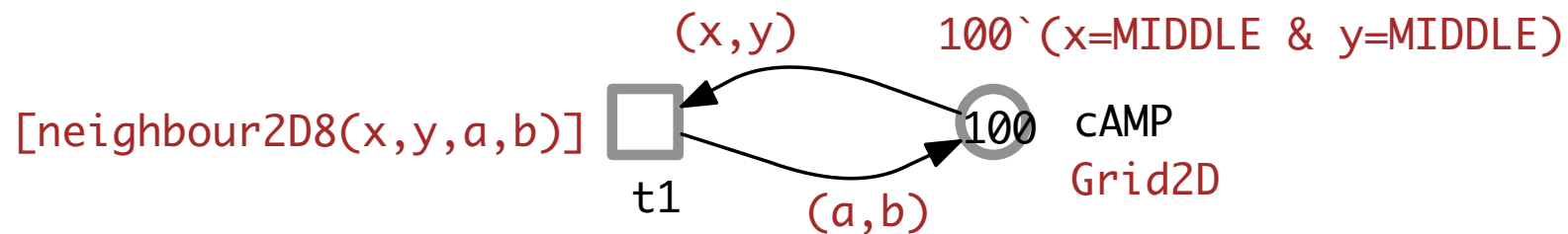
**function** neighbour2D8 (CD1 x, CD2 y, CD1 a, CD2 b) **bool**:

// (a,b) is one of the up to eight neighbours of (x,y)

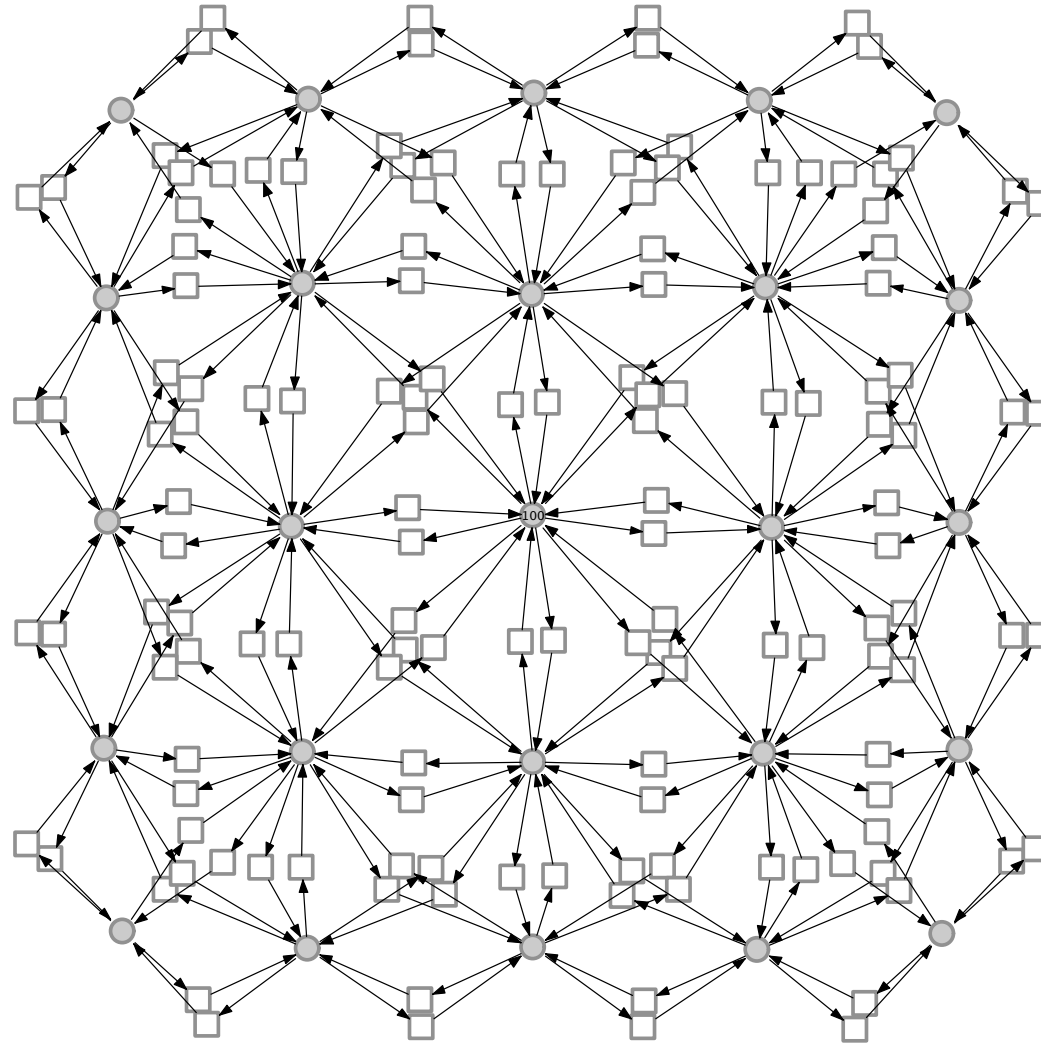
(a=x-1 | a=x | a=x+1) & (b = y-1 | b=y | b=y+1)

& (!(a=x & b=y))

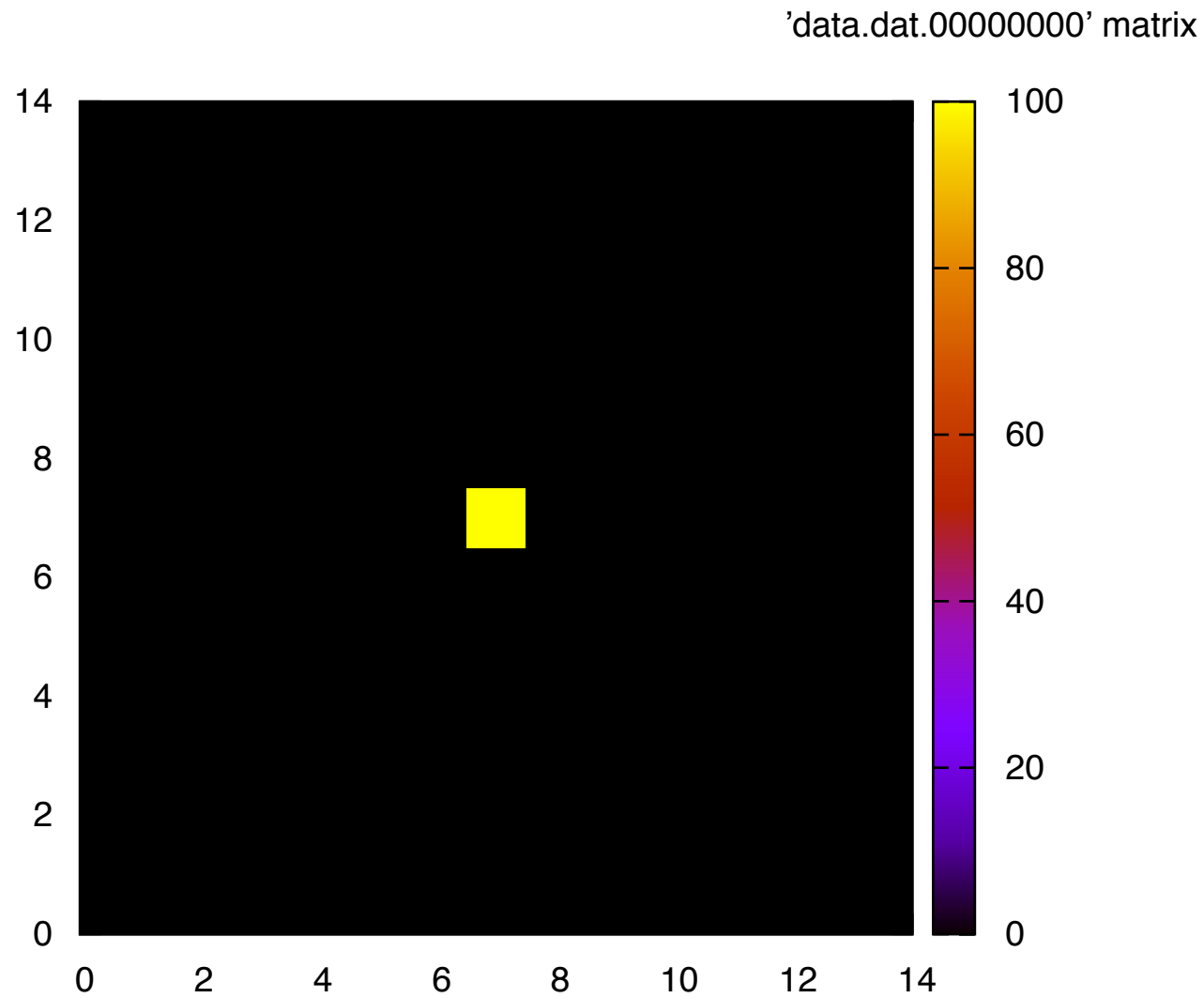
& (1<=a & a<=D1) & (1<=b & b<=D2);

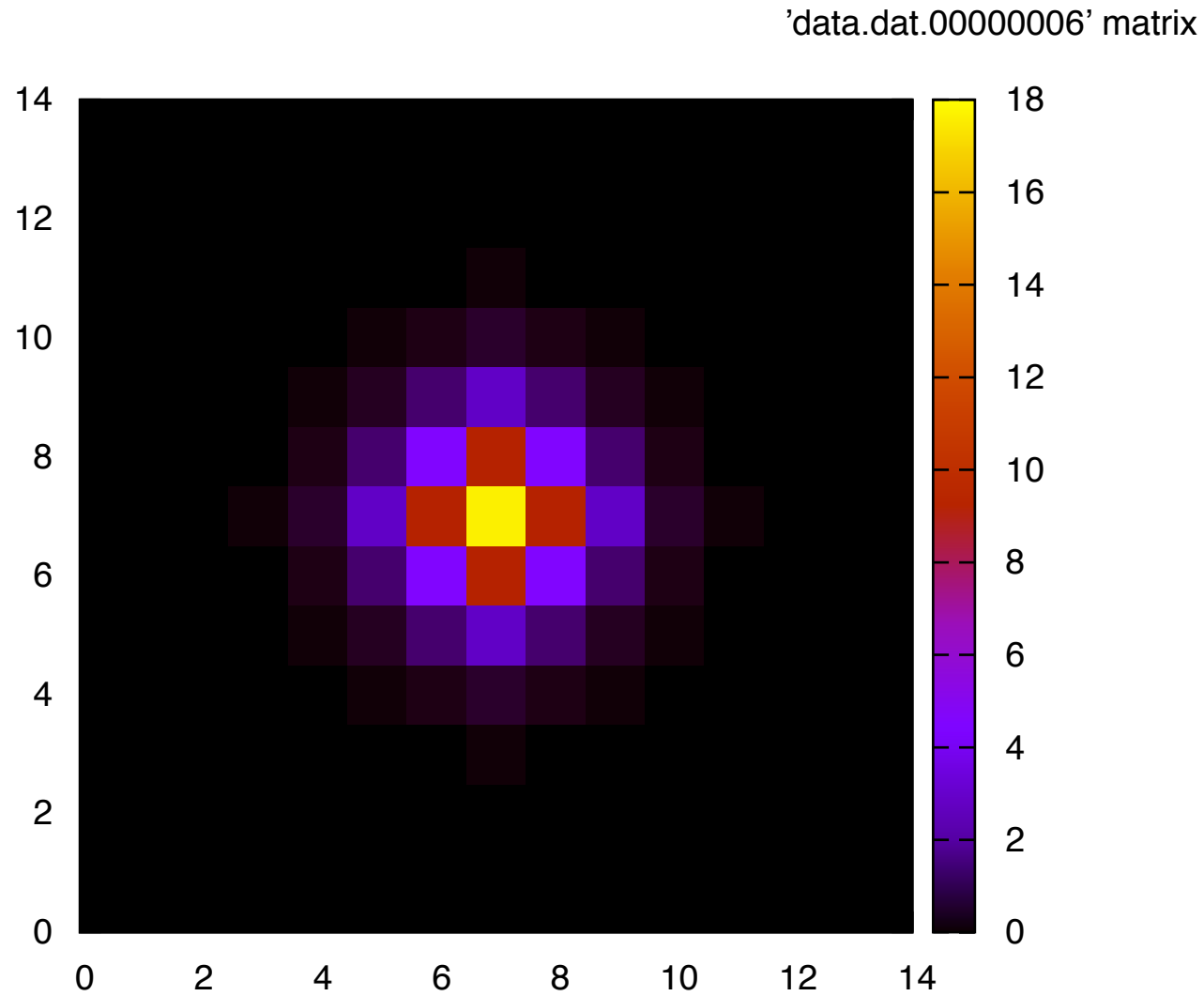


# Ex1: DIFFUSION - 2D8 NEIGHBOURHOOD

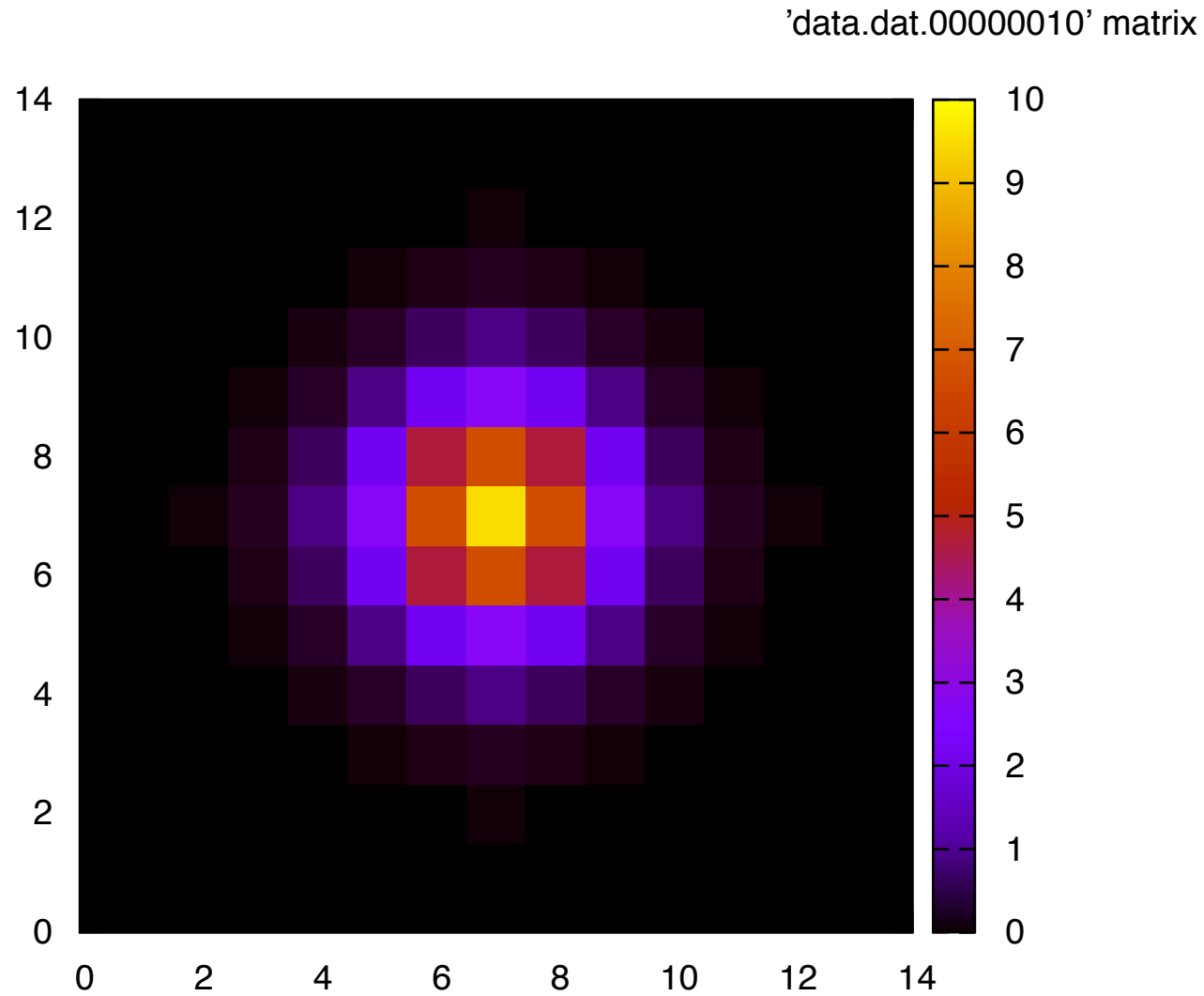


# Ex1: DIFFUSION - 2D4 NEIGHBOURHOOD, 15X15

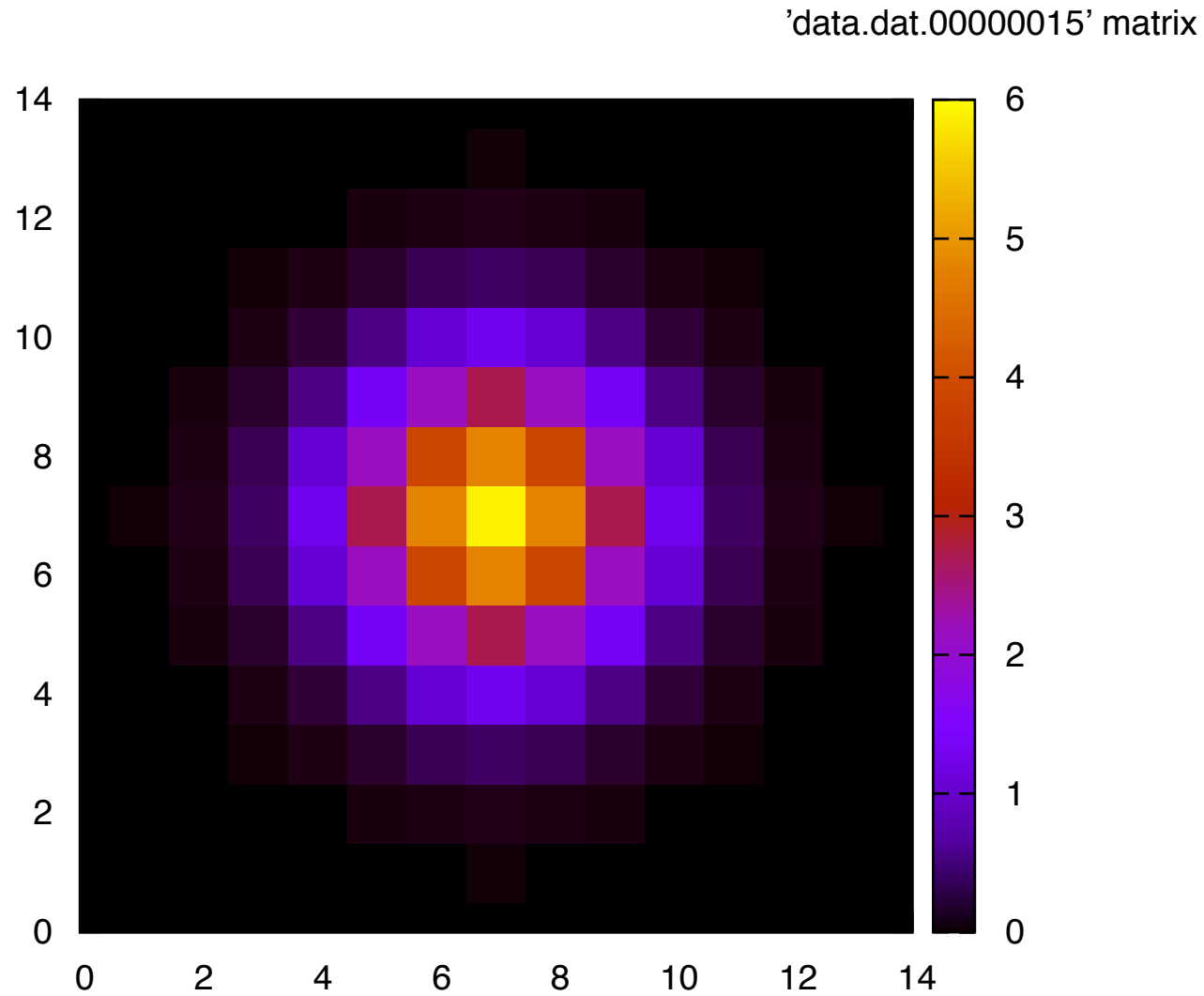




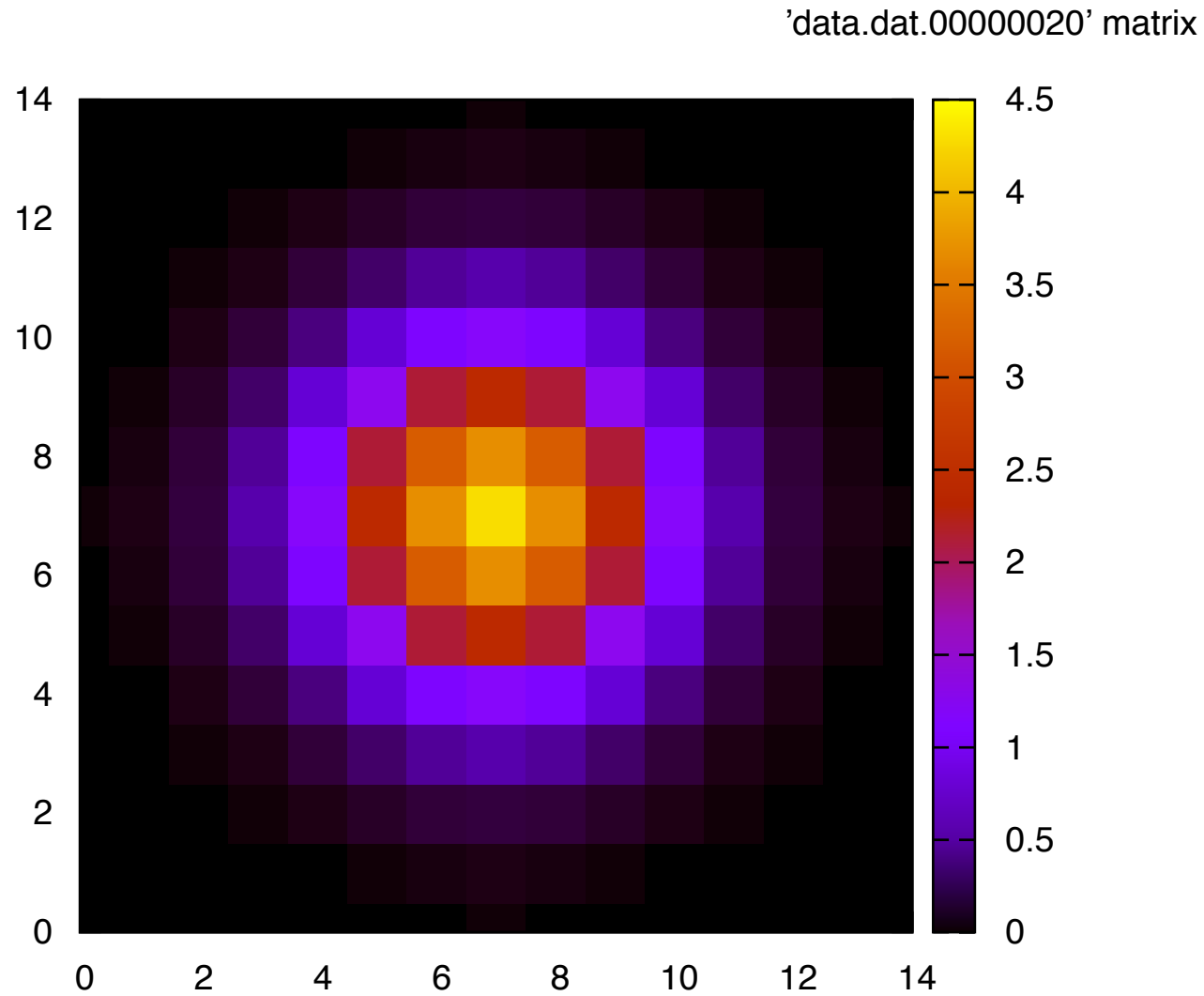
# Ex1: DIFFUSION - 2D4 NEIGHBOURHOOD, 15X15



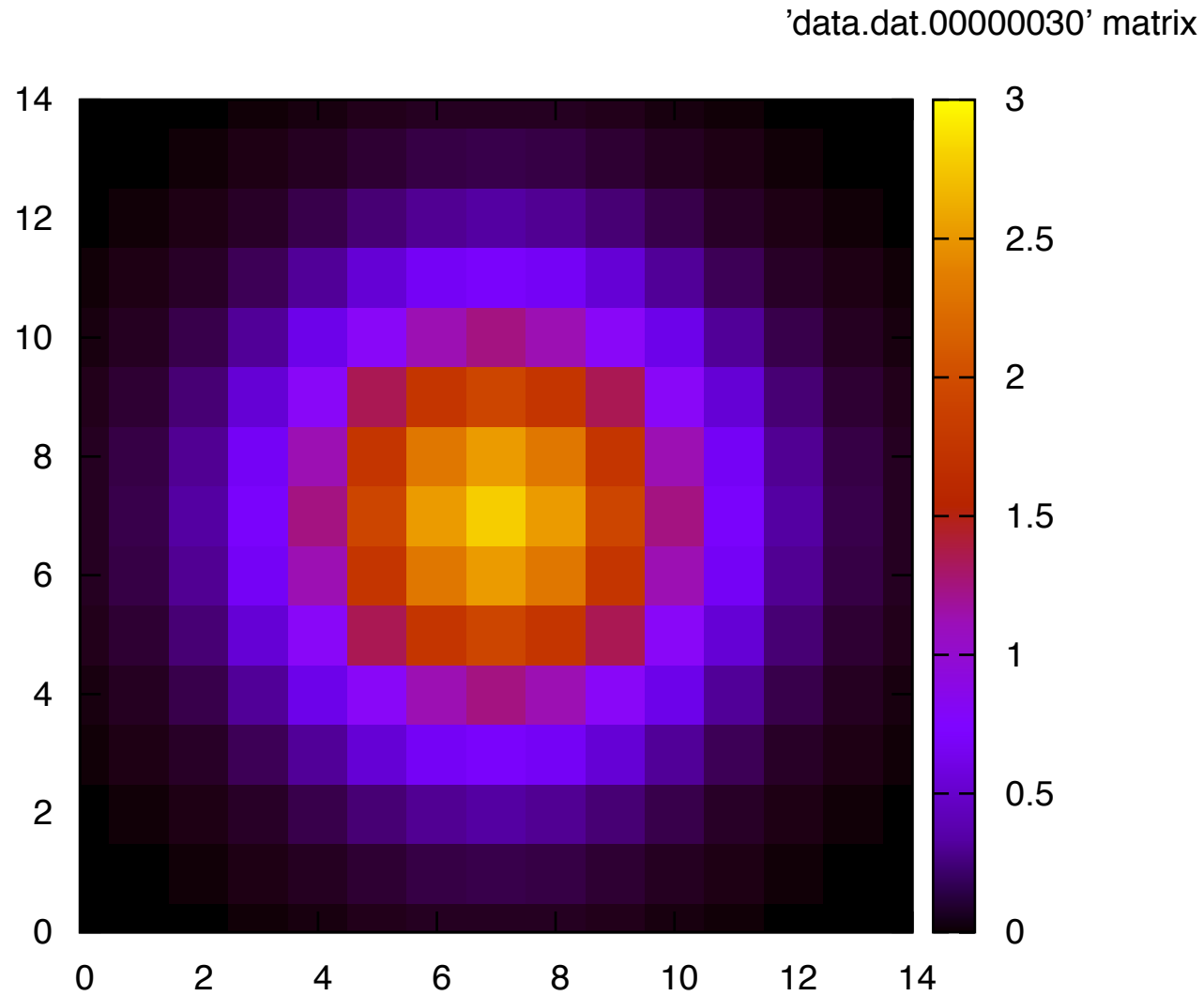
# Ex1: DIFFUSION - 2D4 NEIGHBOURHOOD, 15X15



# Ex1: DIFFUSION - 2D4 NEIGHBOURHOOD, 15X15

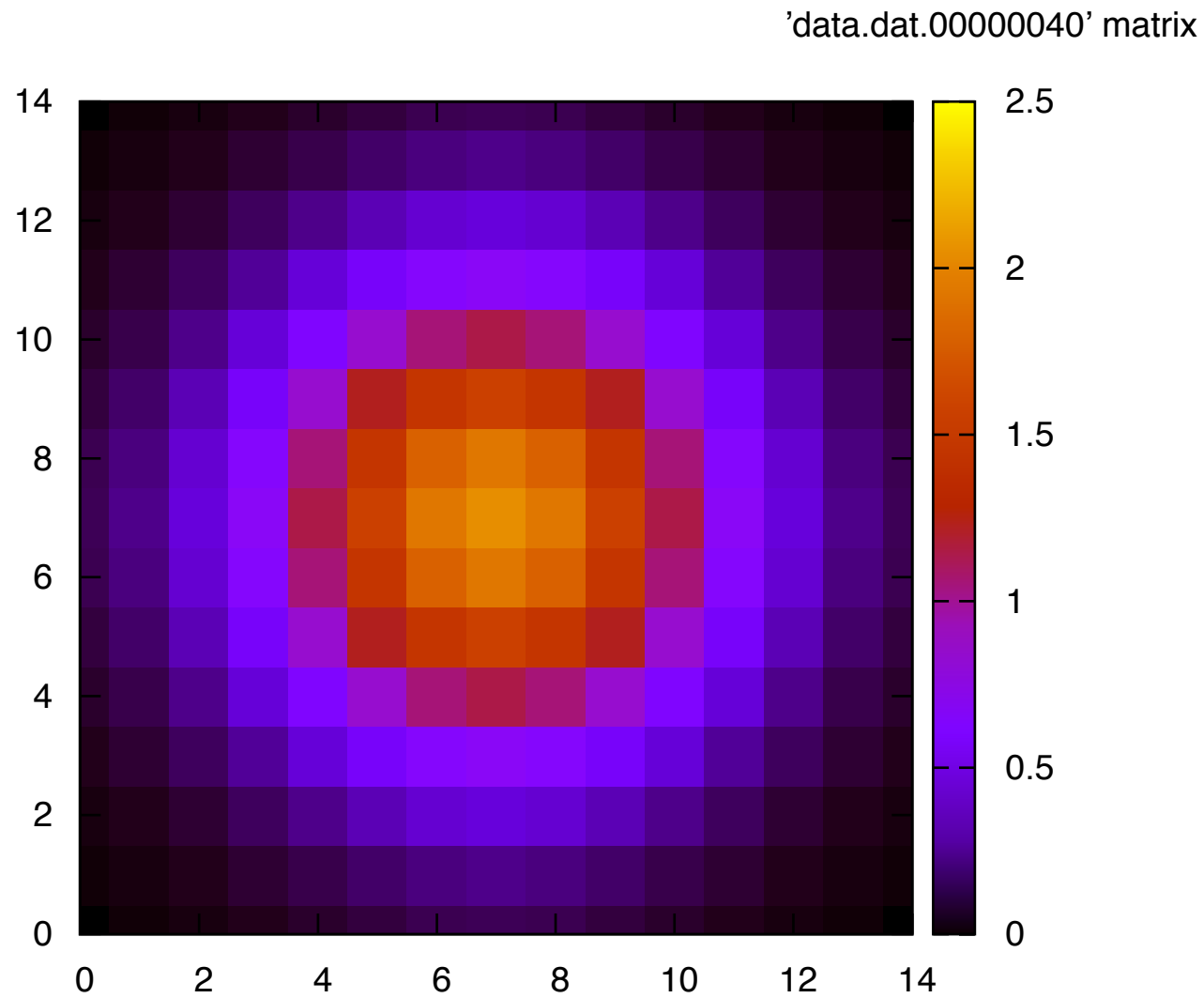


# Ex1: DIFFUSION - 2D4 NEIGHBOURHOOD, 15X15

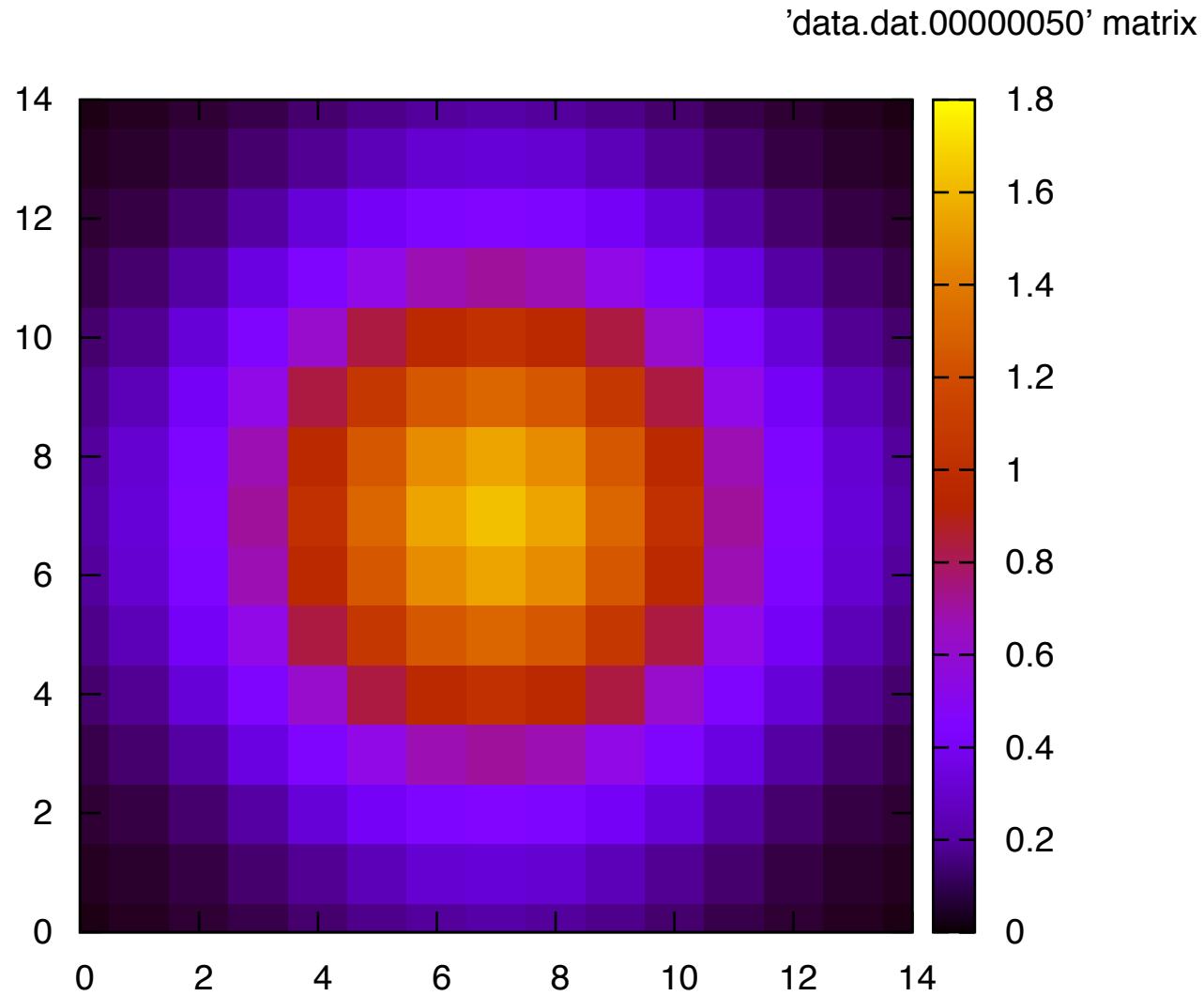


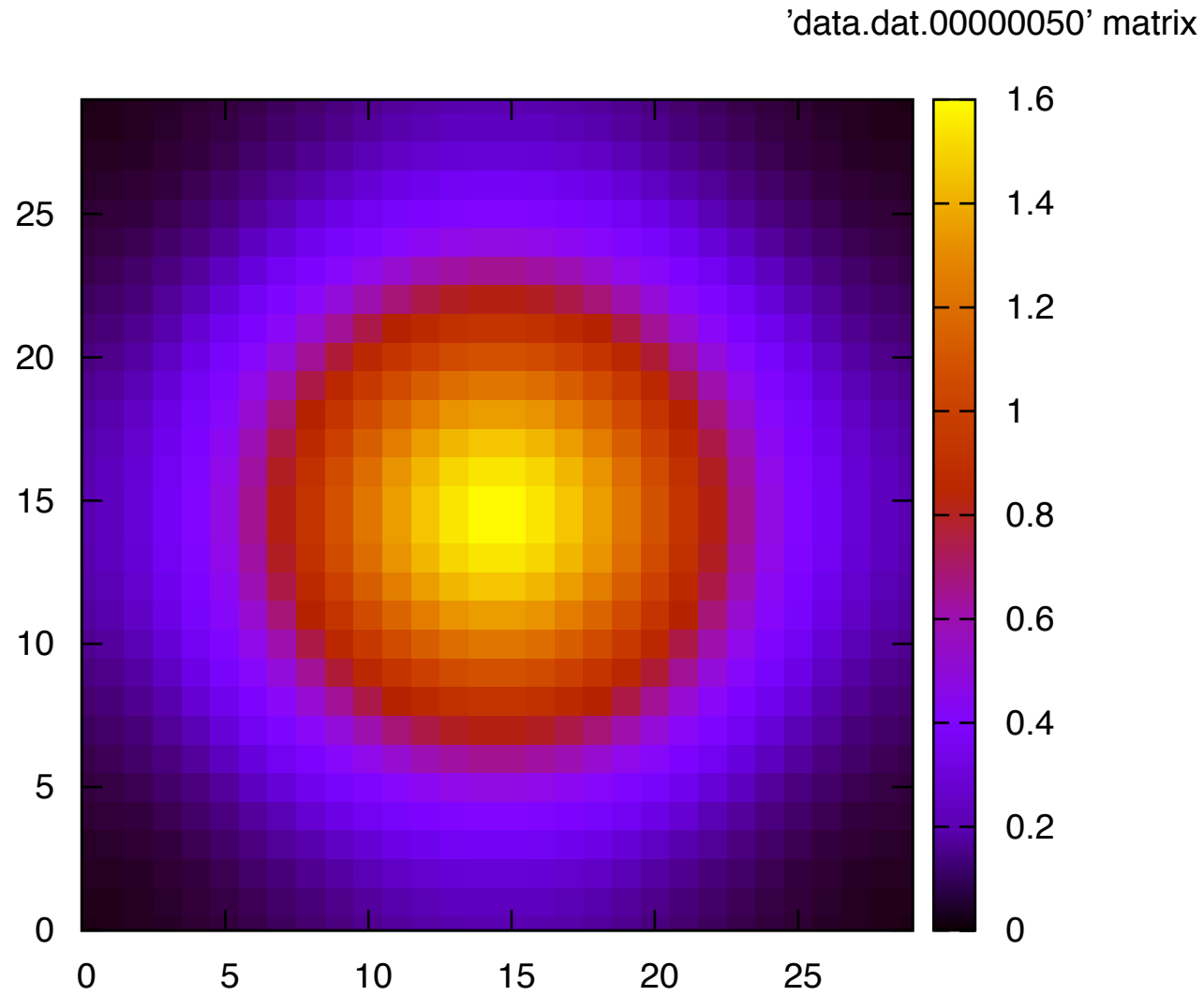


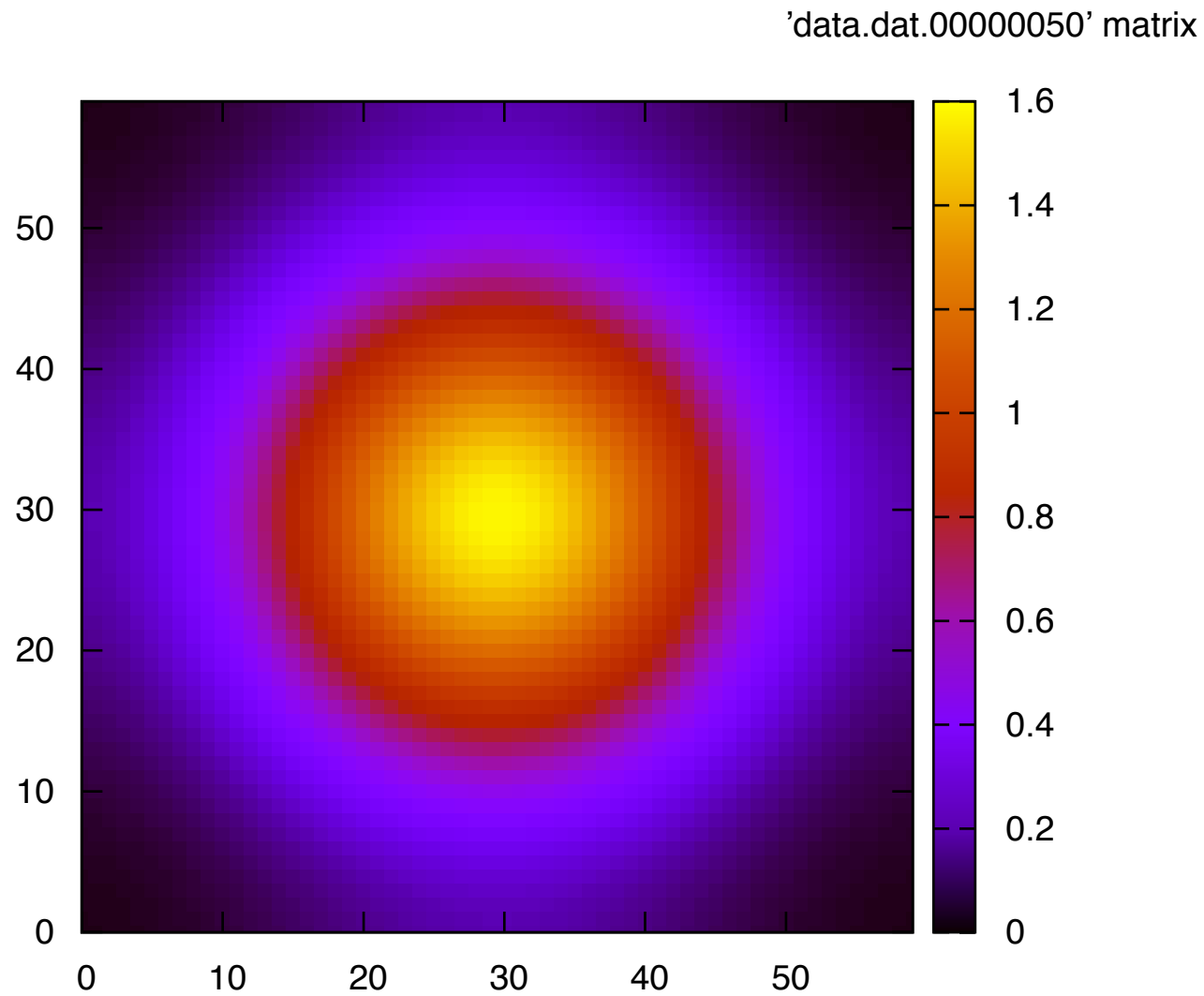
# Ex1: DIFFUSION - 2D4 NEIGHBOURHOOD, 15X15

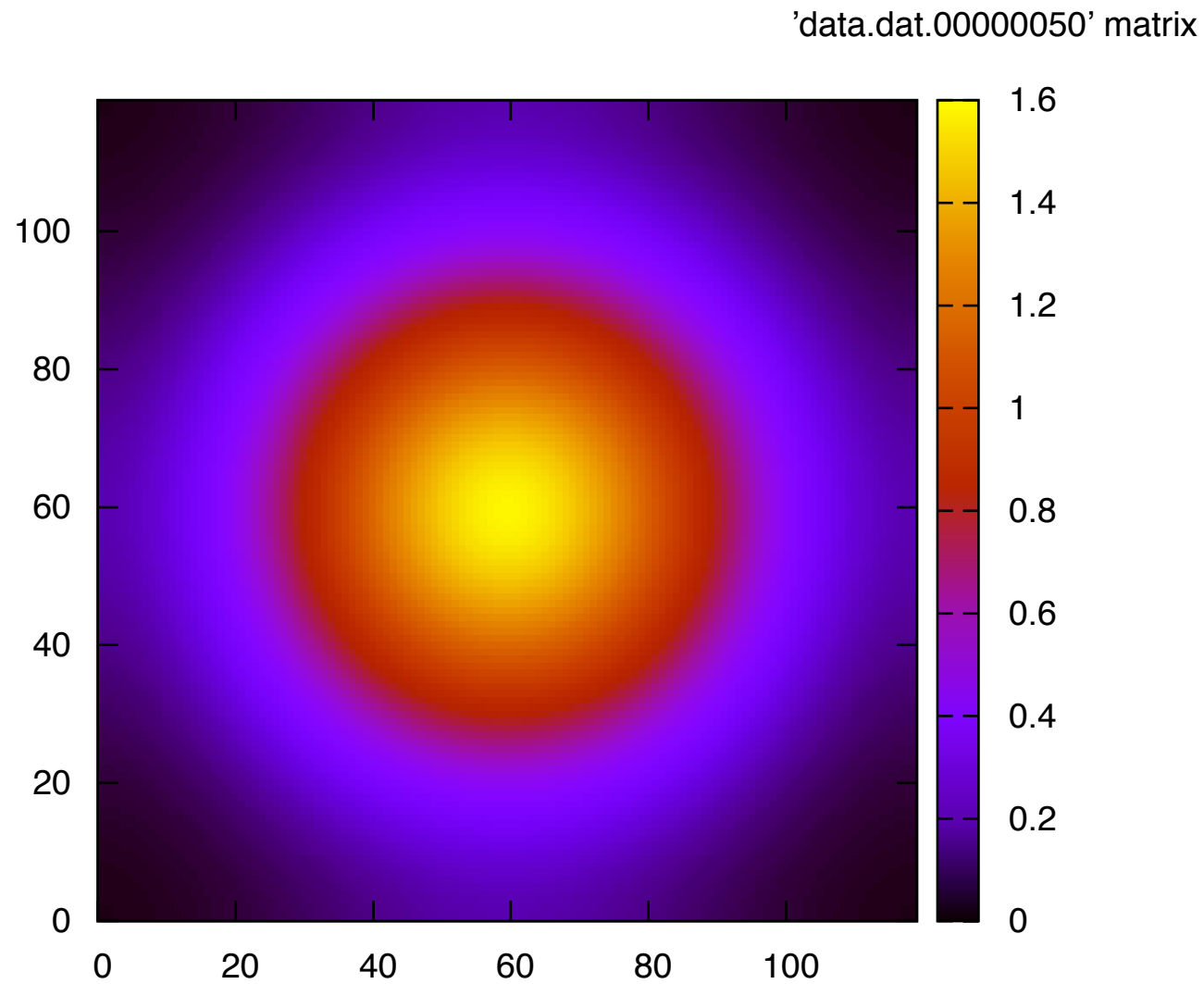


# Ex1: DIFFUSION - 2D4 NEIGHBOURHOOD, 15X15









# EXAMPLE 2: TURING PATTERNS

### “How the Leopard Got Its Spots”

#### □ **morphogenesis**

- > *developmental pattern formation in bio systems*
- > *the process that controls the organized spatial distribution of cells*
- > *tiger stripes, leopard spots, the precisely spaced rows of alligator teeth, etc.*

#### □ **Turing's theory of biological pattern formation, 1952**

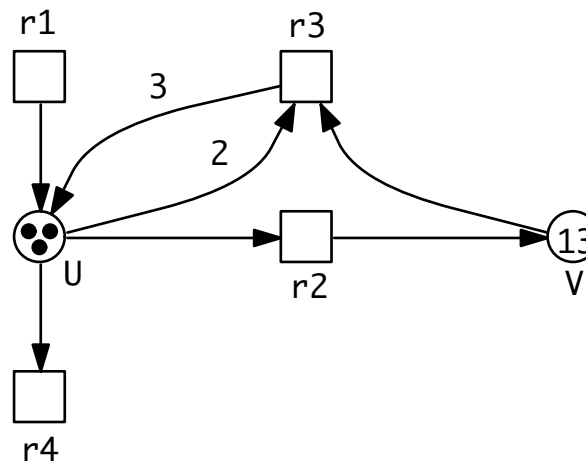
- > *patterns form as result of*  
*the interactions between two chemicals*  
*that spread throughout a system at different rates*

#### □ **highly simplified and idealised take on biological patterning**

#### □ **mathematical challenge**

- > *For which parameters do stable/oscillating Turing patterns exist ?*
- > *analysis of stability, multistability, bifurcation of non-linear PDE*

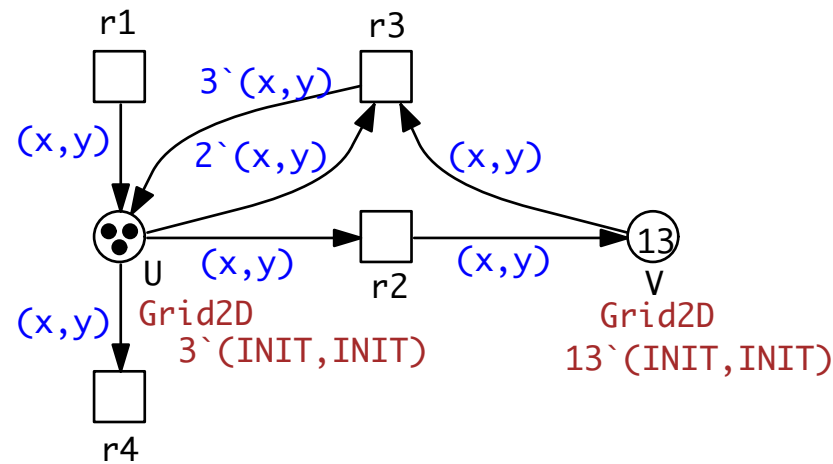
r1:  $\rightarrow U$   
r2:  $U \rightarrow V$   
r3:  $2U + V \rightarrow 3U$   
r4:  $U \rightarrow$



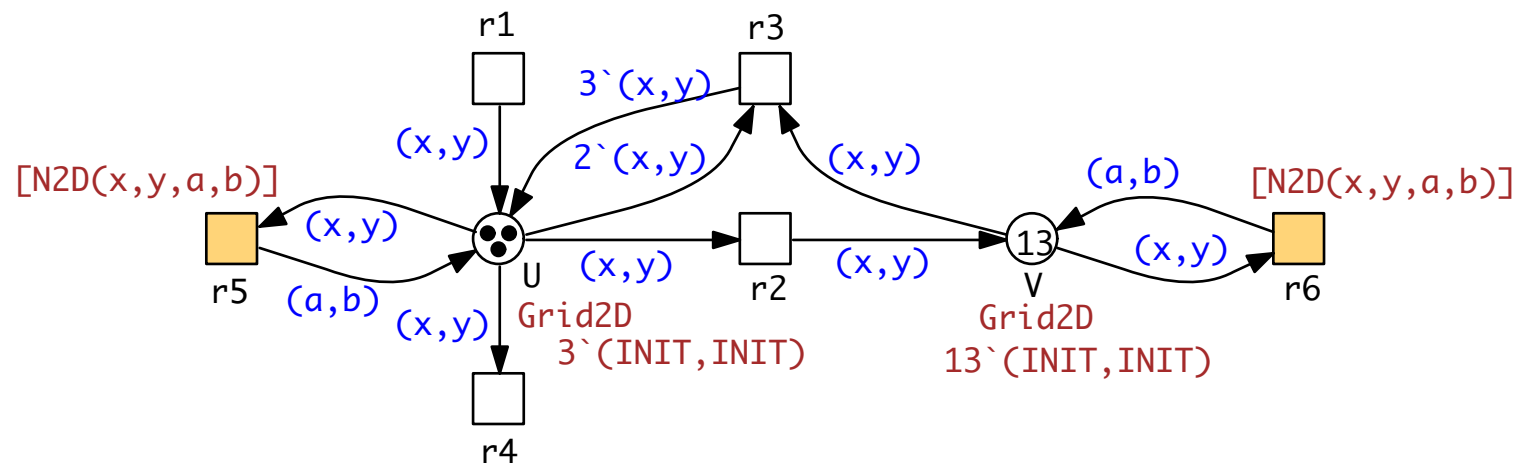
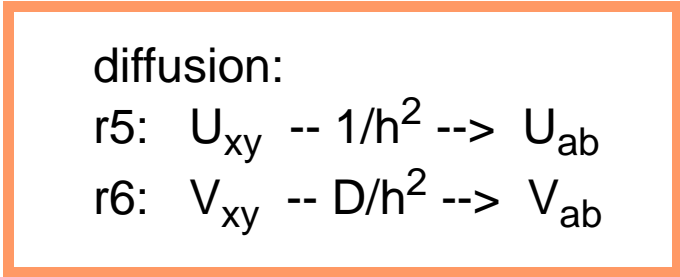
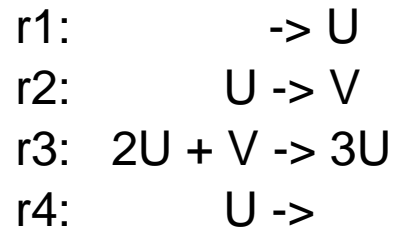


- r1:  $\rightarrow U$
- r2:  $U \rightarrow V$
- r3:  $2U + V \rightarrow 3U$
- r4:  $U \rightarrow$

## adding SPACE



# EX2: TURING PATTERNS



*r1 - r4 follow mass action kinetics with rate constants:*  
*r1: a, r2: b, r3: 1, r4: 1;*

### ❑ reactions

-> *version of Brusselator model, <http://en.wikipedia.org/wiki/Brusselator>*

### ❑ parameters

-> *Pena, Perez-Garcia: Stability of Turing patterns in the Brusselator model, Physical Review 2001*

->  *$a = 4.5$ ,  $b = 0.04 \dots 0.98$ ,  $D = 128$ ,  $h = 0.8$*

### ❑ unfolding

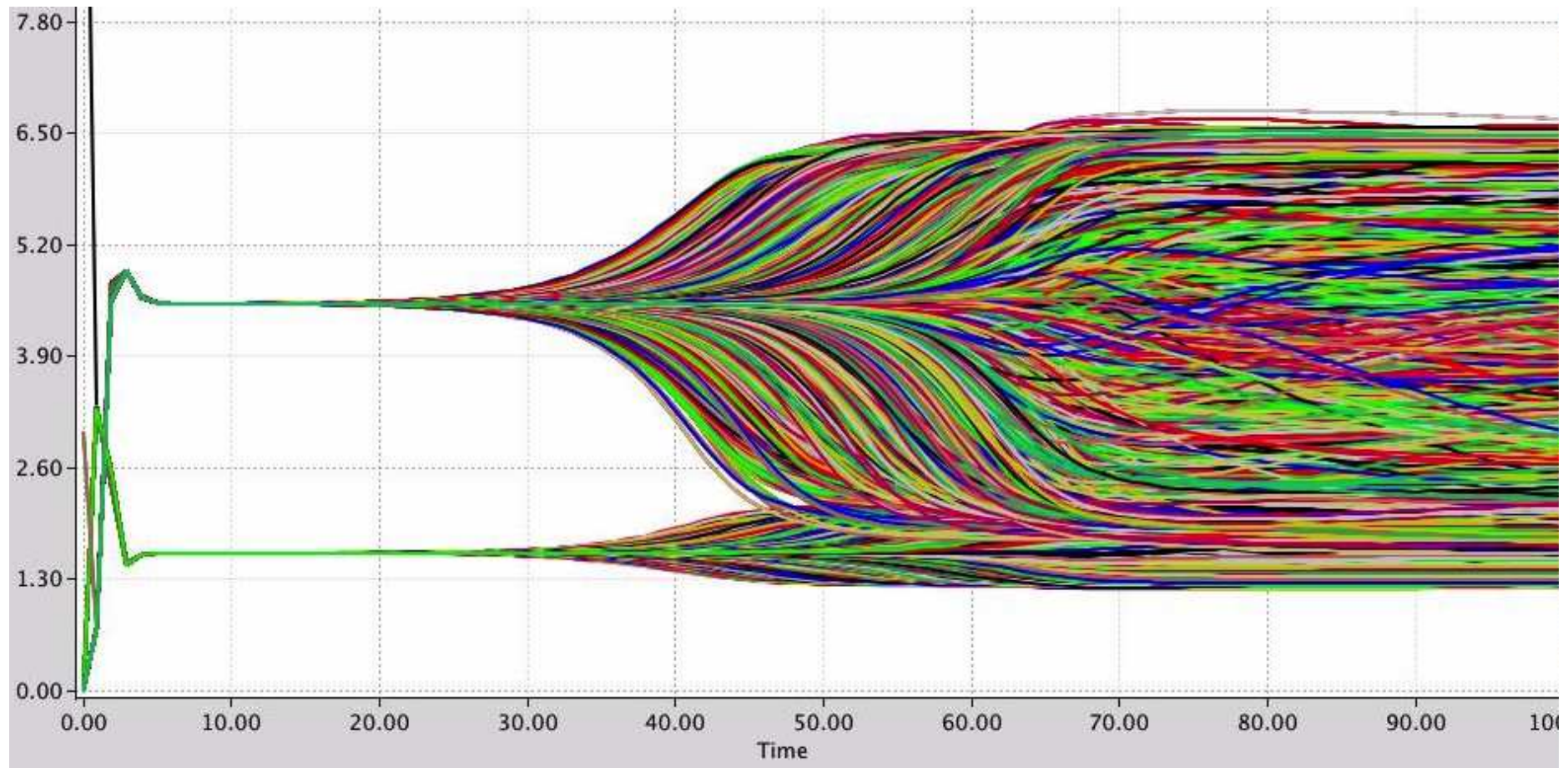
-> *runtime (constraint solver, 4 threads): 128 sec*

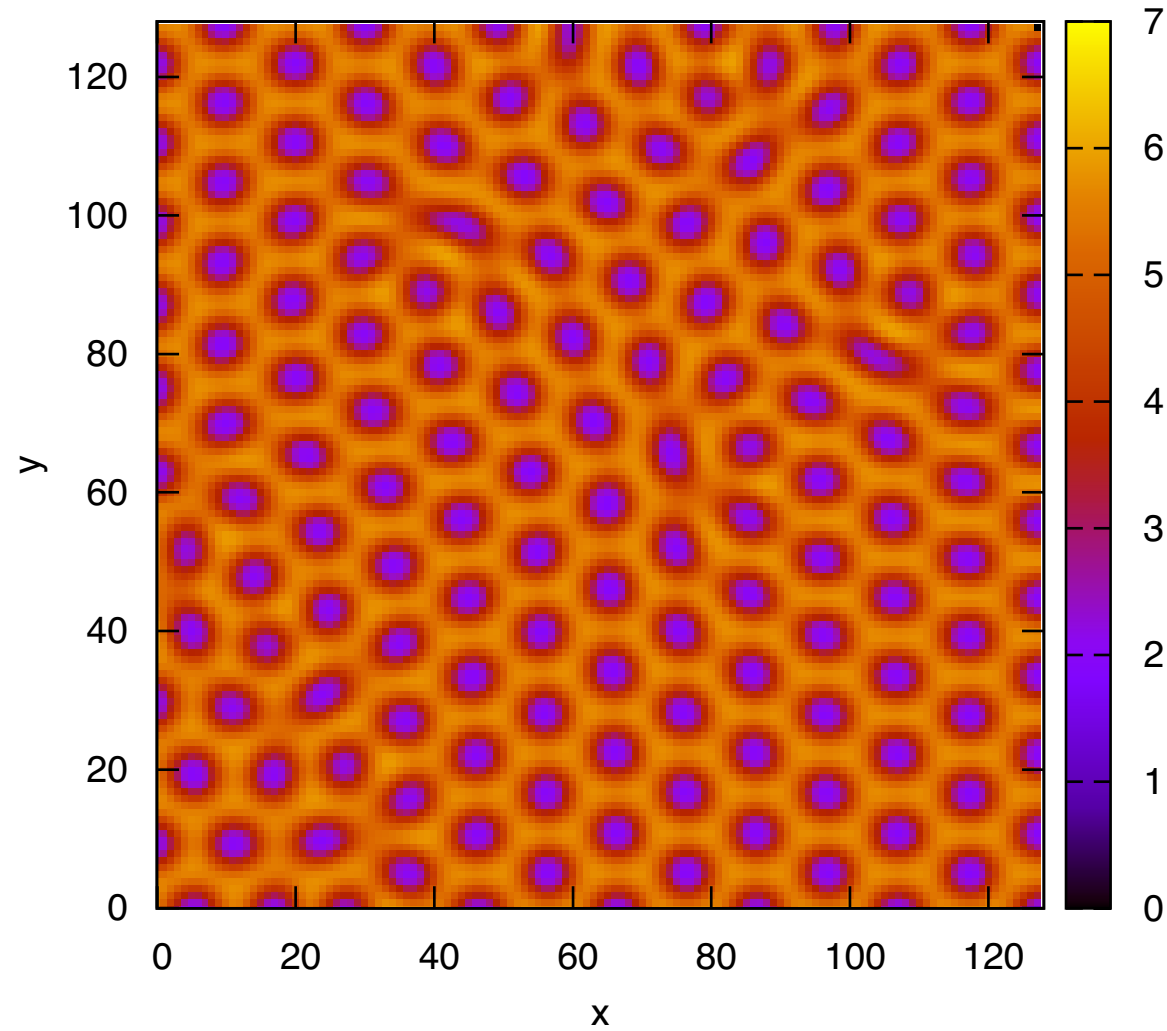
-> *places: 32,768, transitions: 324,616*

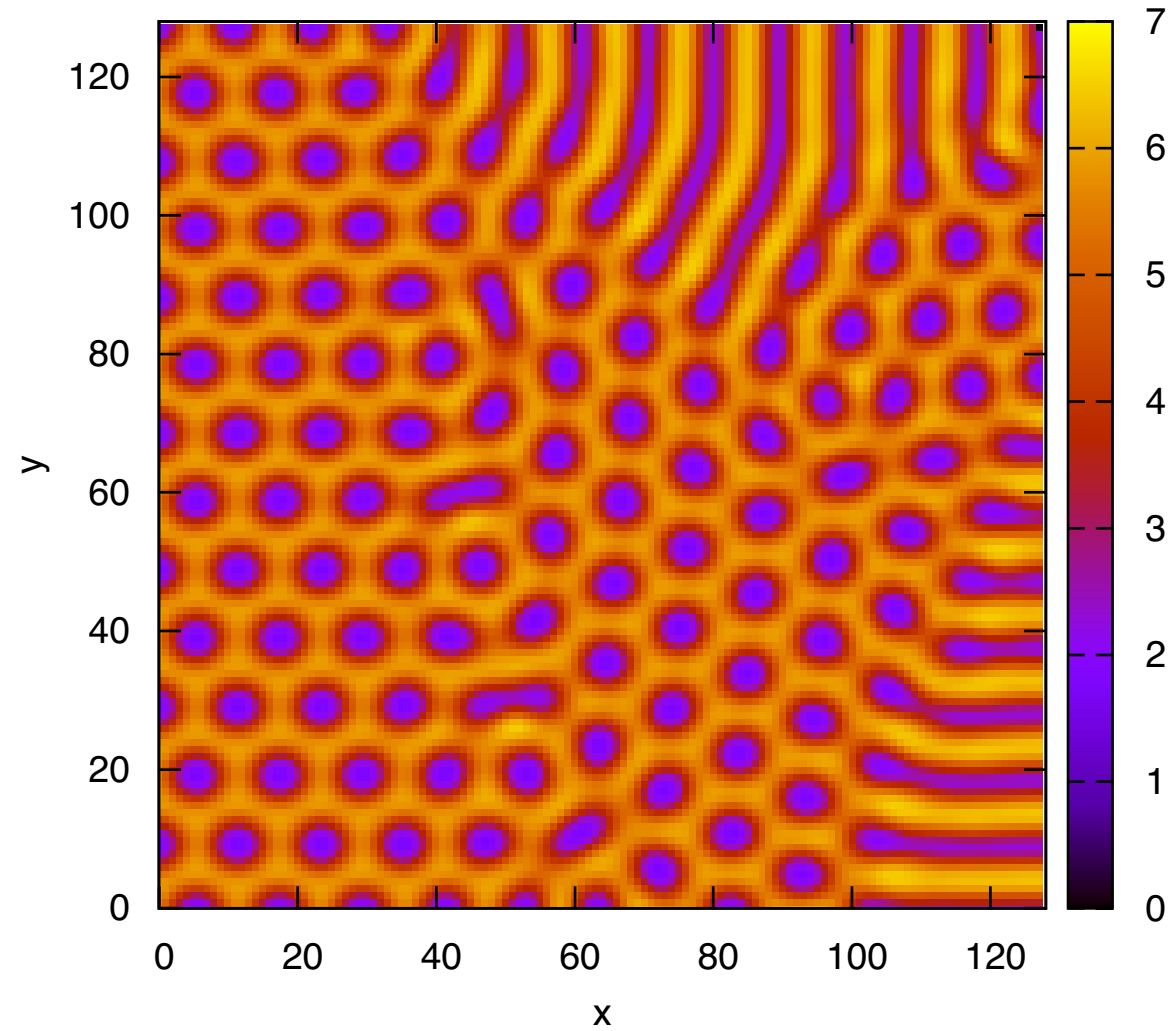
### ❑ continues simulation

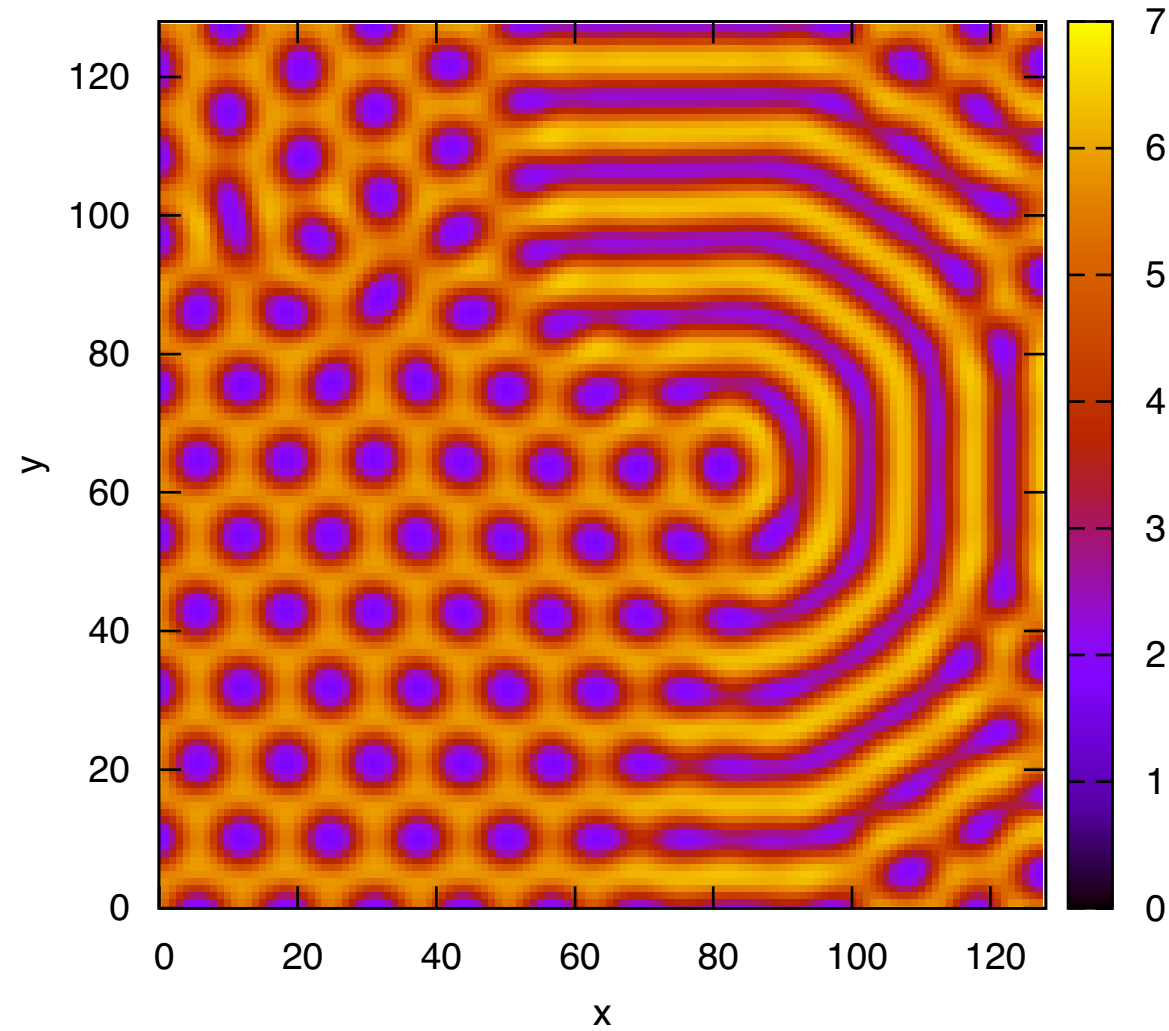
-> *BDF (Backward Differentiation Formulae, higher-order stiffly stable solver)*

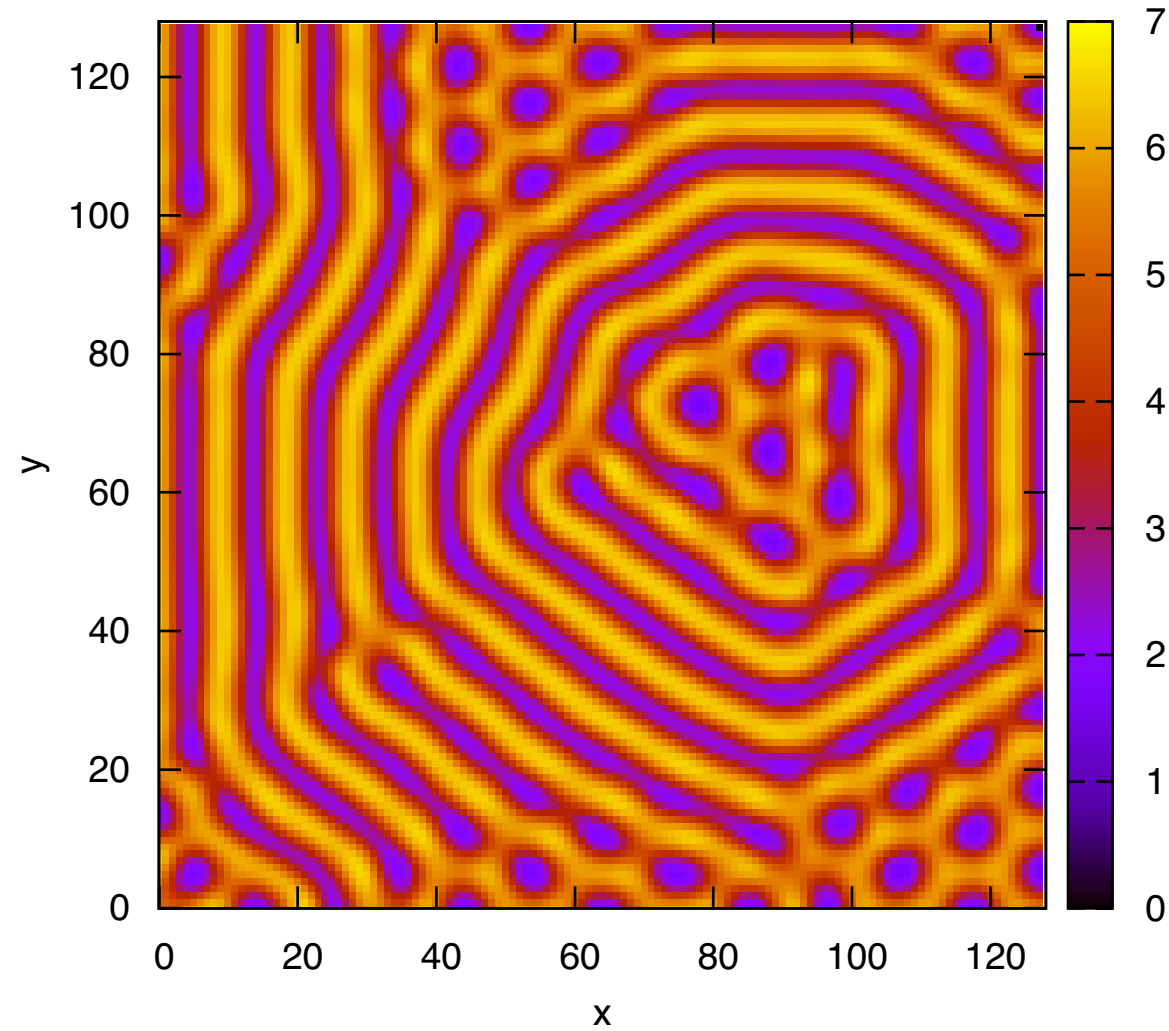
-> *simulation time: 5,000 -> runtime: about 30h*



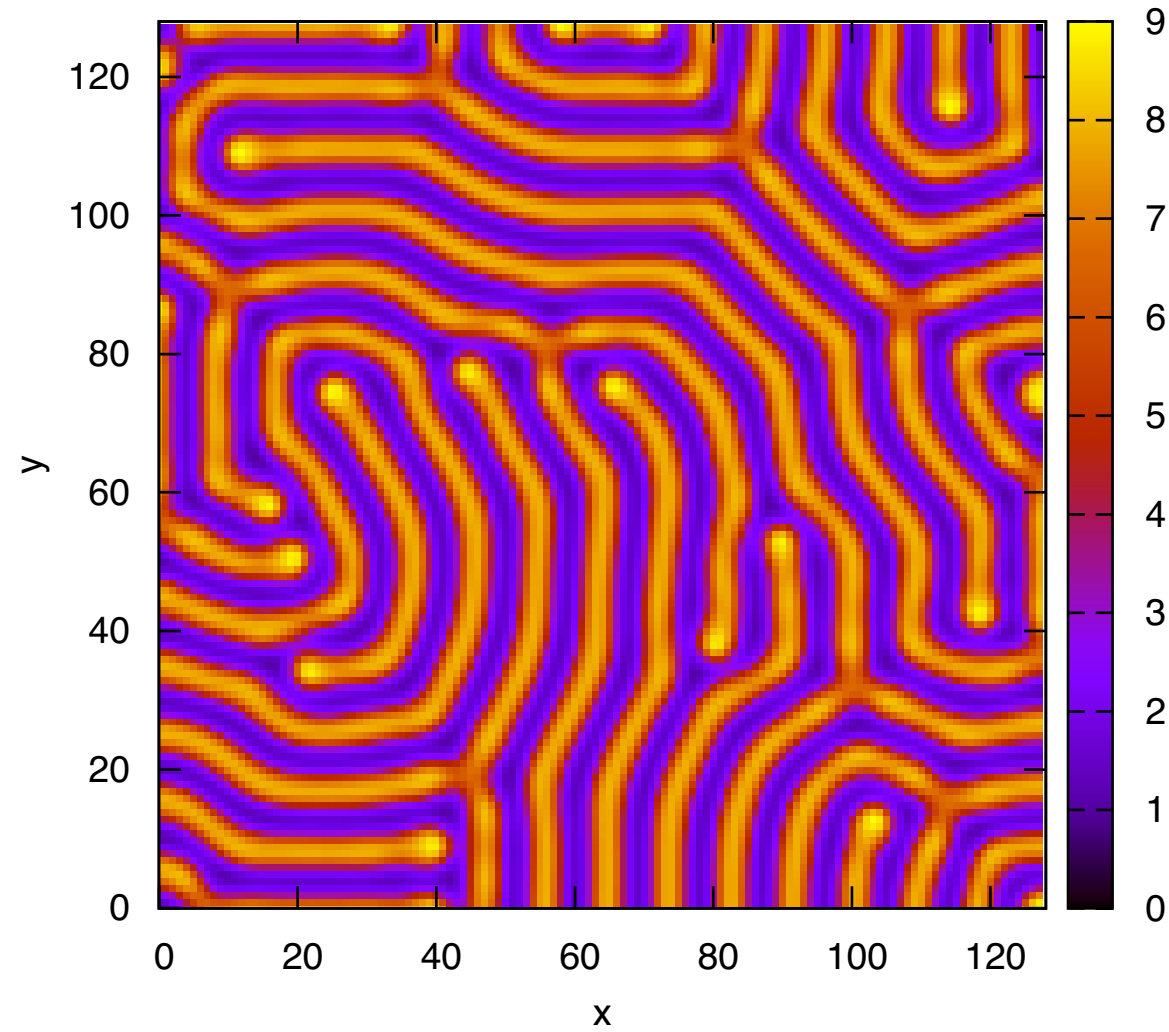


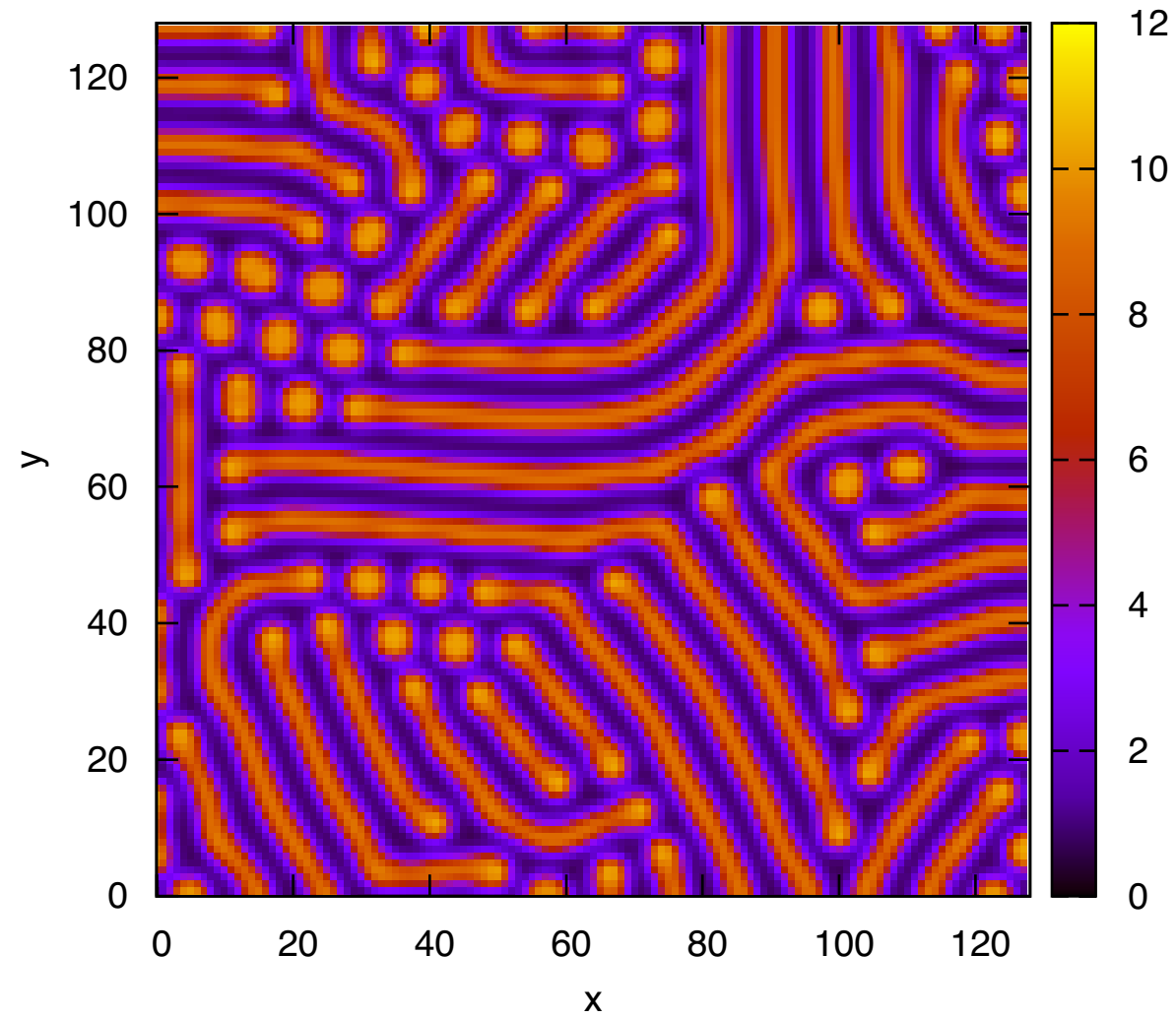


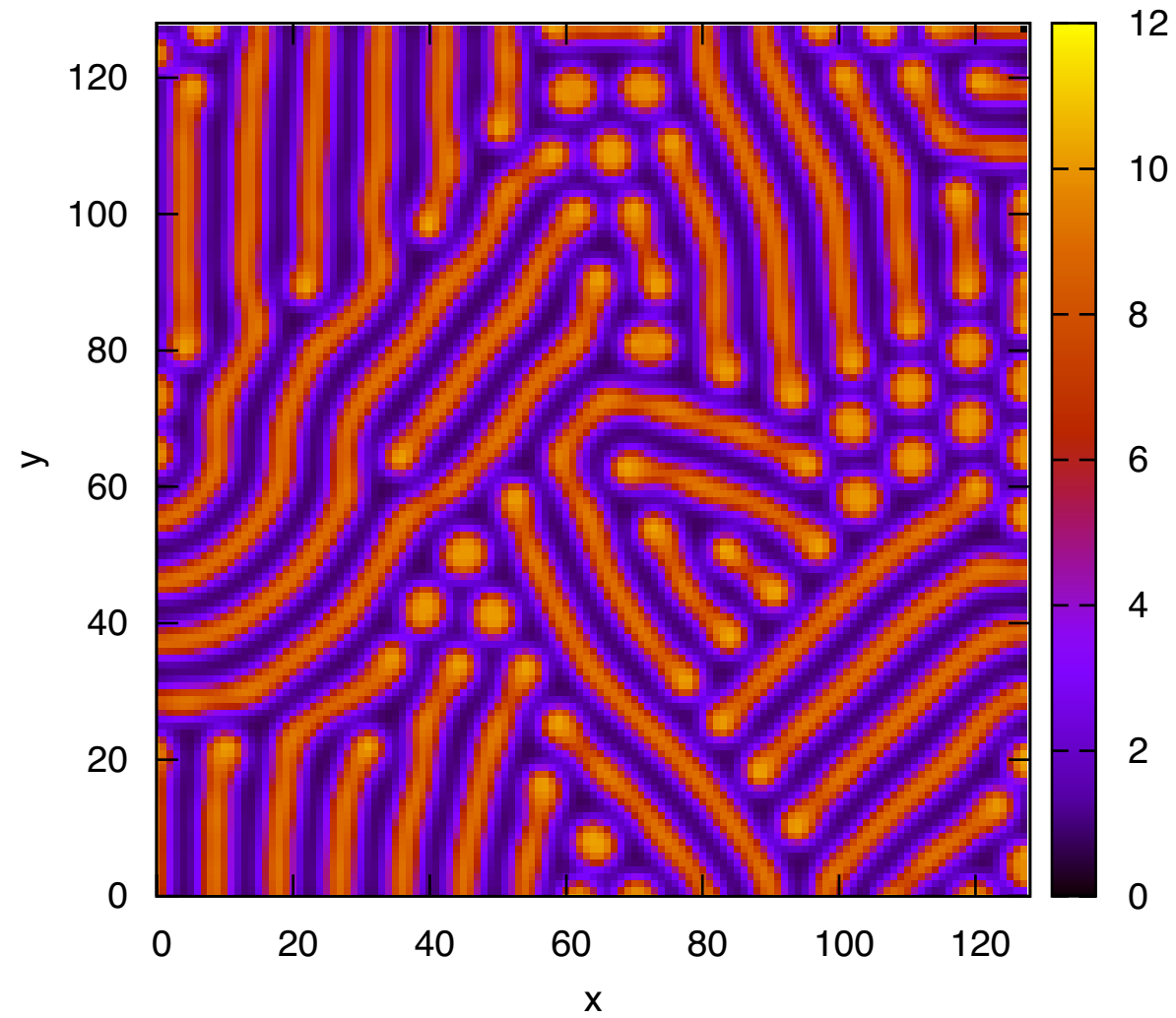


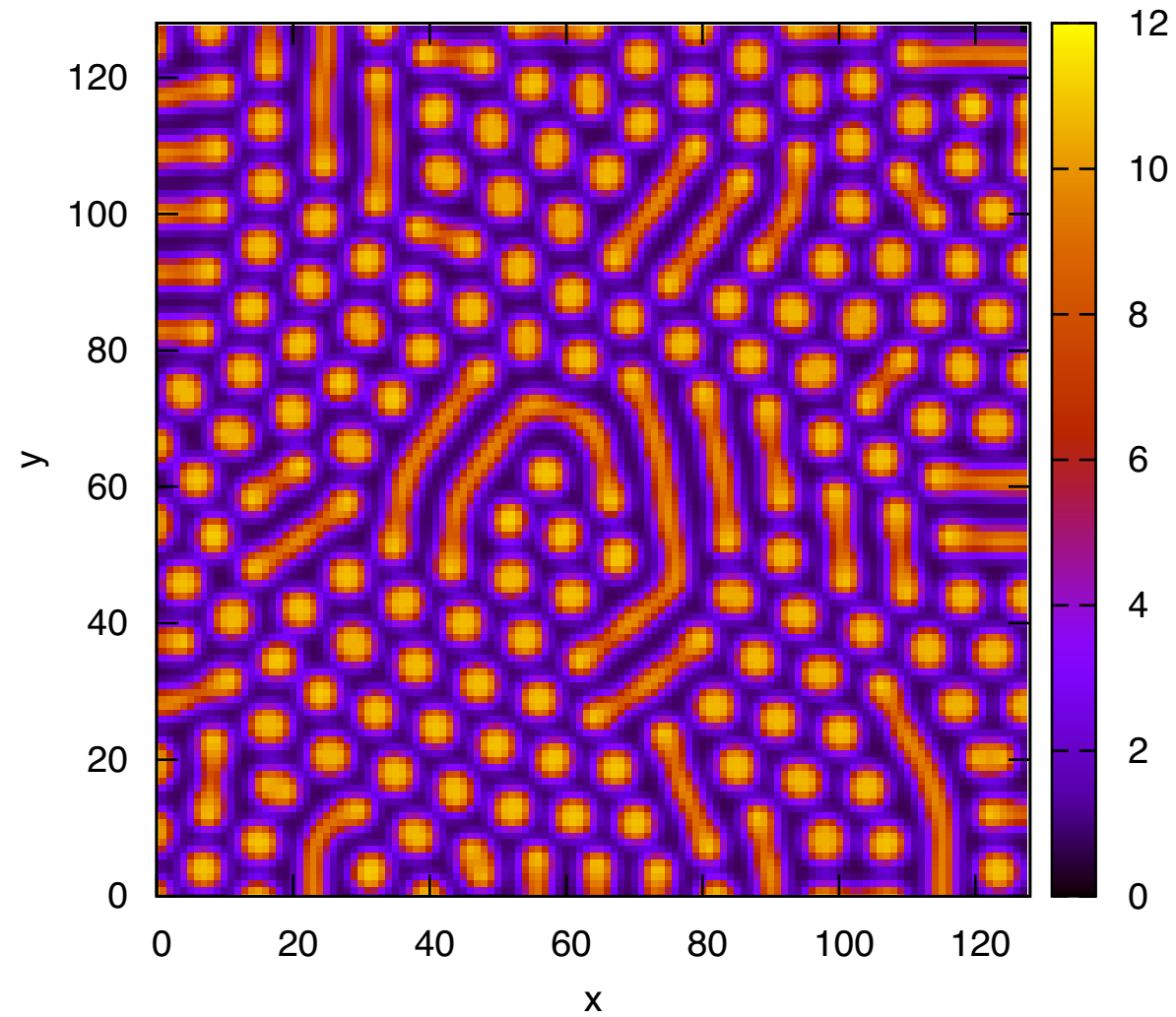


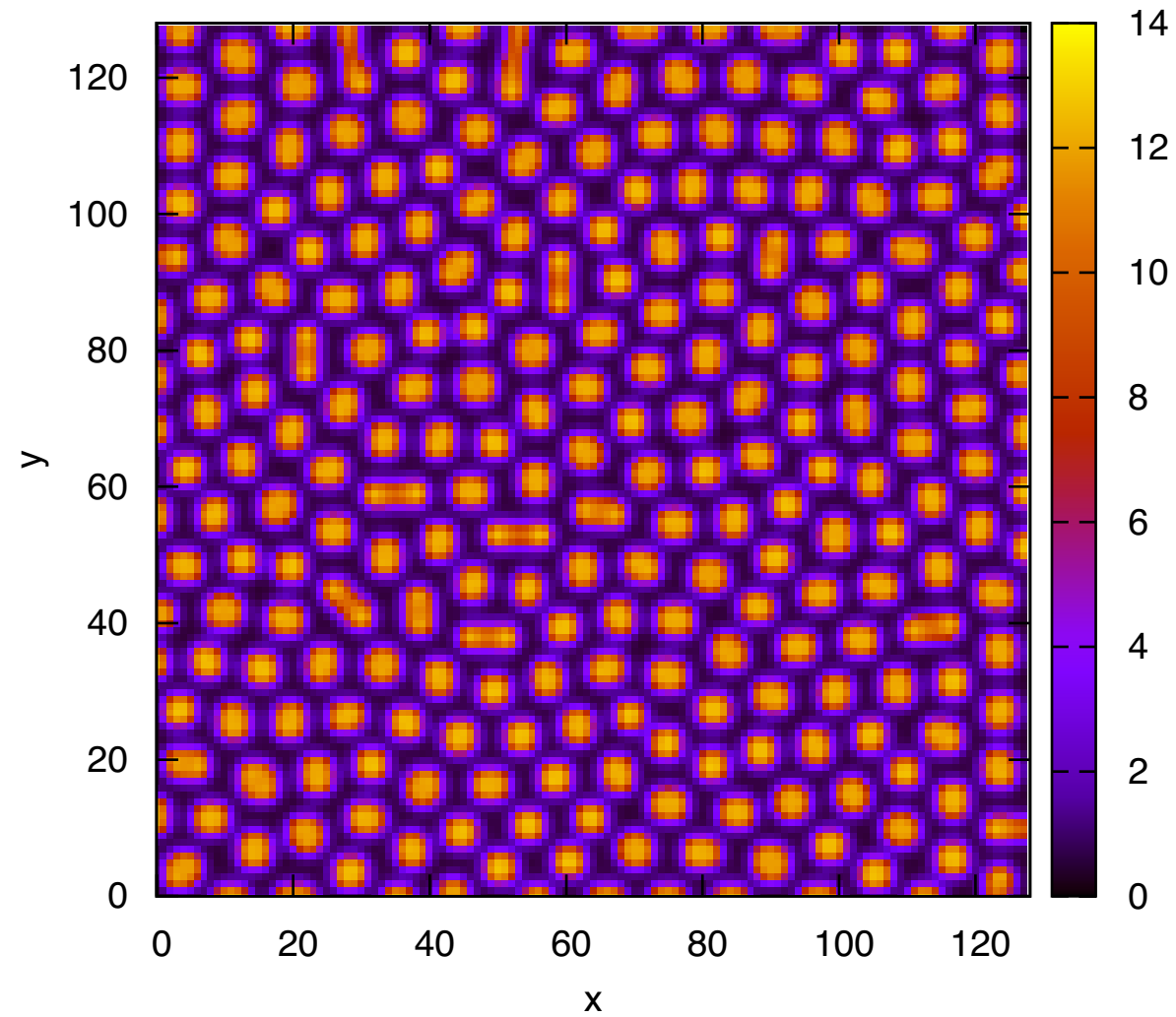












# **EXAMPLE 3:**

## **PHASE VARIATION IN MULTISTRAIN CELL COLONIES**

### □ phase variation

-> *method for dealing with rapidly varying environments without requiring random mutations*

### □ contingency genes

-> *populations include variants adapted to “foreseeable” frequently encountered environmental or selective conditions*

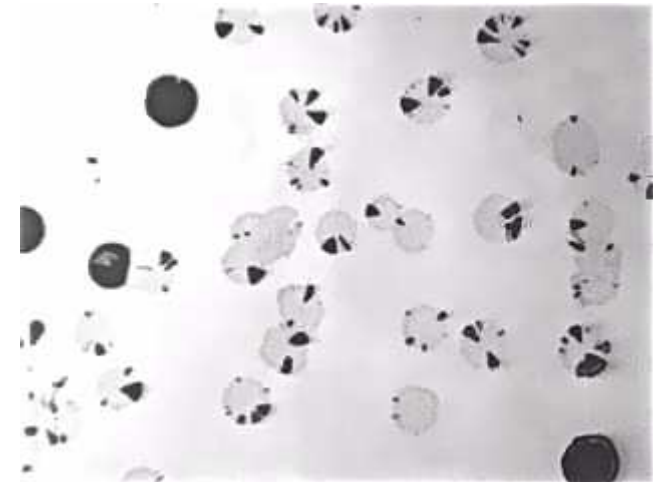
### □ stochastic gene switching process

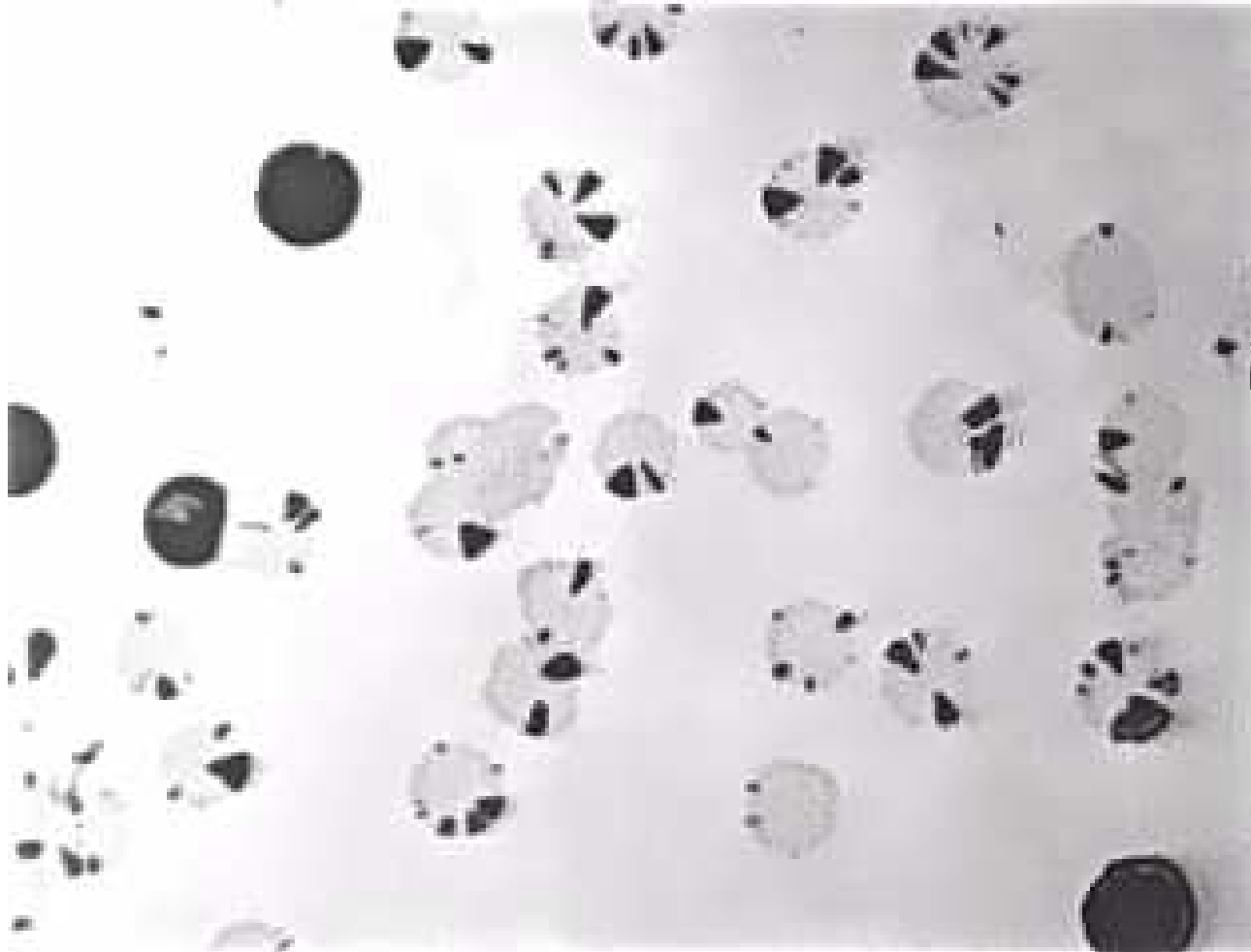
-> *controlled by reversible gene mutations, inversions, or epigenetic modification*

-> *e.g. switch between two phenotypes A, B*

### □ colonial sectoring

-> *observable effect in cultures grown in vitro*





*(courtesy of N Saunders)*



*Microbiology* (2003), 149, 485–495

DOI 10.1099/mic.0.25807-0

### Mutation rates: estimating phase variation rates when fitness differences are present and their impact on population structure

Nigel J. Saunders,<sup>1†</sup> E. Richard Moxon<sup>1</sup> and Mike B. Gravenor<sup>2</sup>

#### Correspondence

Nigel J. Saunders

saunders@molbiol.ox.ac.uk

<sup>1</sup>Molecular Infectious Diseases Group, Institute of Molecular Medicine, University of Oxford, Headington, Oxford OX3 9DS, UK

<sup>2</sup>Institute for Animal Health, Compton, Berkshire RG20 7NN, UK

---

Phase variation is a mechanism of ON–OFF switching that is widely utilized by bacterial pathogens. There is currently no standardization to how the rate of phase variation is determined experimentally.

Microbiology (2003), 149, 485–495

DOI 10.1099/mic.0.25807-0

Mutation rates: estimating phase variation rates when fitness differences are present and their impact on population structure

Nigel J. Saunders<sup>1</sup>, Richard Moxon<sup>1</sup> and Mike B. Gravenor<sup>2</sup>

Correspondence

Nigel J. Saunders

saunders@microbiology.ox.ac.uk

<sup>1</sup>Molecular Infectious Diseases Group, Institute of Molecular Medicine, University of Oxford, Headington, Oxford OX3 9DS, UK

<sup>2</sup>Institute for Animal Health, Compton, Berkshire RG20 7NN, UK

Phase variation is a mechanism of ON–OFF switching that is widely utilized by bacterial pathogens. There is currently no standardization to how the rate of phase variation is determined experimentally.

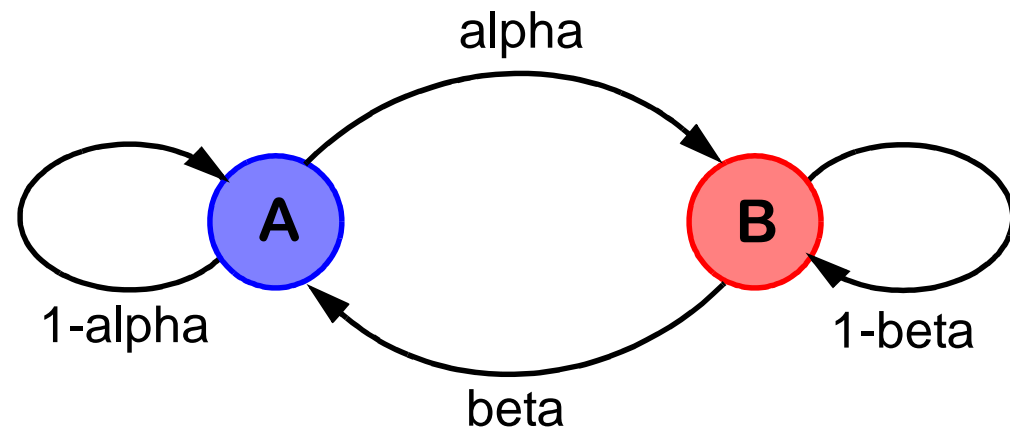
□ **two cell types: phenotype A and B**

□ **cell divide**

- > *cell division may involve mutation of the offspring*
- > *parent cell keeps its phenotype*

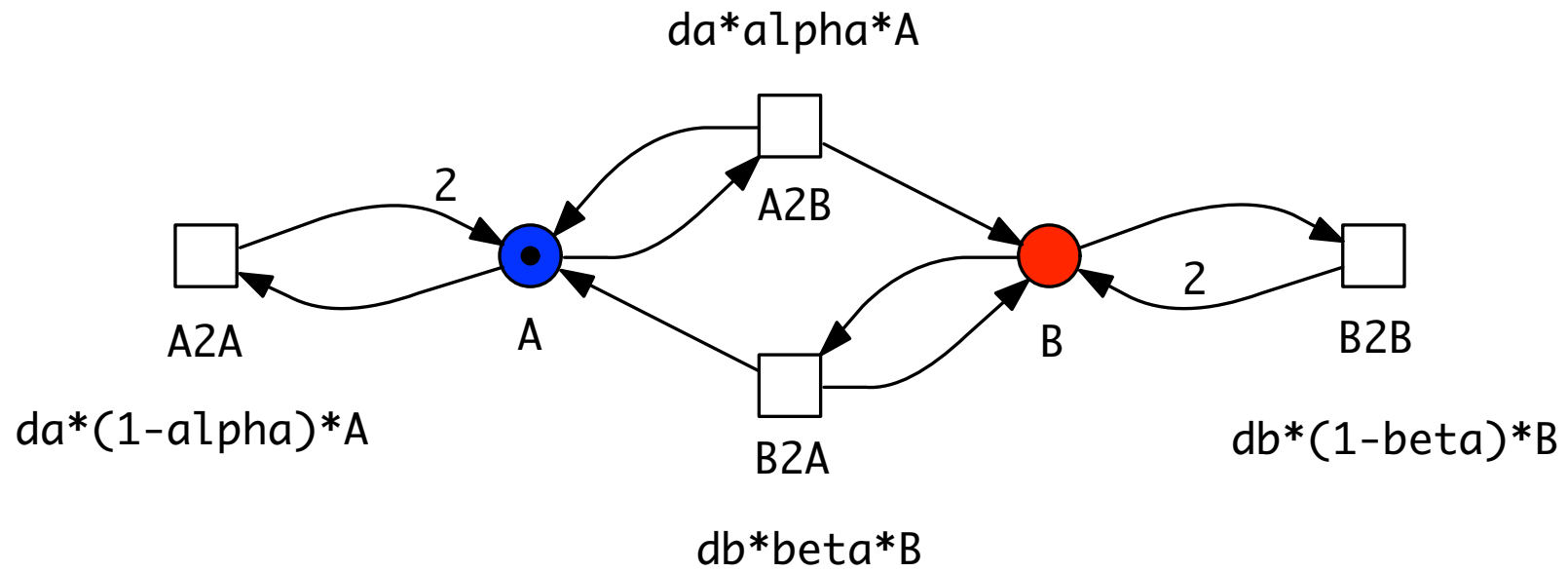
□ **model parameters**

- > *alpha = beta - mutation rates*
- > *da, db - fitness of A, B*
- > *da/db - relative fitness*

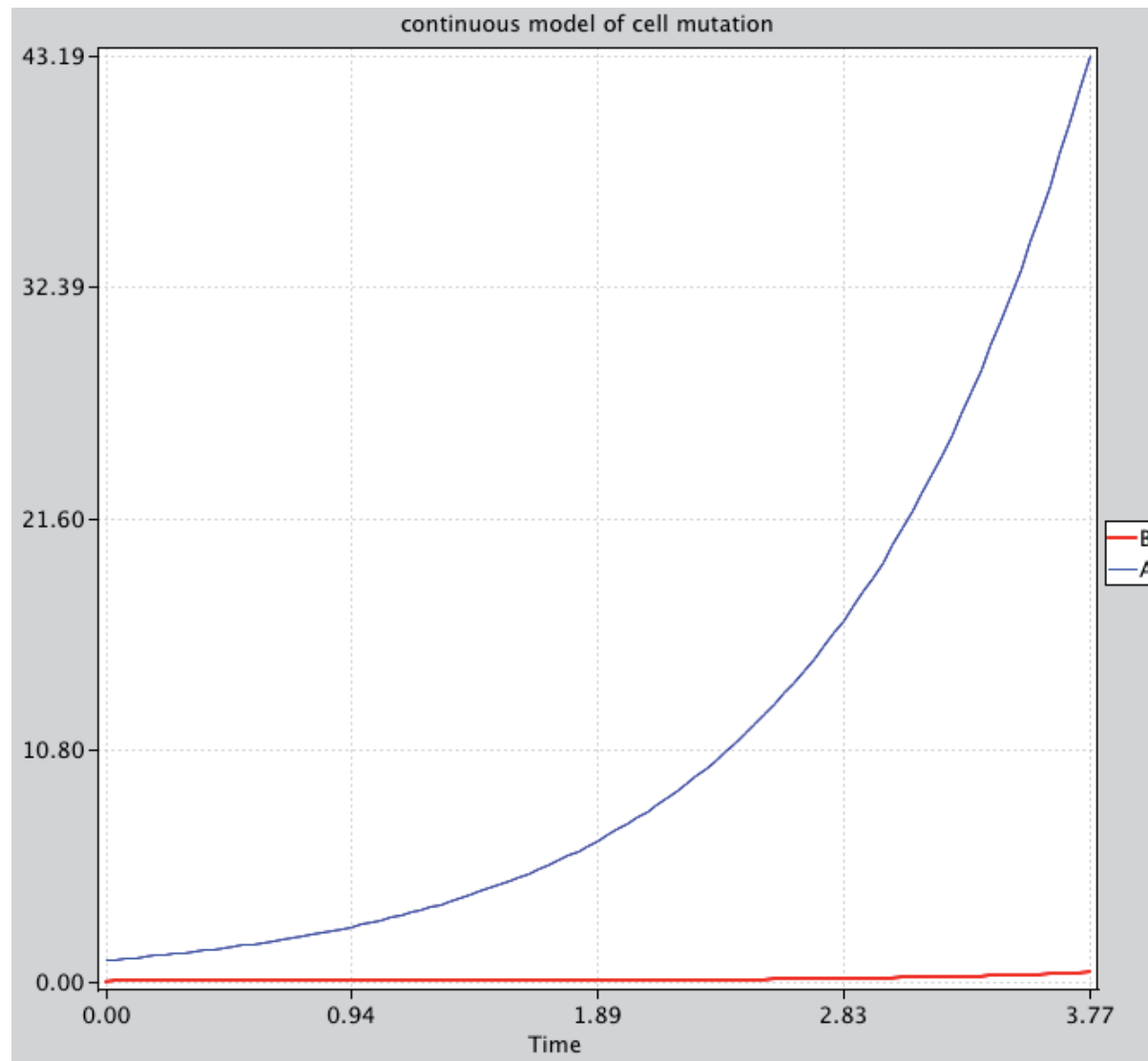


□ **output**

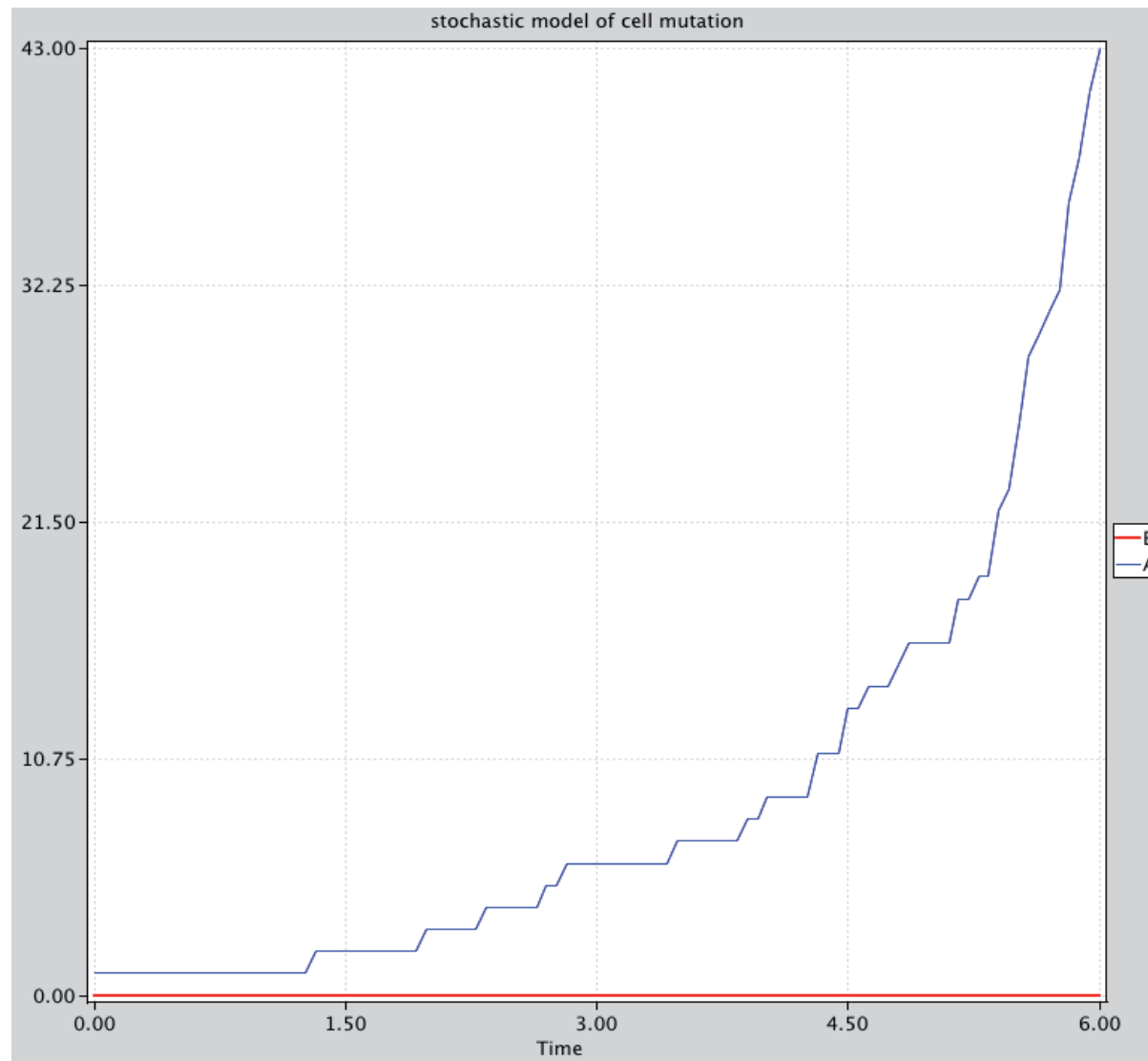
- > *total number of cells*
- > *proportion of A =  $A / (A + B)$*
- > *proportion of B =  $B / (A + B)$*



# EX3: CELL COLONIES, CONTINUOUS PLOT



# EX3: CELL COLONIES, STOCHASTIC PLOT



# ... AND THEN THERE WAS COLOUR

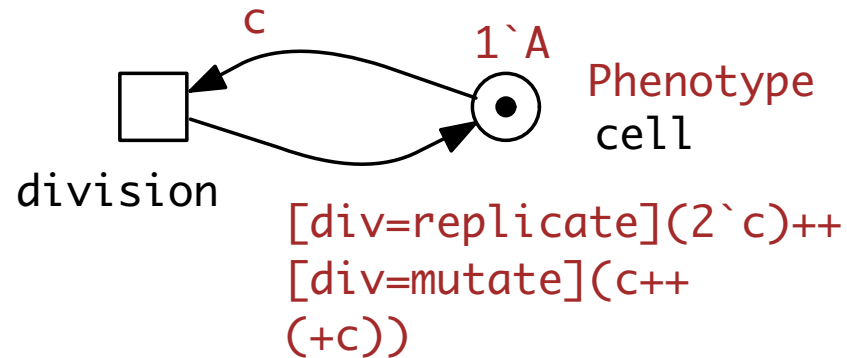


*Kew Gardens,  
24/04/2011*



**colorset** Phenotype = **enum with** A, B;

**colorset** DivisionType = **enum with** replicate , mutate ;



```
(c=A) & (div=replicate) : cell*da*(1-alpha)
(c=A) & (div=mutate) : cell*(da*alpha)
(c=B) & (div=replicate) : cell*(db*(1-beta))
(c=B) & (div=mutate) : cell*(db*beta)
```



```
colorset Phenotype = enum with A, B;  
colorset DivisionType = enum with replicate, mutate;
```



**ADDING SPACE**

```
[div=replicate](2`c)++  
[div=mutate](c-  
(+))
```

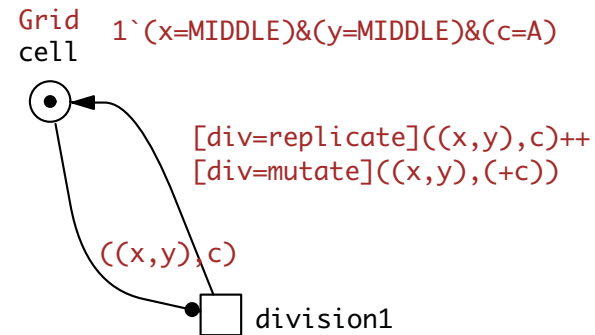
**CONTROLLING COLONY SPREADING**

```
(c=A) & (div=replicate) : cell*(da*(1-alpha))  
(c=A) & (div=mutate) : cell*(da*alpha)  
(c=B) & (div=replicate) : cell*(db*(1-beta))  
(c=B) & (div=mutate) : cell*(db*beta)
```

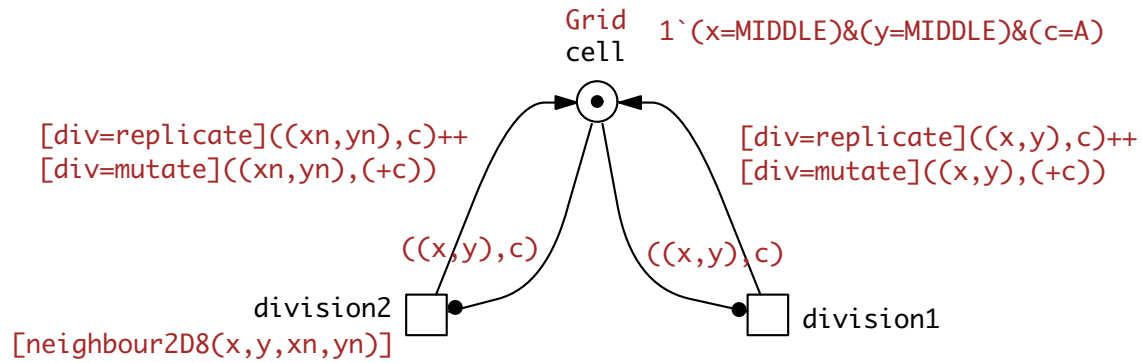
**CONTROLLING THICKNESS**

**CONTROLLING COLONY SIZE**

**colorset Grid = product with Grid2D x Phenotype;**

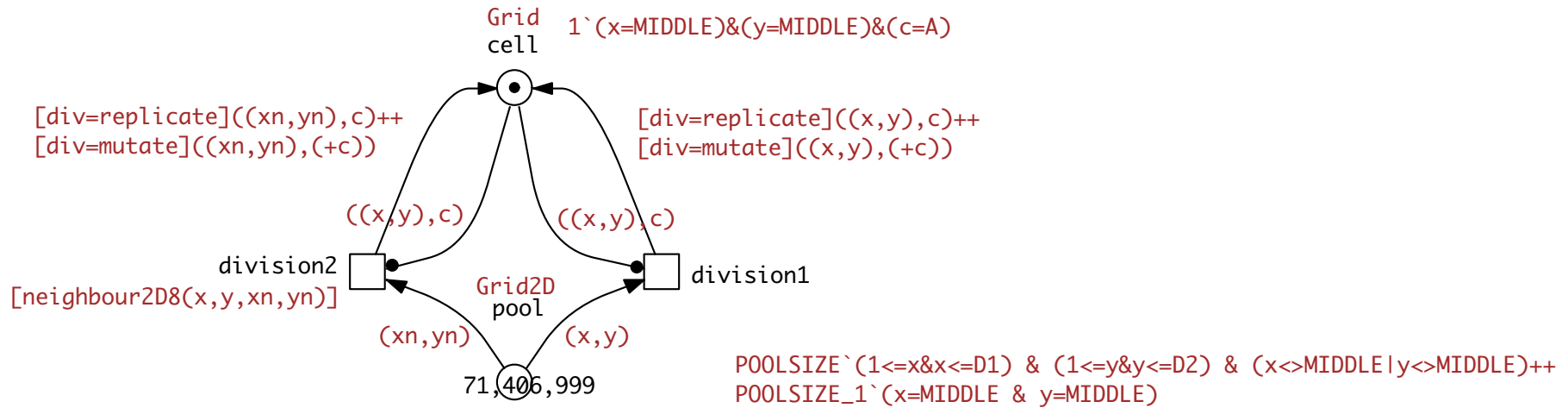


**colorset Grid = product with Grid2D x Phenotype;**



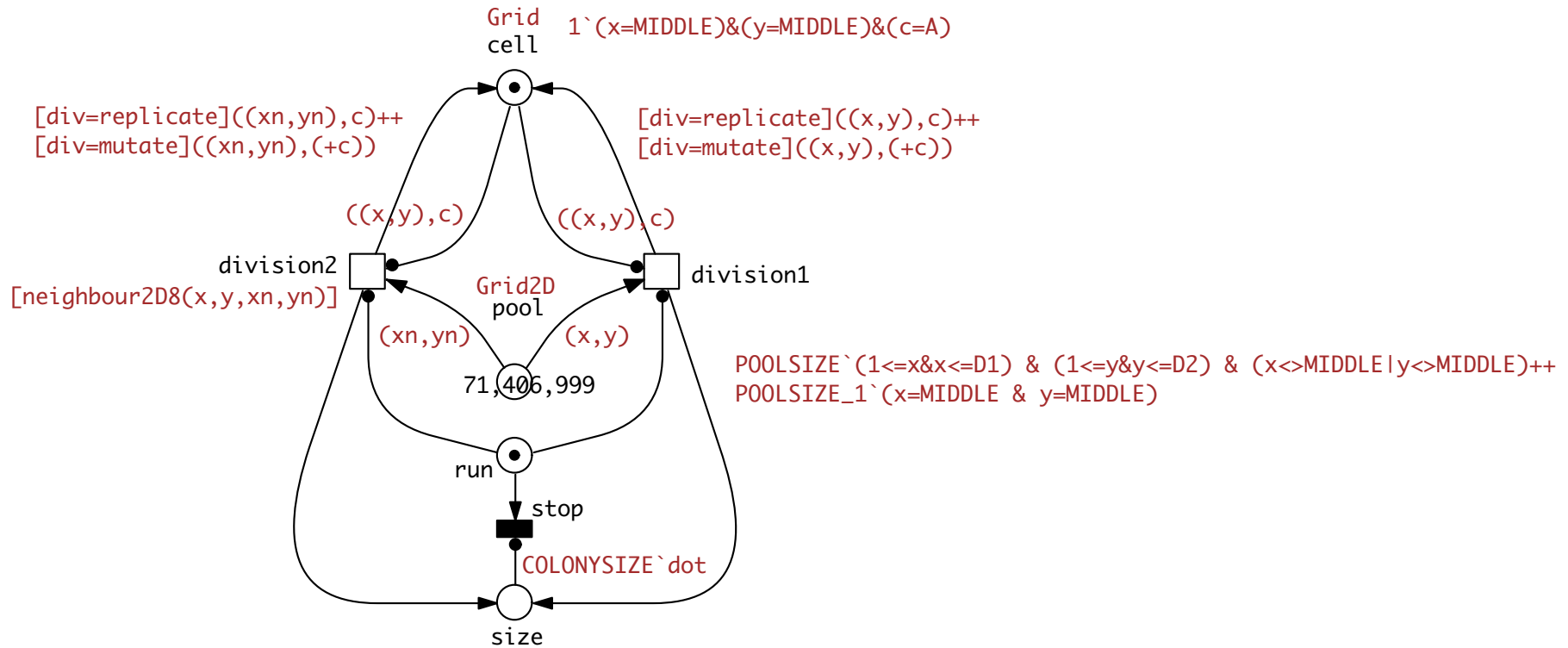
# EX3: CELL COLONIES, CONTROLLING THICKNESS

**colorset Grid = product with Grid2D x Phenotype;**



# EX3: CELL COLONIES, CONTROLLING COLONY SIZE

**colorset Grid = product with Grid2D x Phenotype;**



### ❑ model assumptions

- > *“If phase variation occurs, the progeny consists of one A and one B”  
(Saunders 2003)*
- > *It is always the mutant who goes to a neighbouring position, if any.*
- > *constant biofilm thickness (so far)*

### ❑ colony size - 24 h

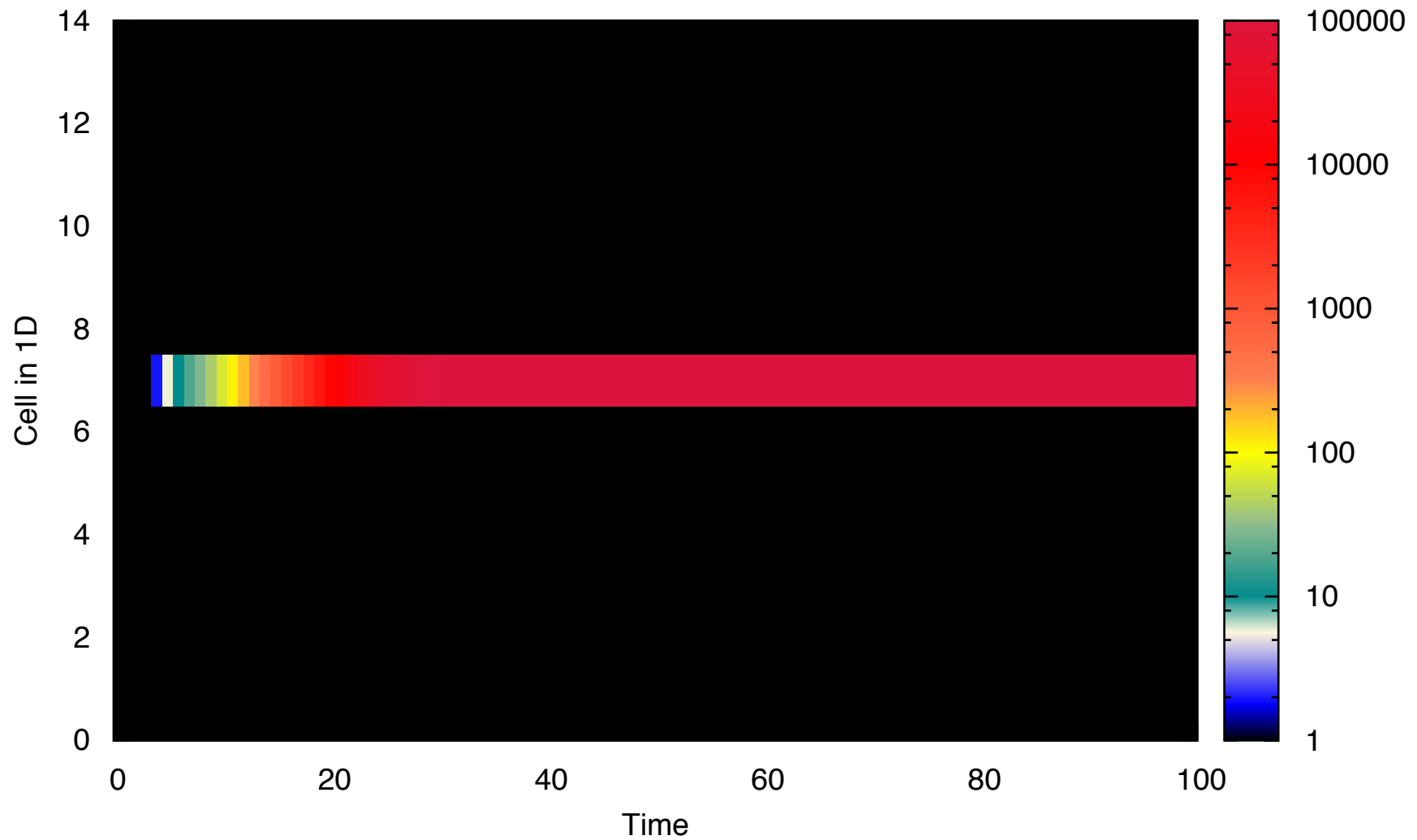
- > *25 generations: 33.5 E+06*
- > *26 generations: 67 E+06*
- > *COLONYSIZE = 70,000,000*

### ❑ grid size

- > *61 x 61 grid: 11,163 P / 131,044 T; unfolding: 152 sec;*
- > *101 x 101 grid: 30,603 P / 362,404 T; unfolding: 9 min;*
- > *runtime 1 stoch. simulation: 35-40 minutes*

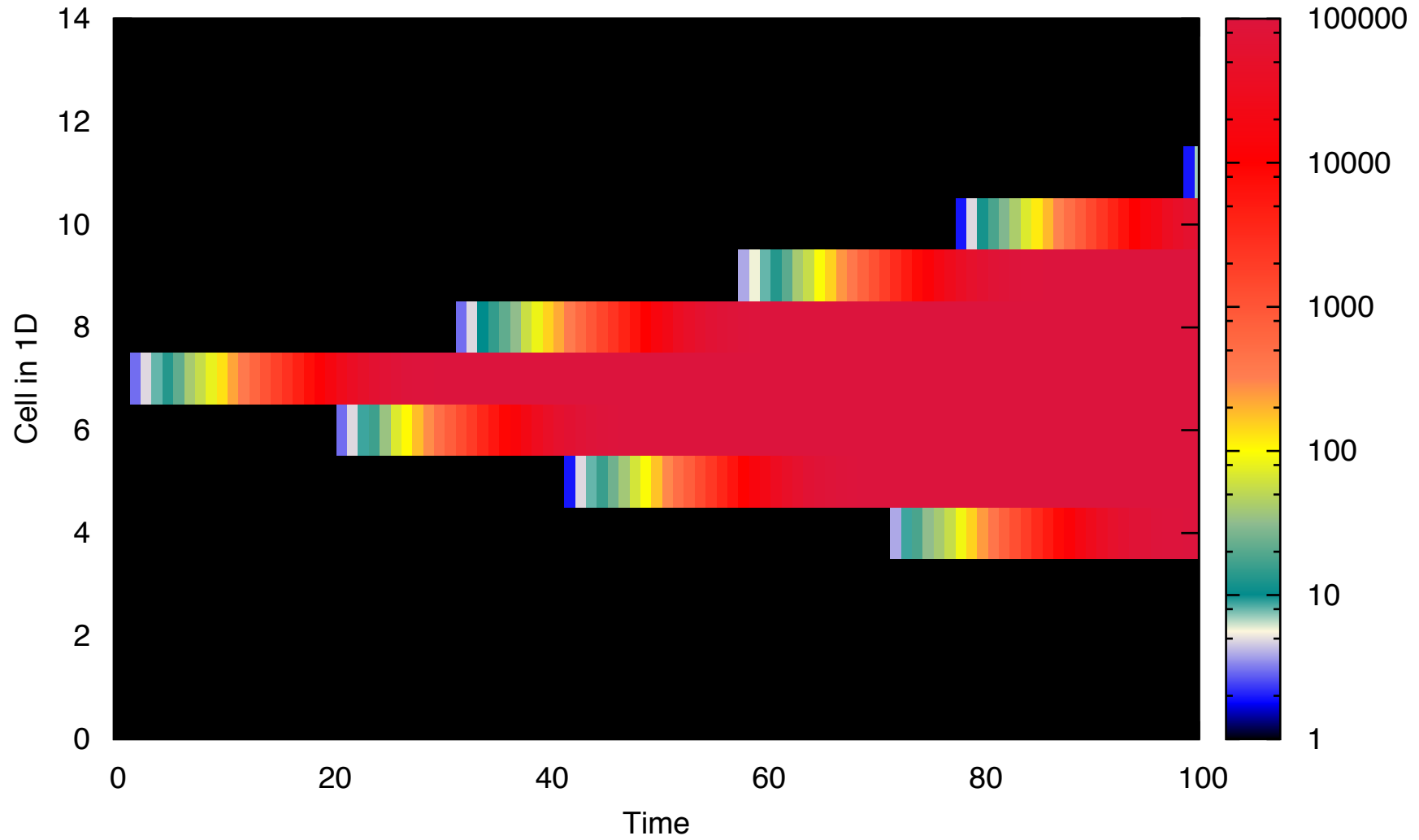
# ... SOME EXPERIMENTS

## Ex3: 1D15 - VARYING MOBILITY, GAMMA = 100

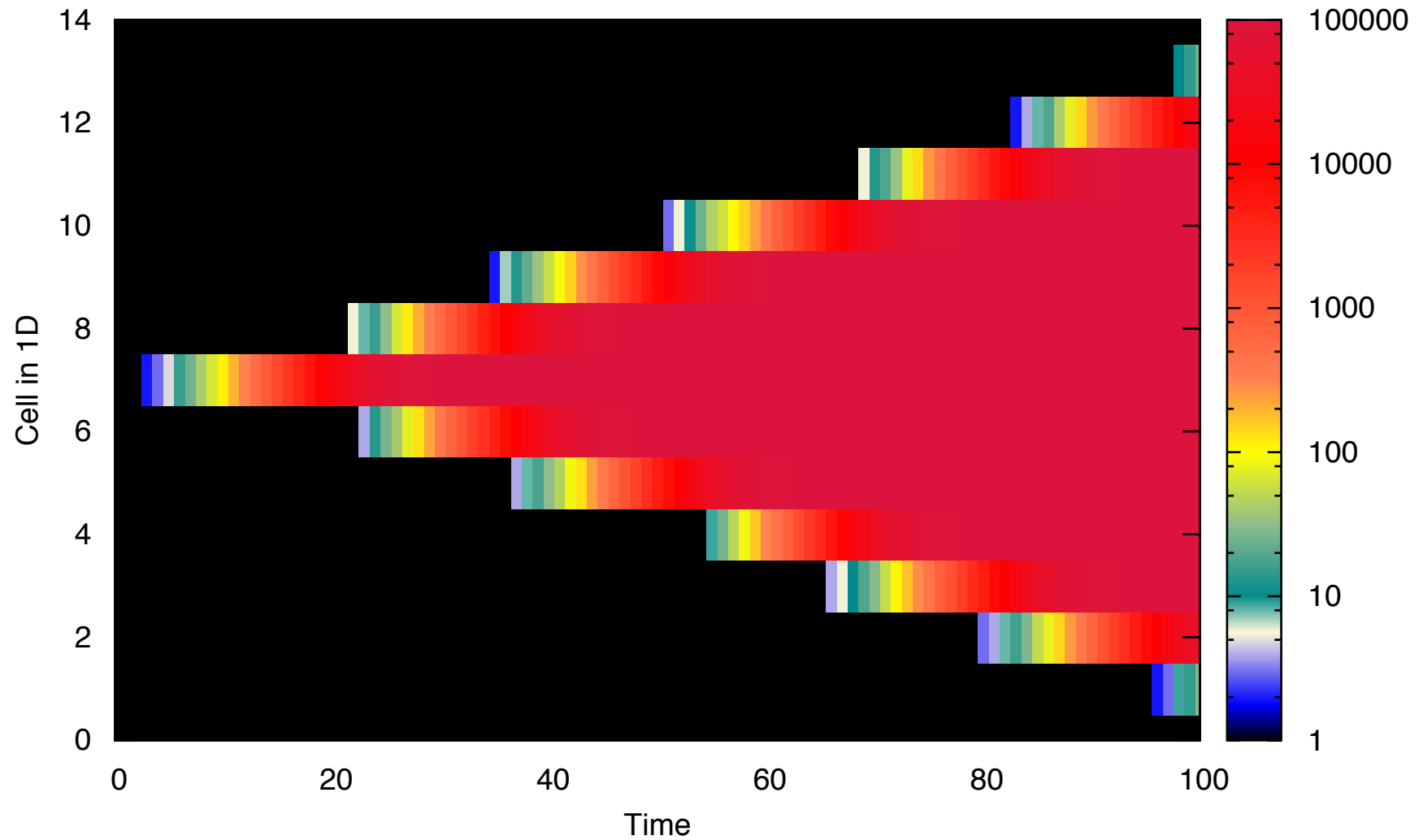




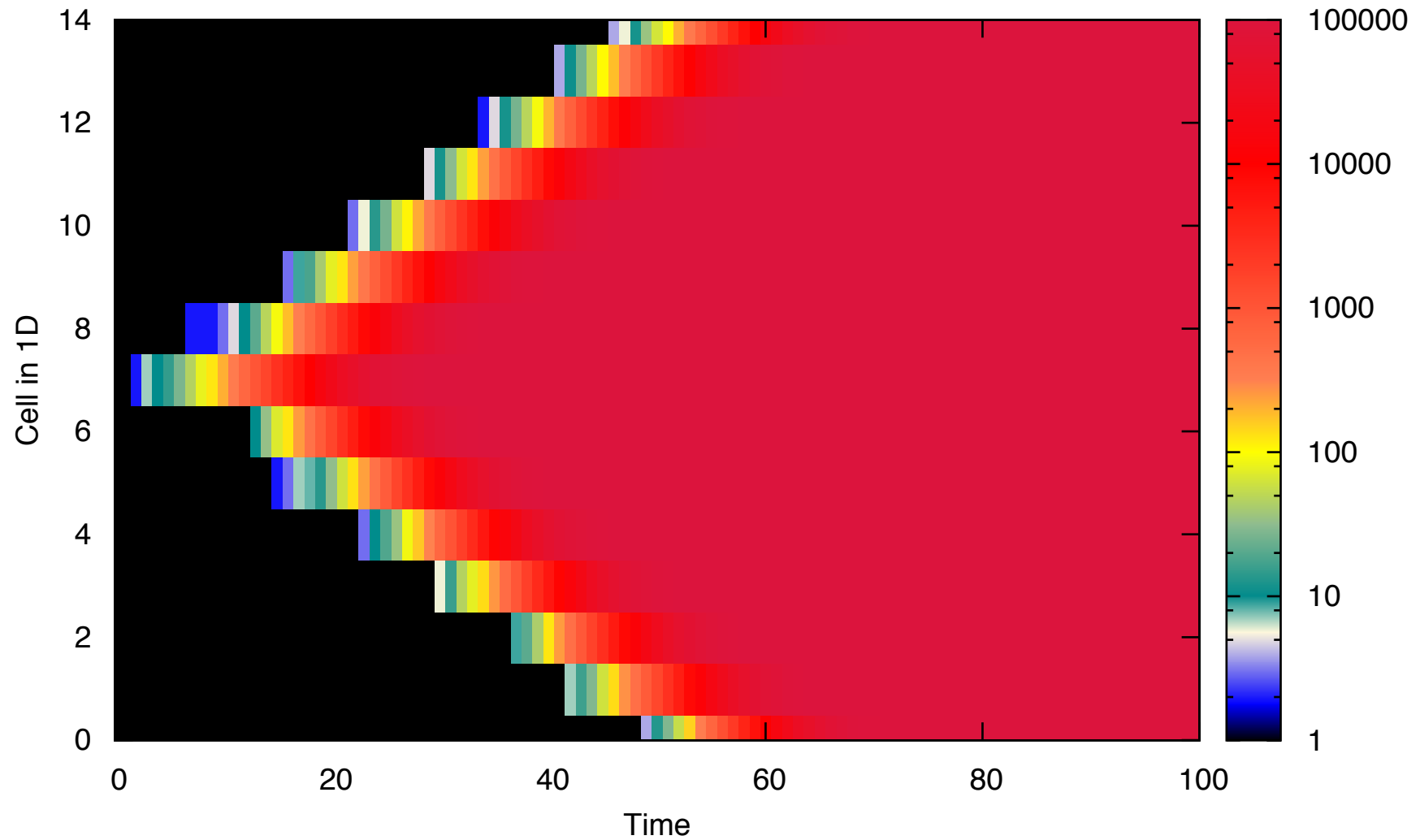
# Ex3: 1D15 - VARYING MOBILITY, GAMMA = 99.999



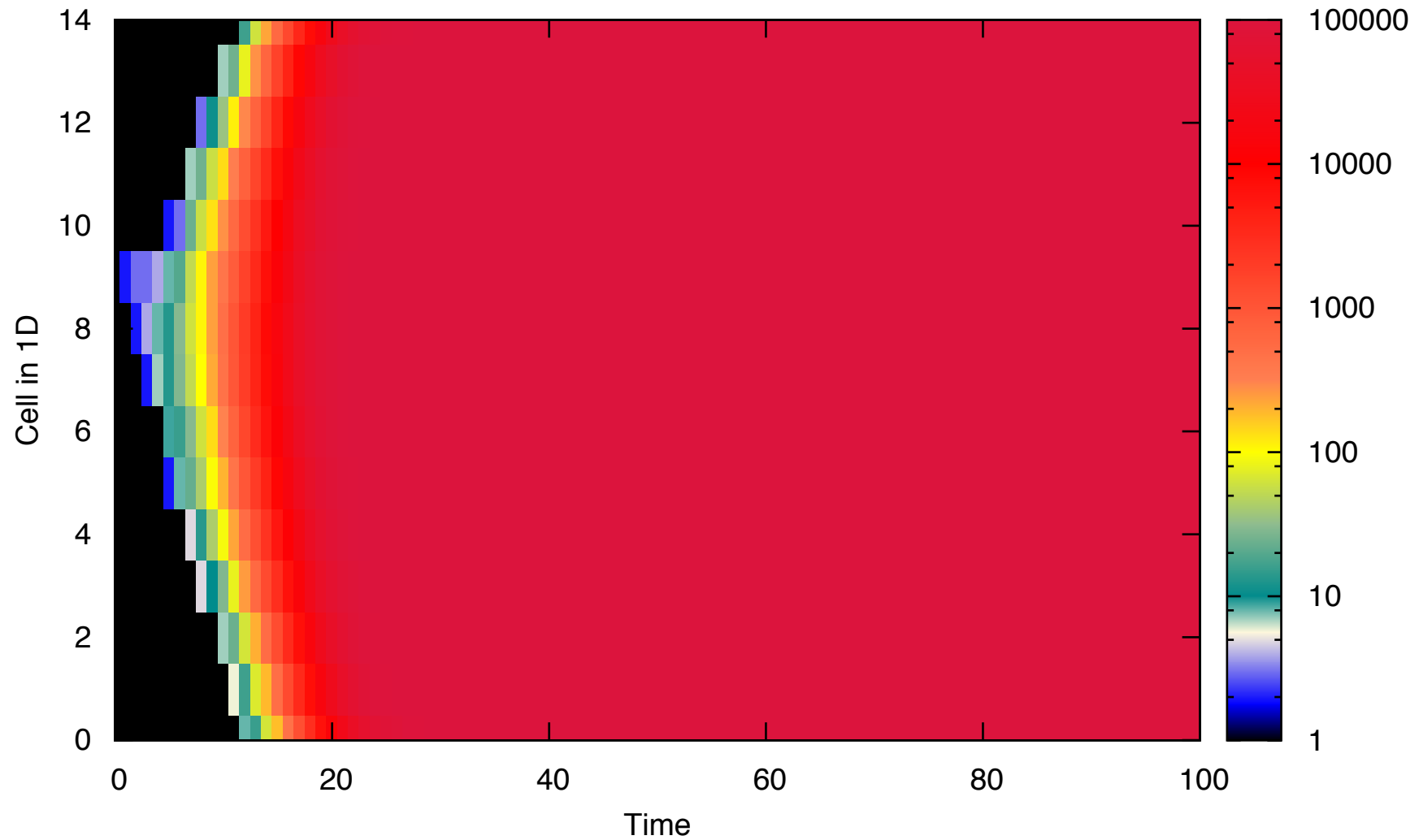
# Ex3: 1D15 - VARYING MOBILITY, GAMMA = 99.99



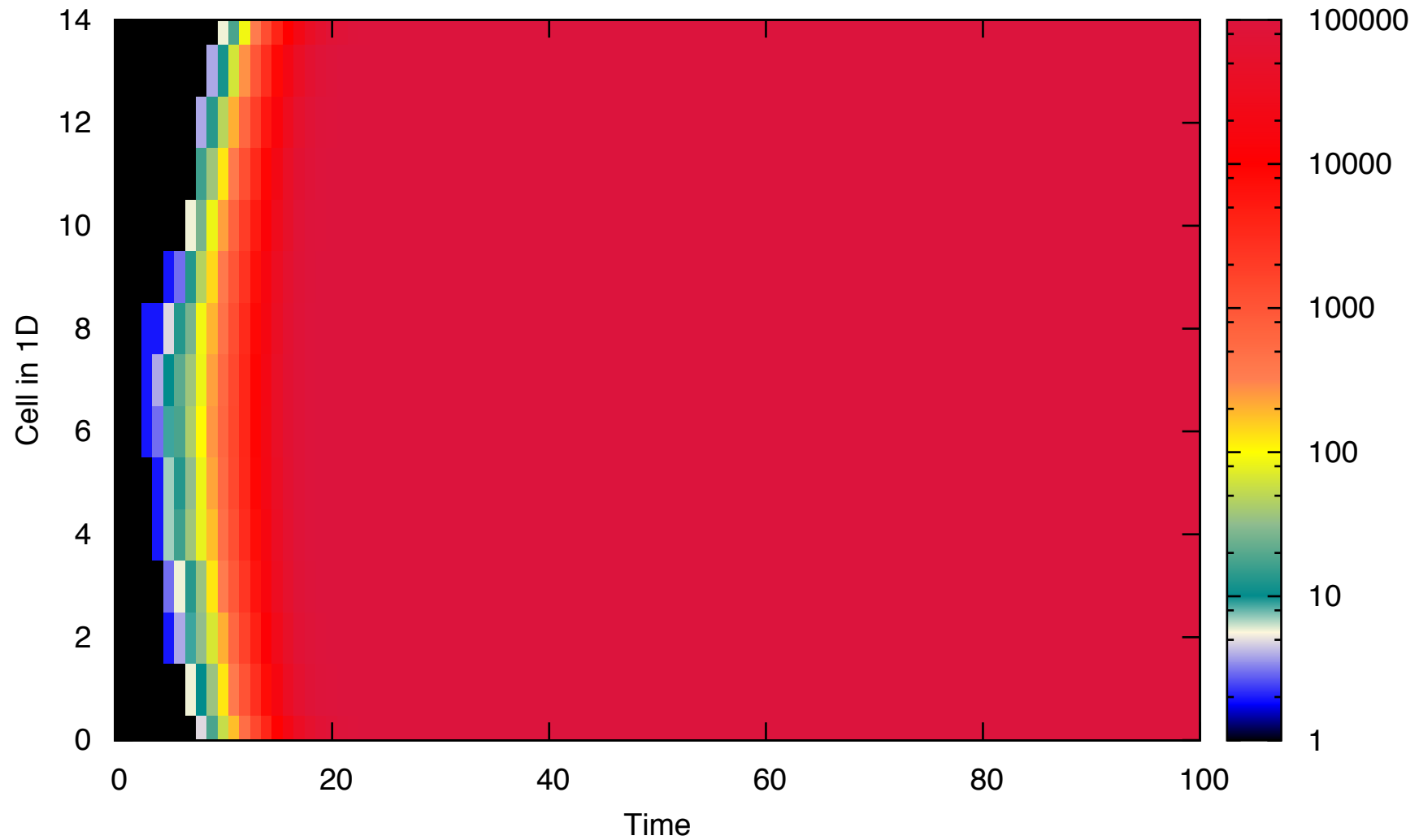
# Ex3: 1D15 - VARYING MOBILITY, GAMMA = 90

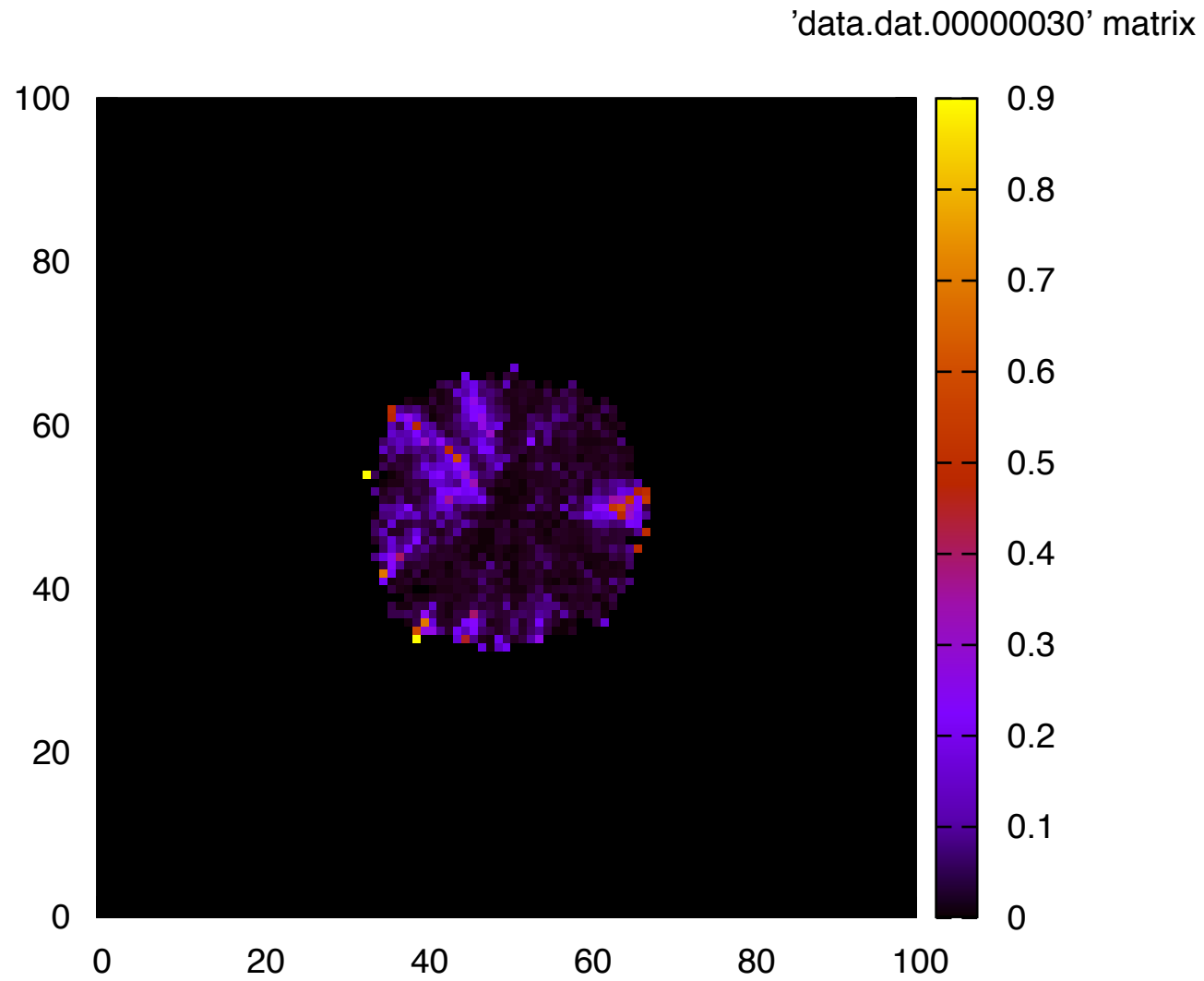


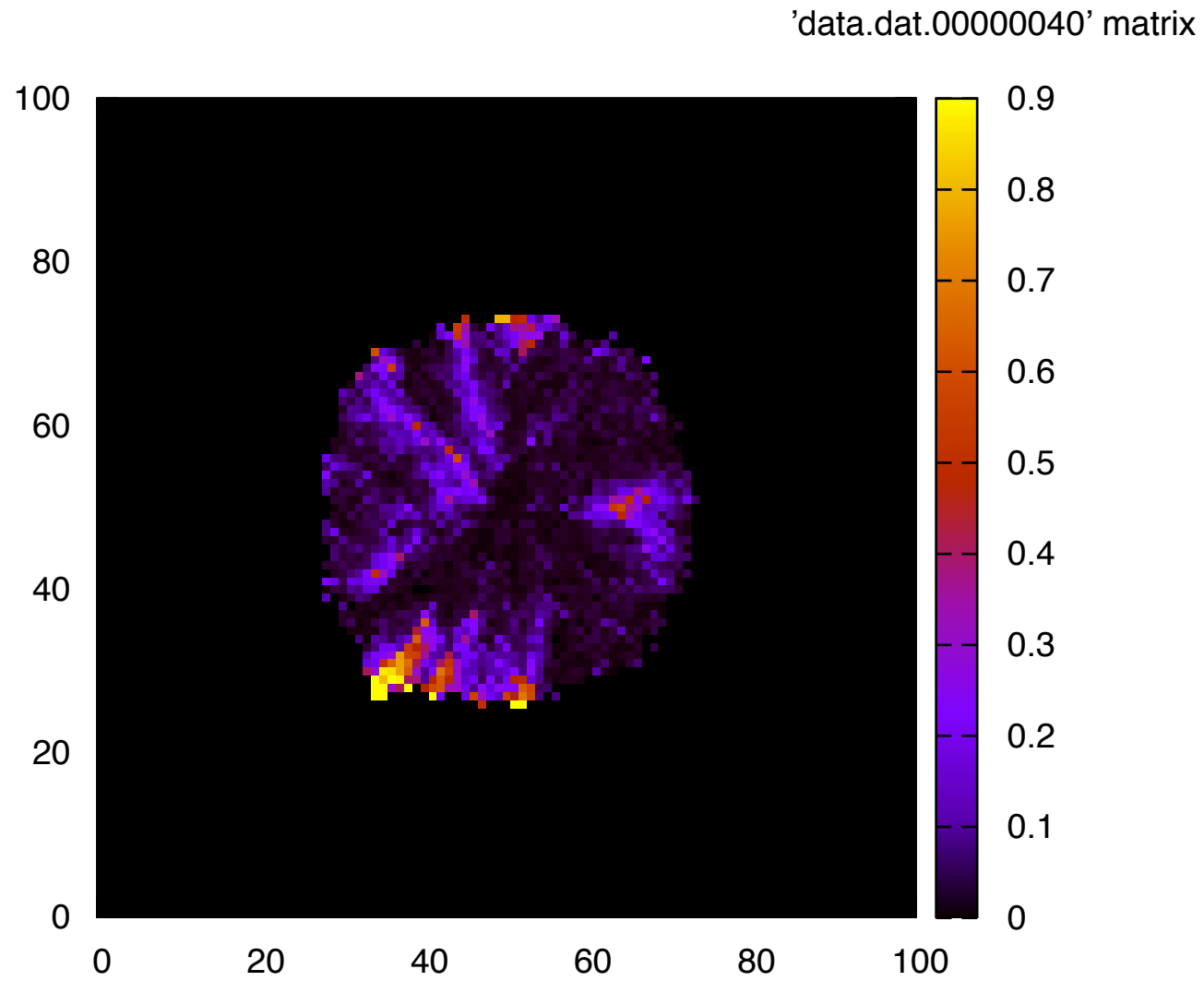
## Ex3: 1D15 - VARYING MOBILITY, GAMMA = 50

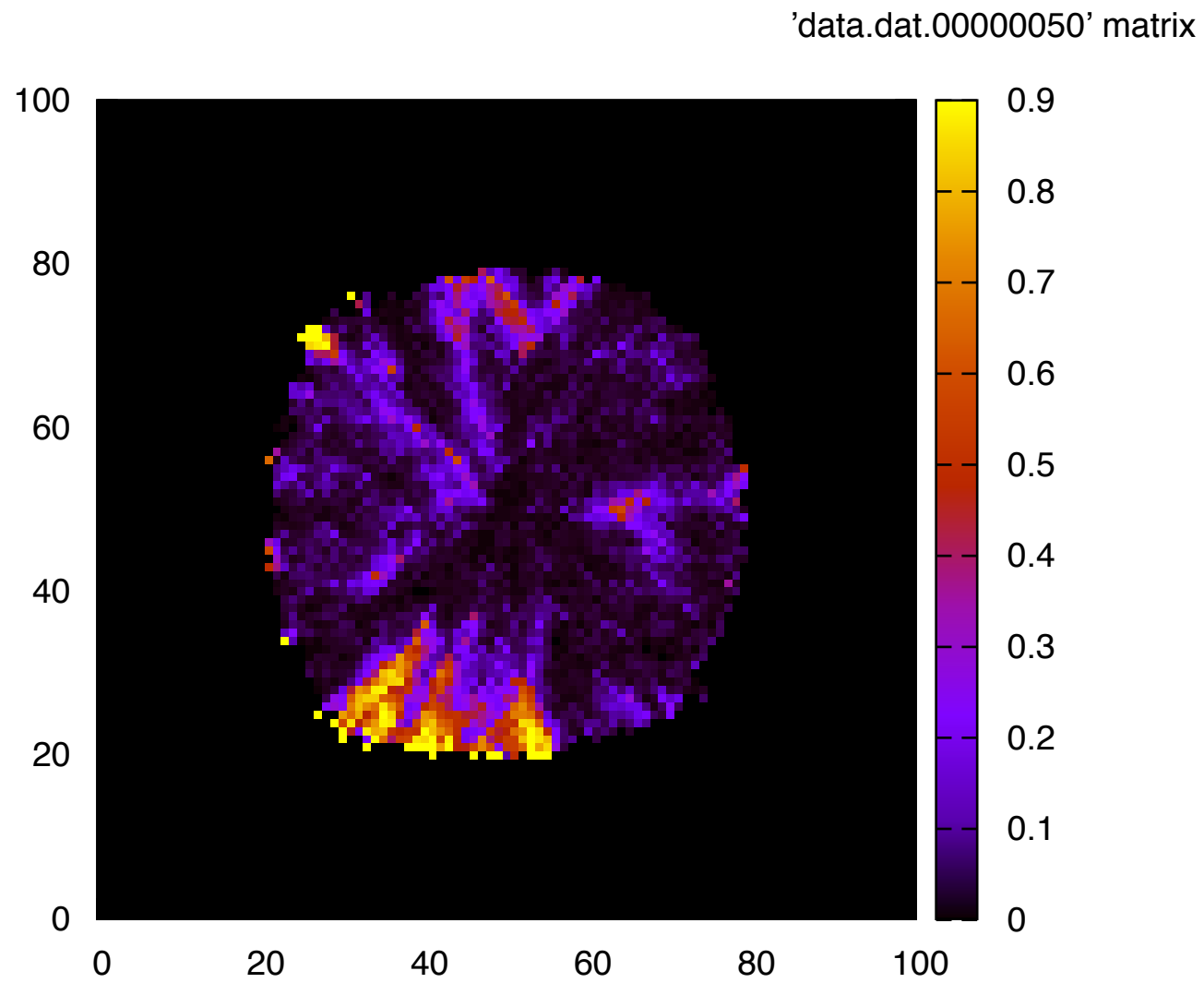


## Ex3: 1D15 - VARYING MOBILITY, GAMMA = 1

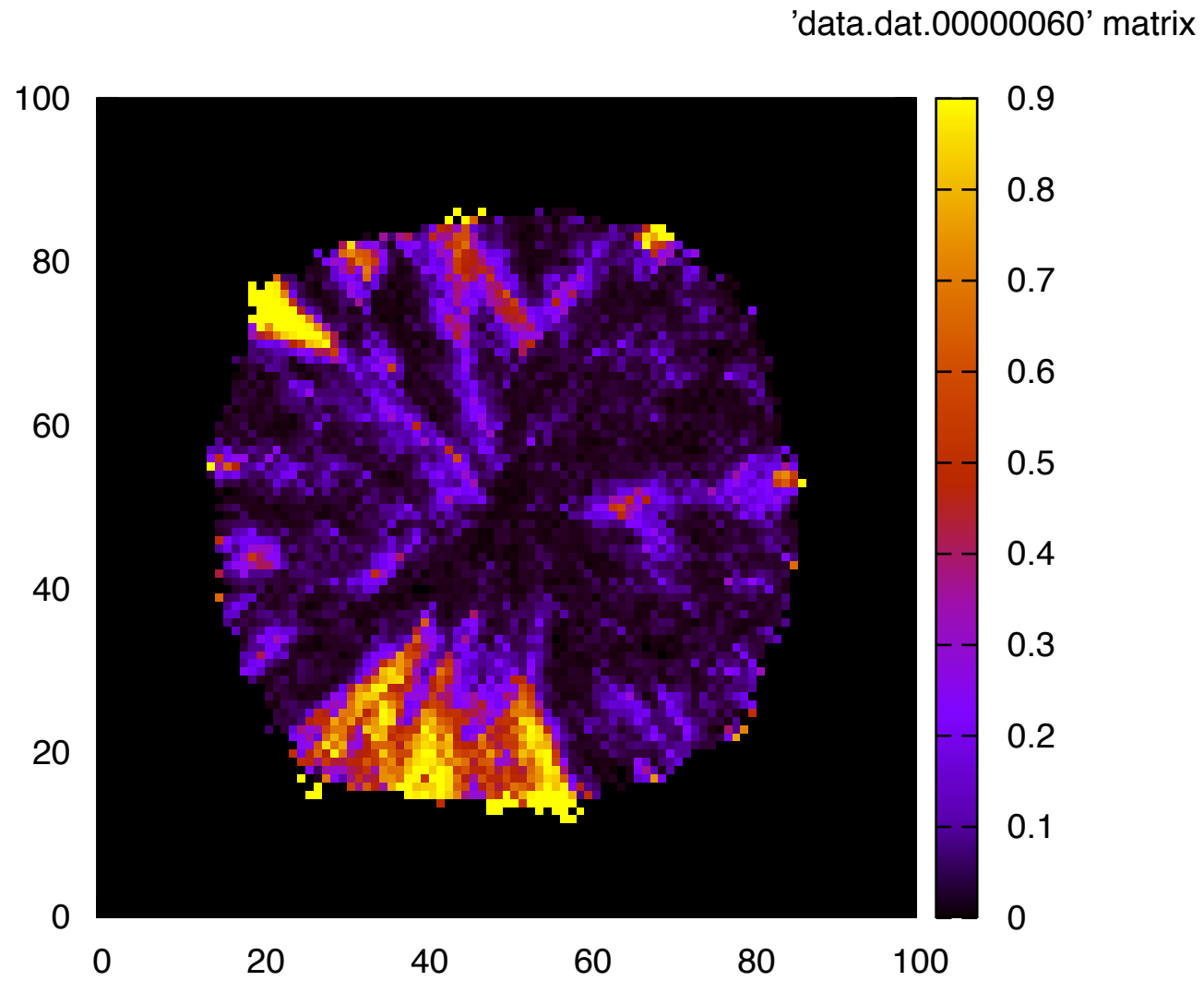


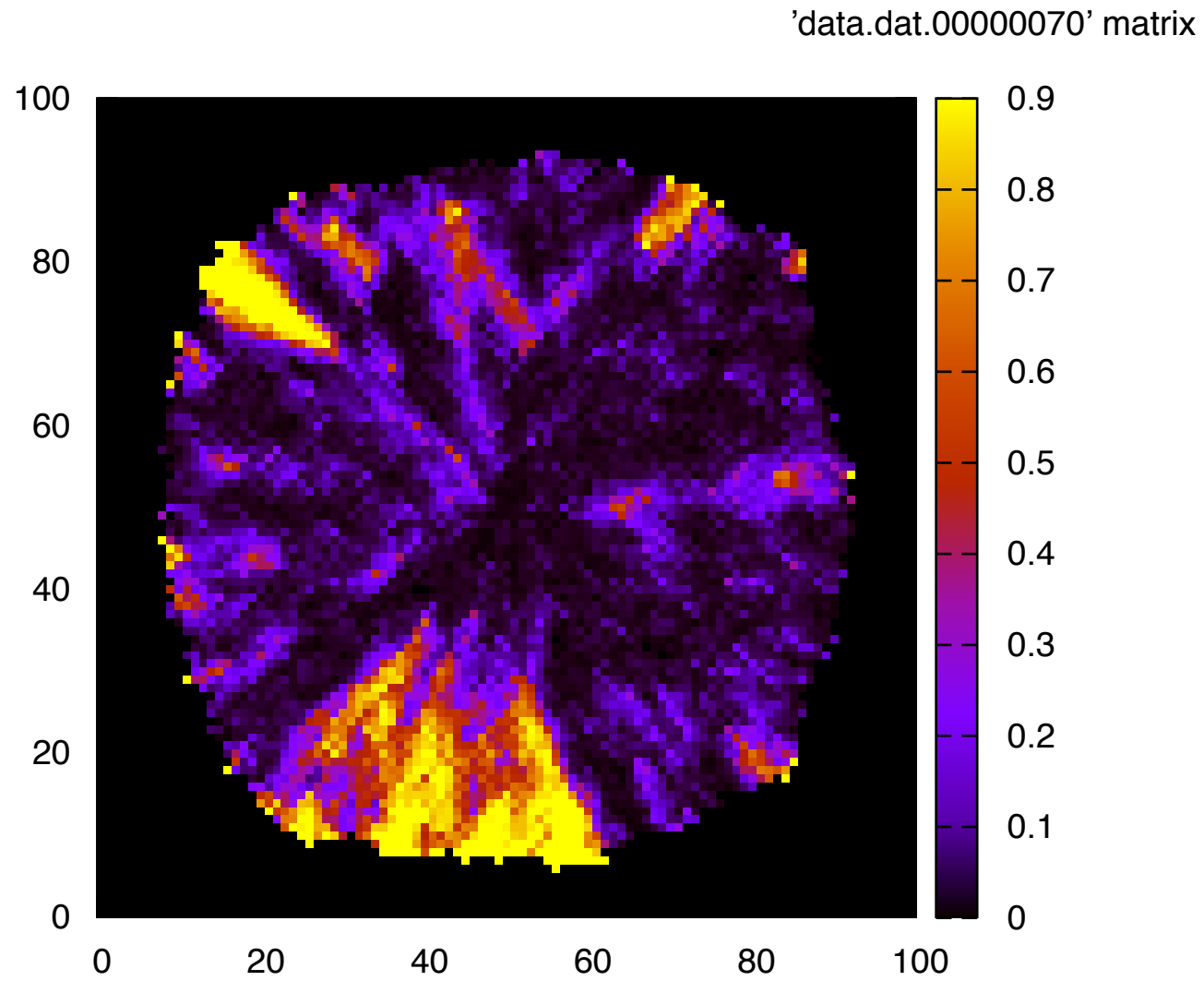


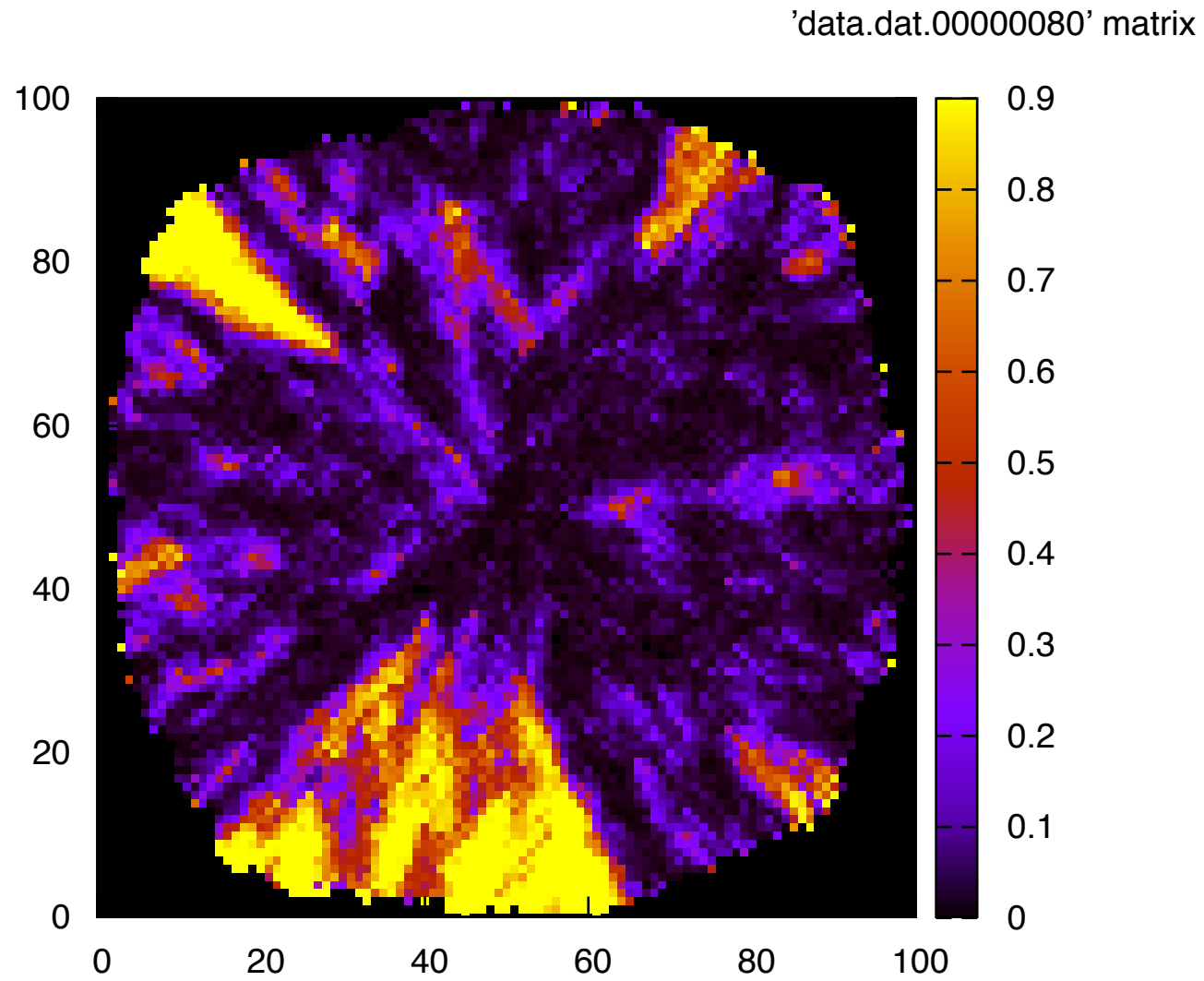


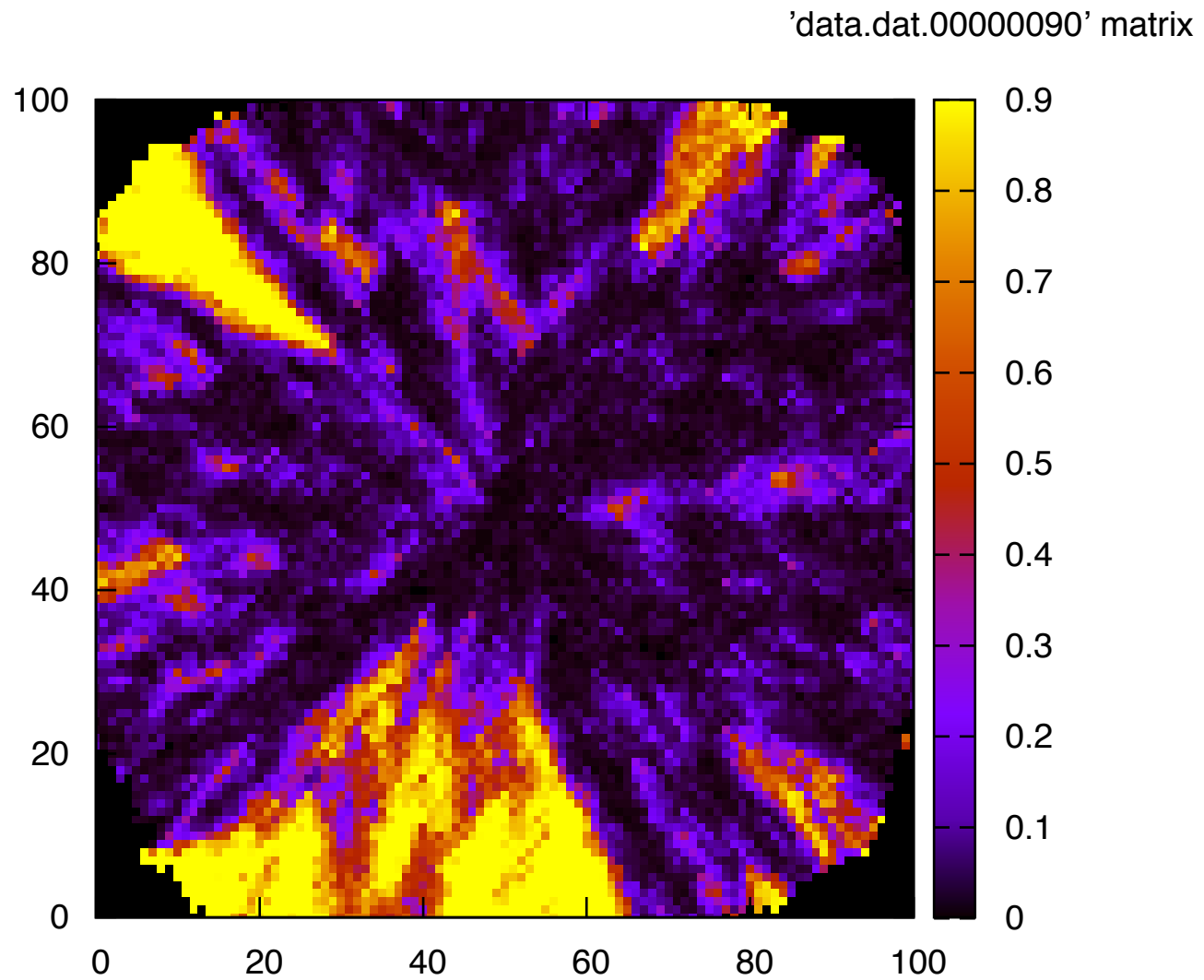




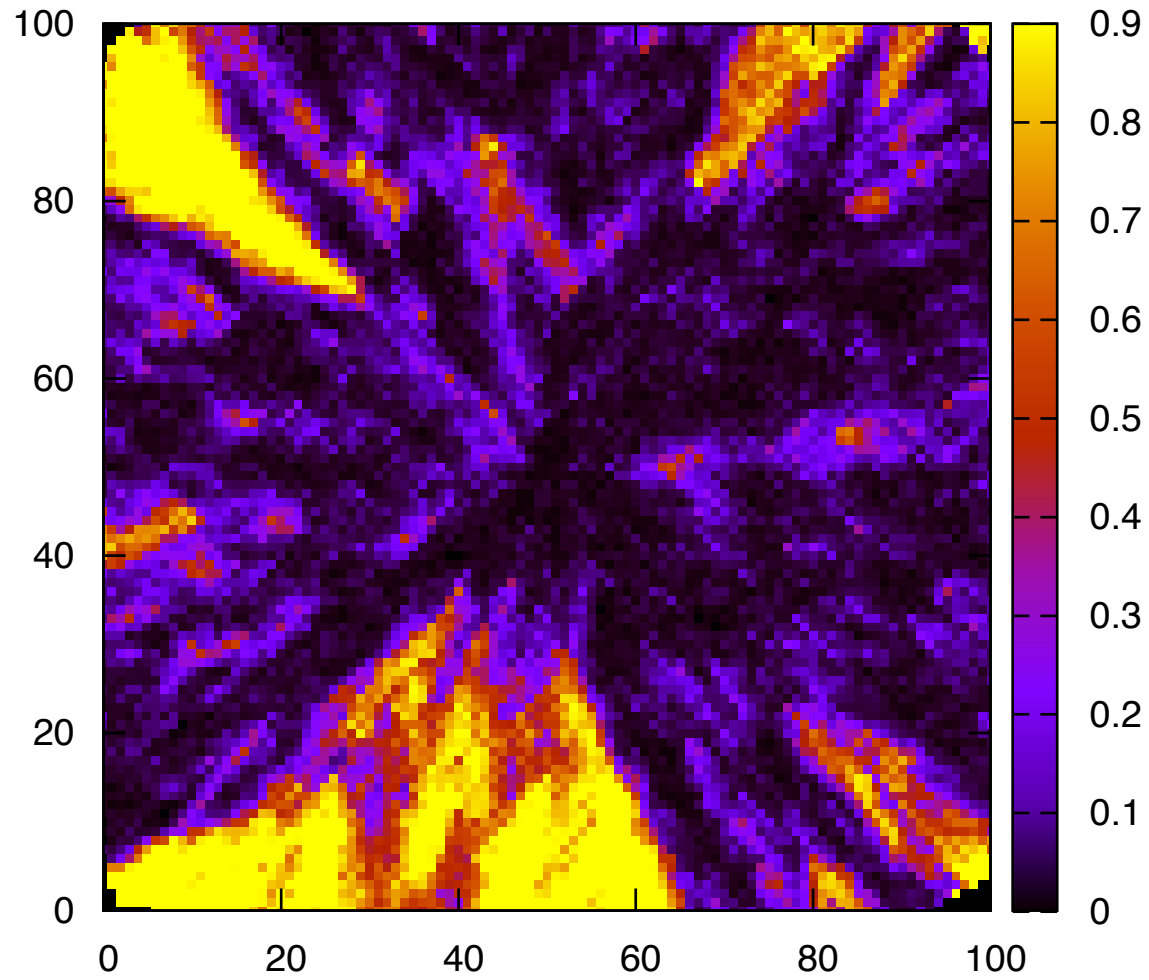


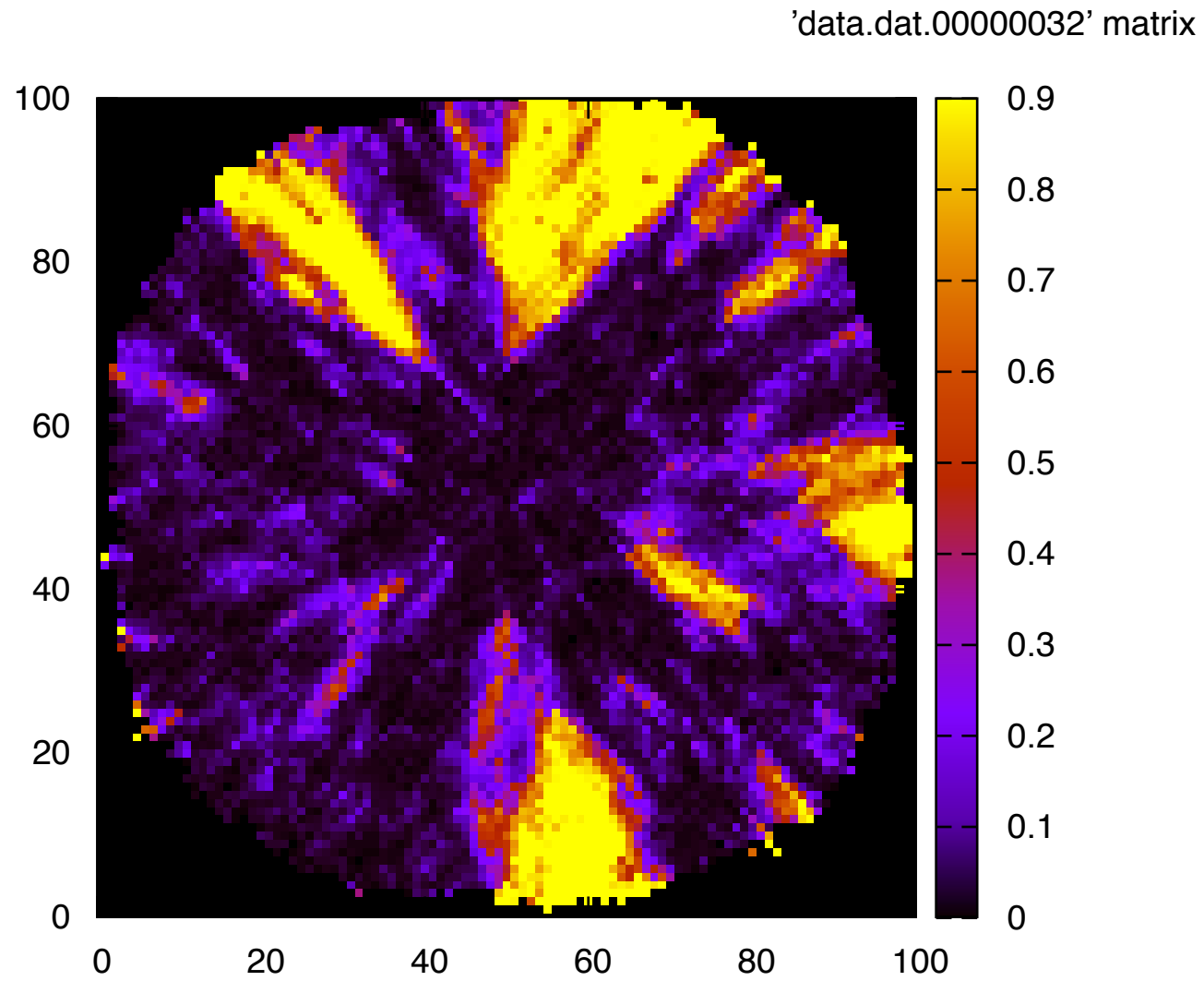


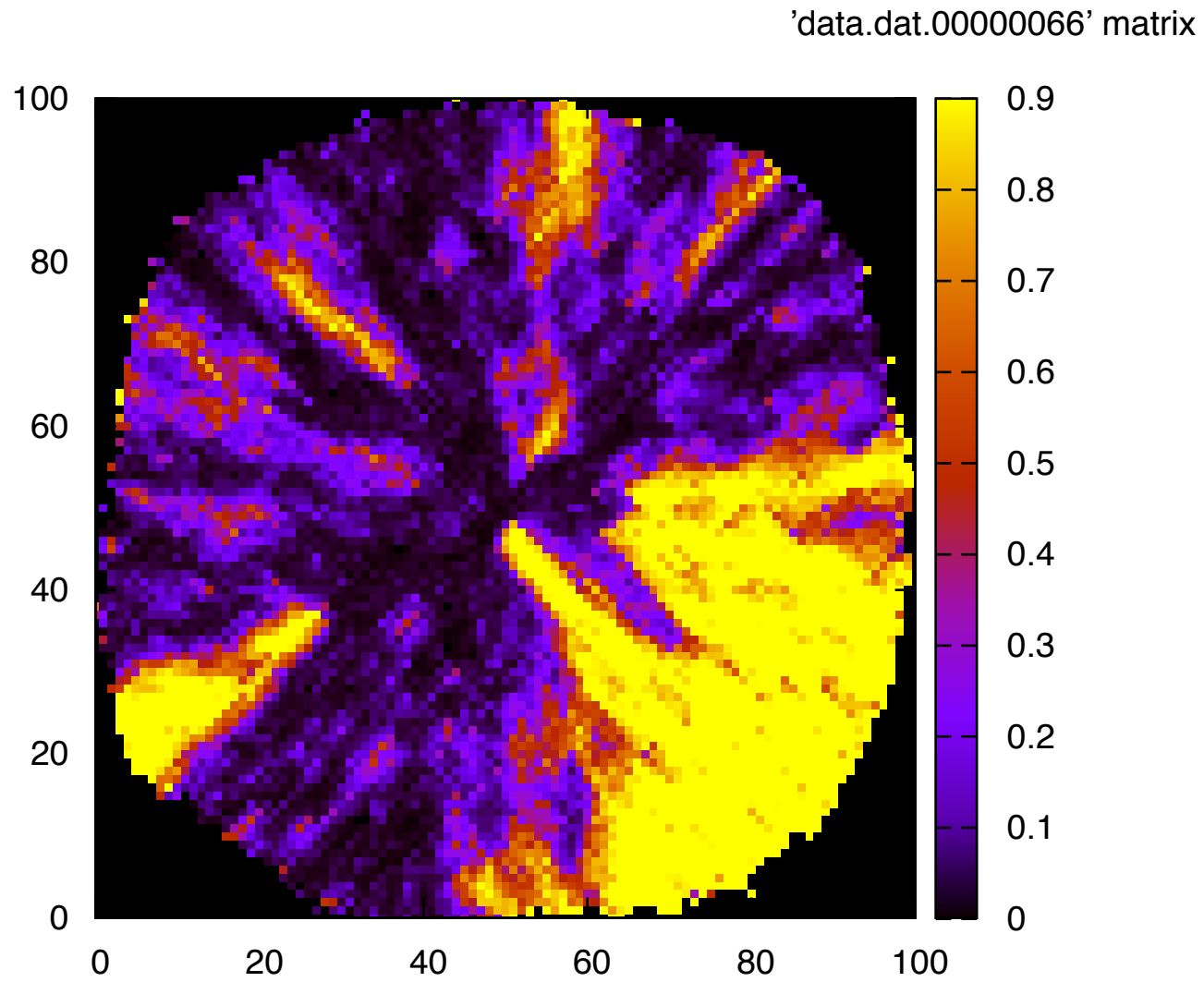




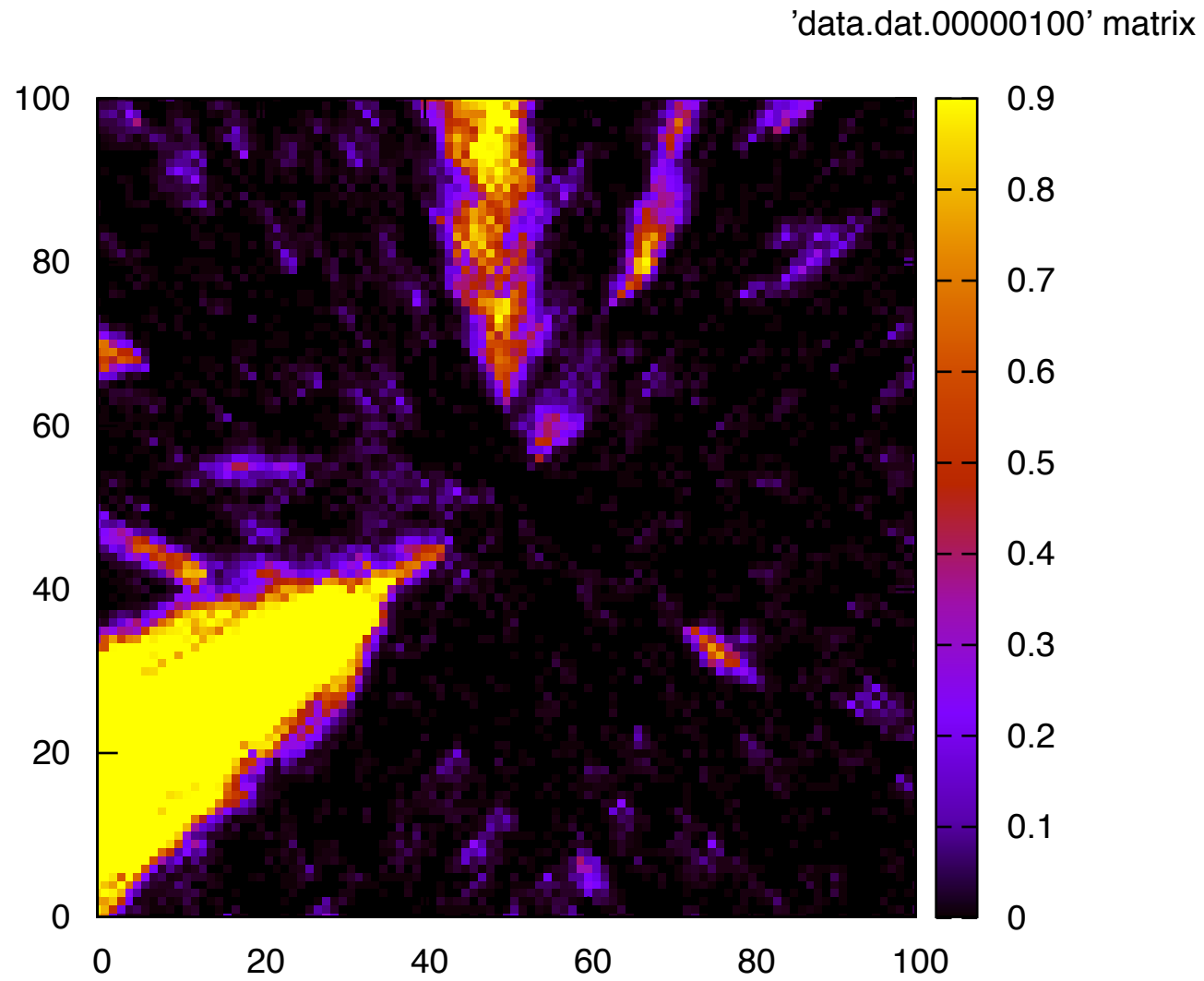
'data.dat.00000100' matrix



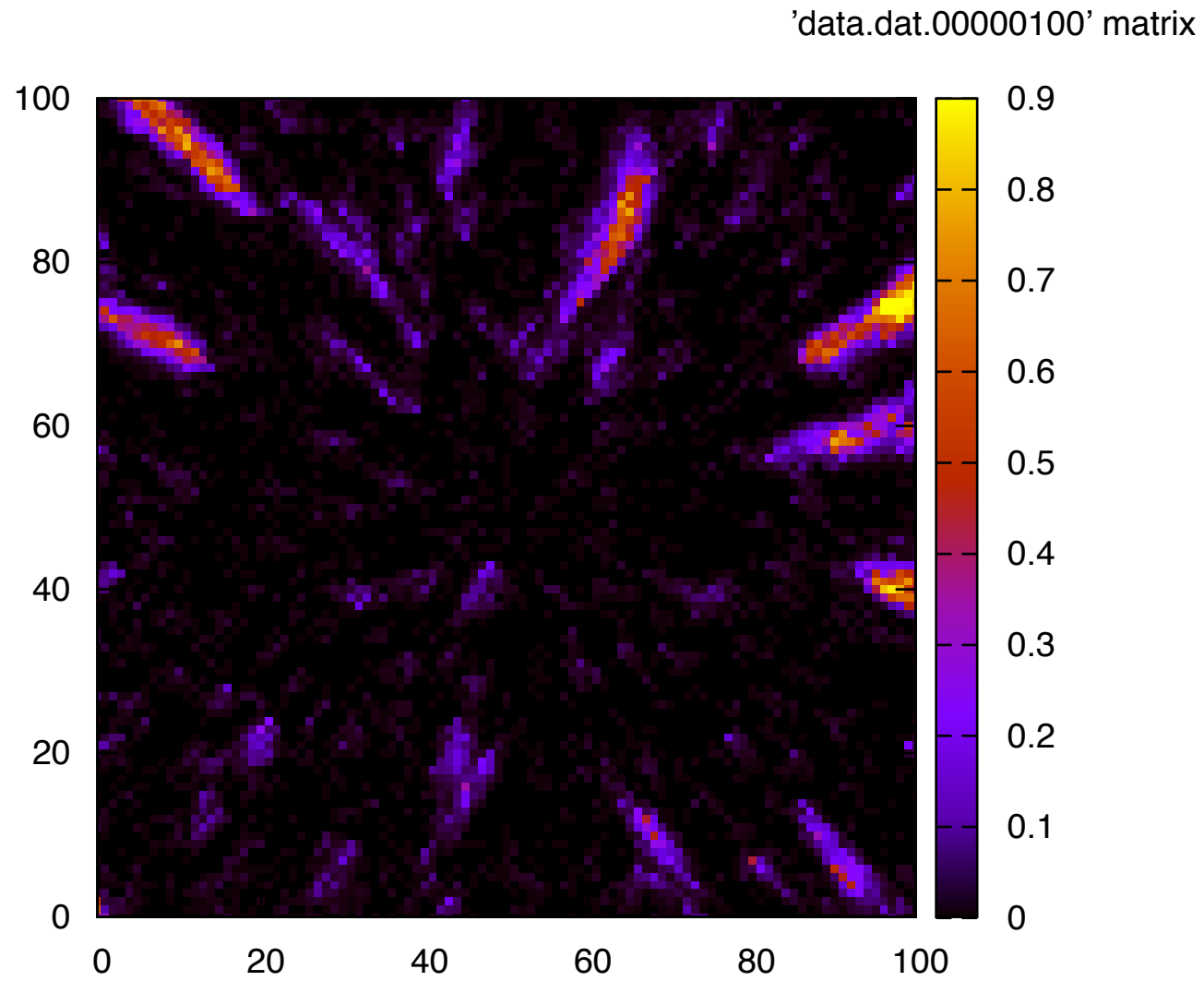




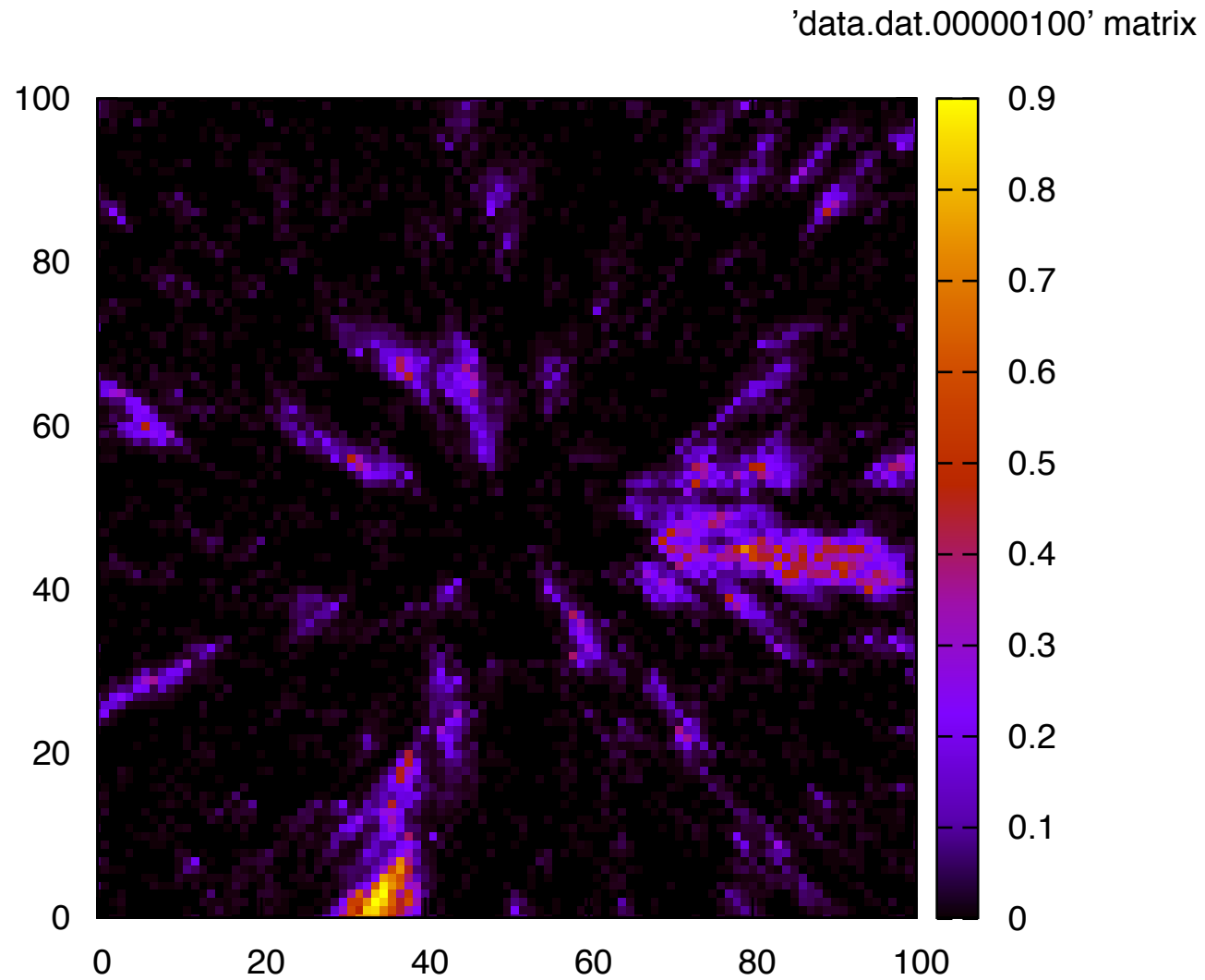
## Ex3: 2D - VARYING FITNESS, TRACE 1 (MEDIUM, F=1)



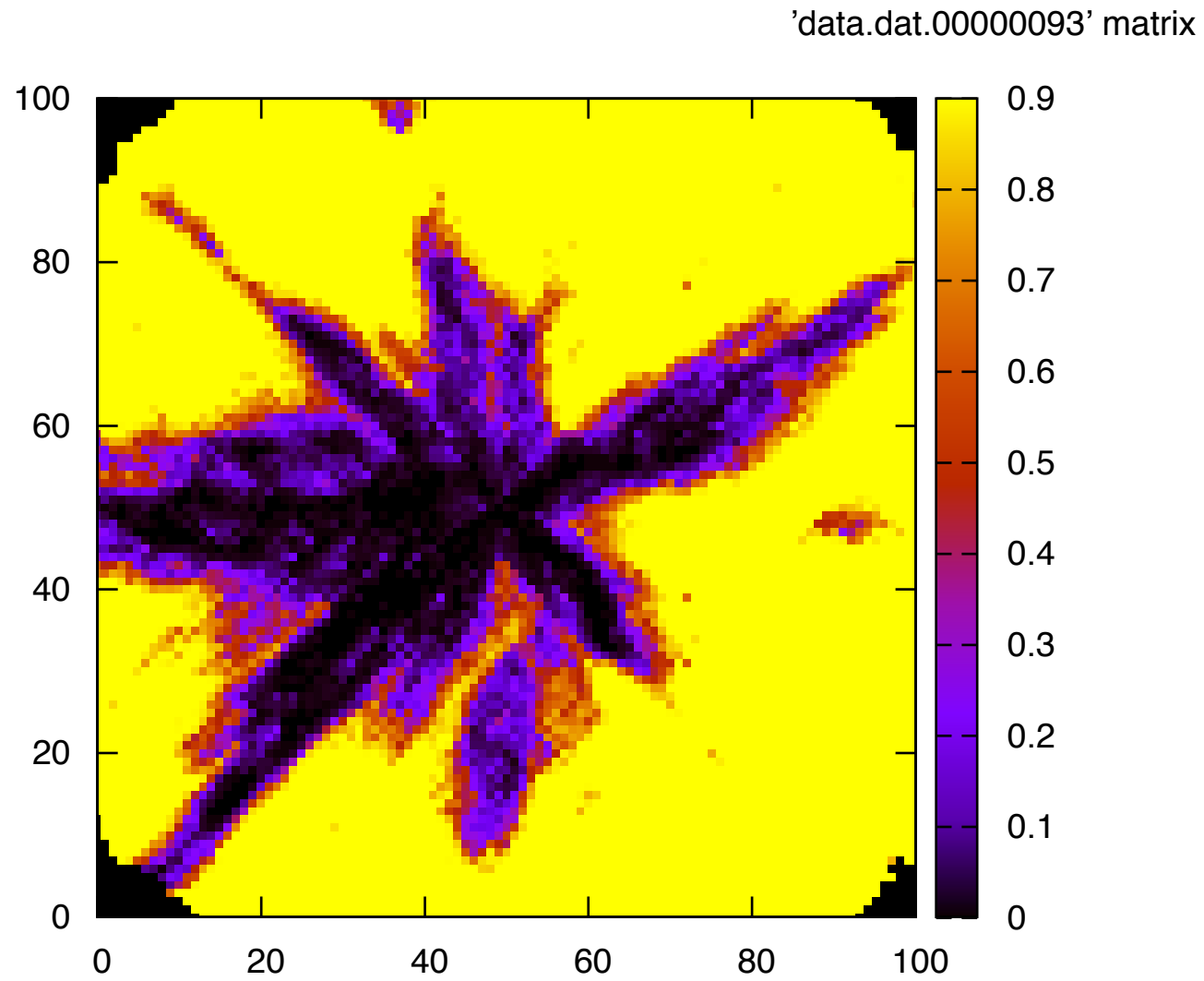




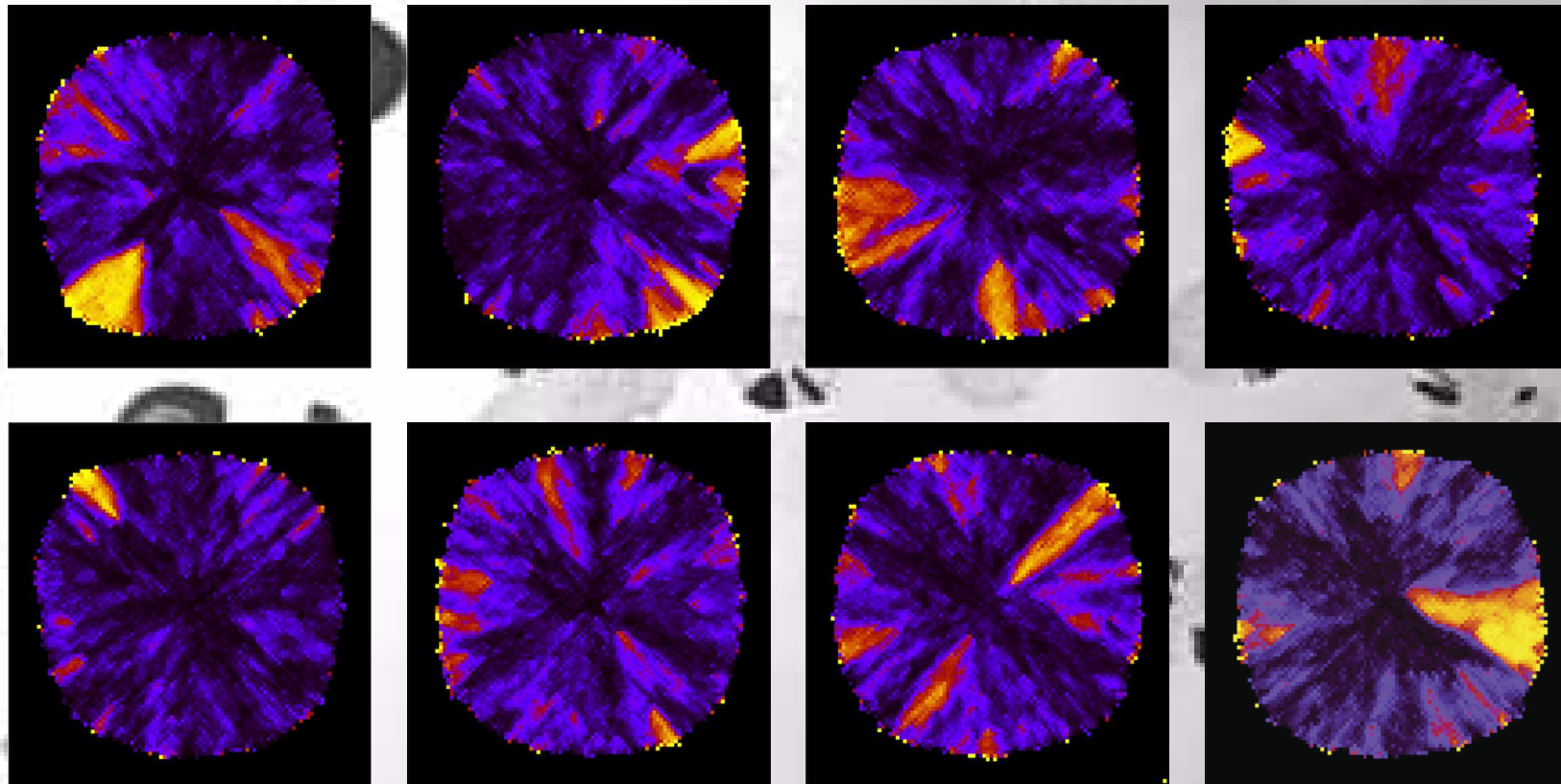
## Ex3: 2D - VARYING FITNESS, TRACE 1 (MEDIUM, $F=0.99$ )



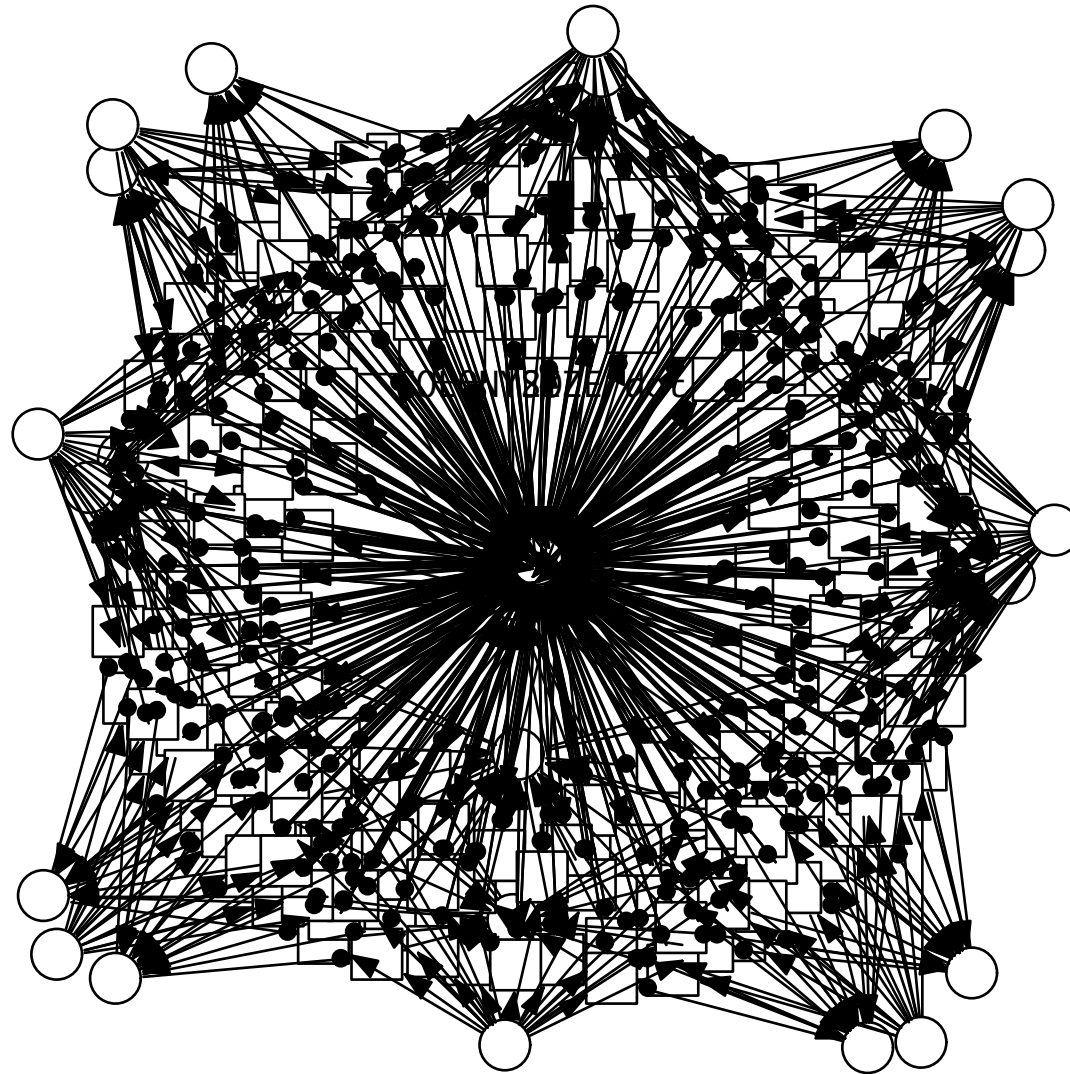
## Ex3: 2D - VARYING FITNESS, TRACE 1 (MEDIUM, $F=0.90$ )



## EX3: SOME FINAL STATES (HIGH, $F=1$ )

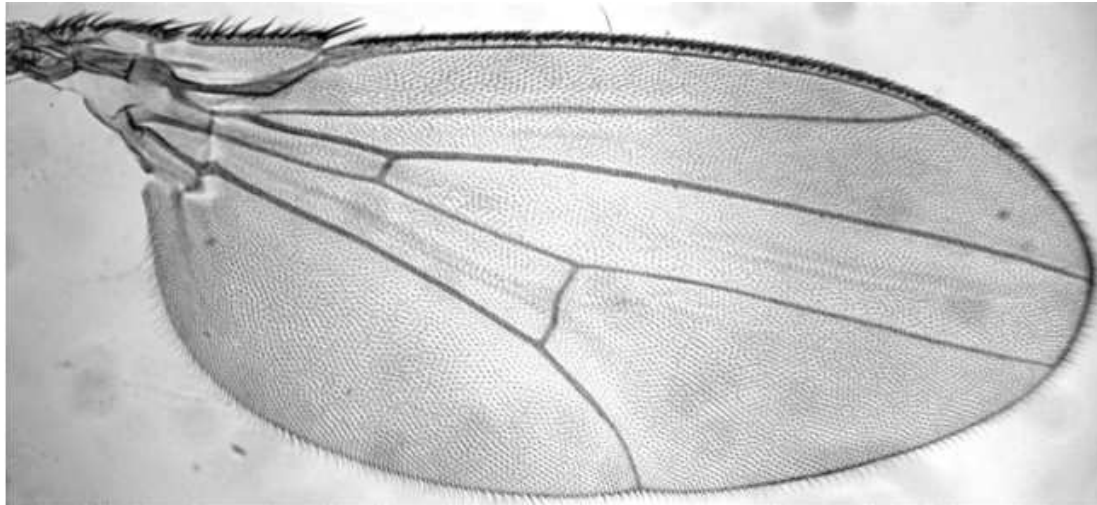


- ❑ **how to analyse visual data?**      **-> CMSB 2013**
  - > *auxiliary variables derived from model variables*
  - > *clustering techniques*
  - > *shape recognition*
  - > *visual analytics*
  
- ❑ **use model to predict**
  - > *mutation rates by measuring the mutation sectors,*  
*... or just the number of sectors?*
  - > *fitness by measuring angle of sectors*
  
- ❑ **possible model extensions / variations**
  - > *fine tuning of biofilm thickness*
  - > *multiple gene on/off and their dependencies*
  - > *log pedigree and/or mobility*

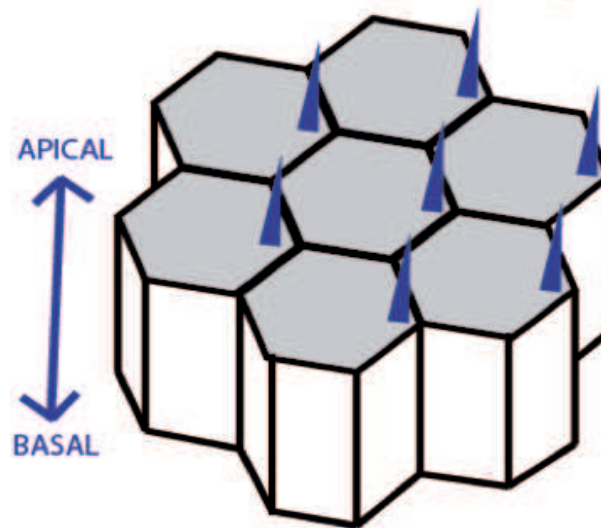


# **EXAMPLE 4: PLANAR CELL POLARITY IN FLY WING**

# EX4 - PLANAR CELL POLARITY

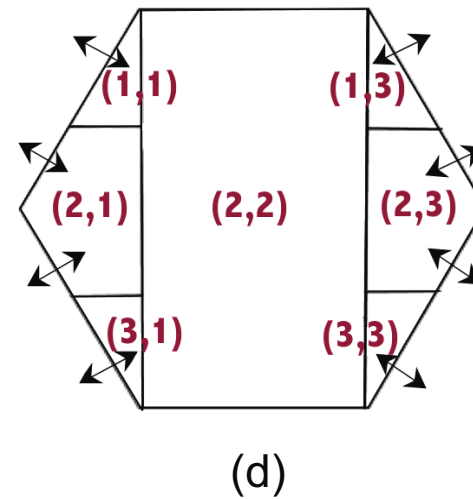
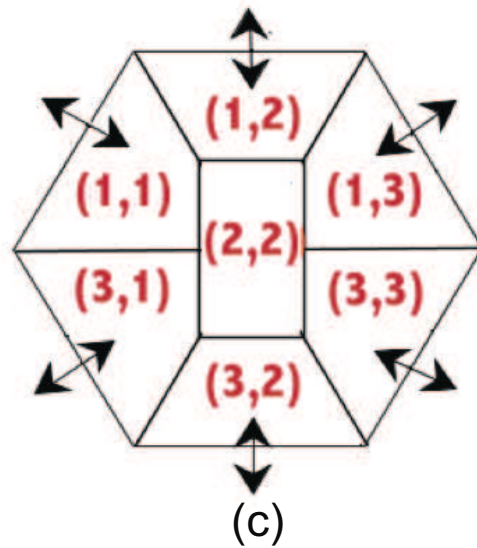
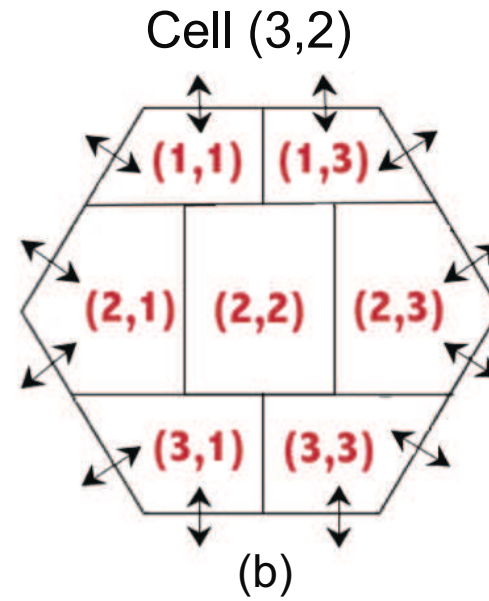
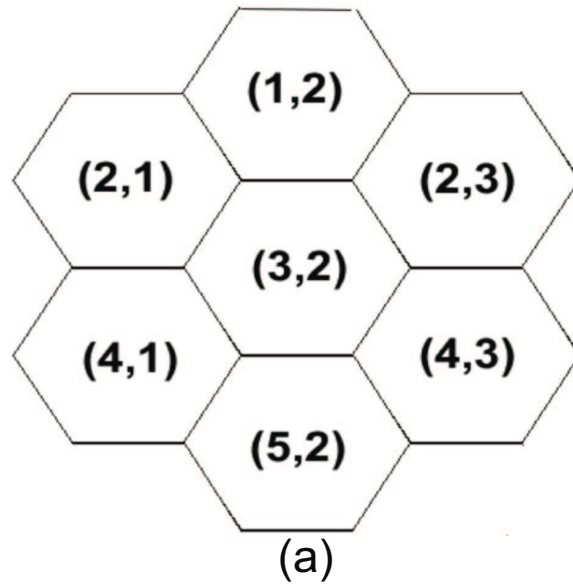


PROXIMAL ← → DISTAL

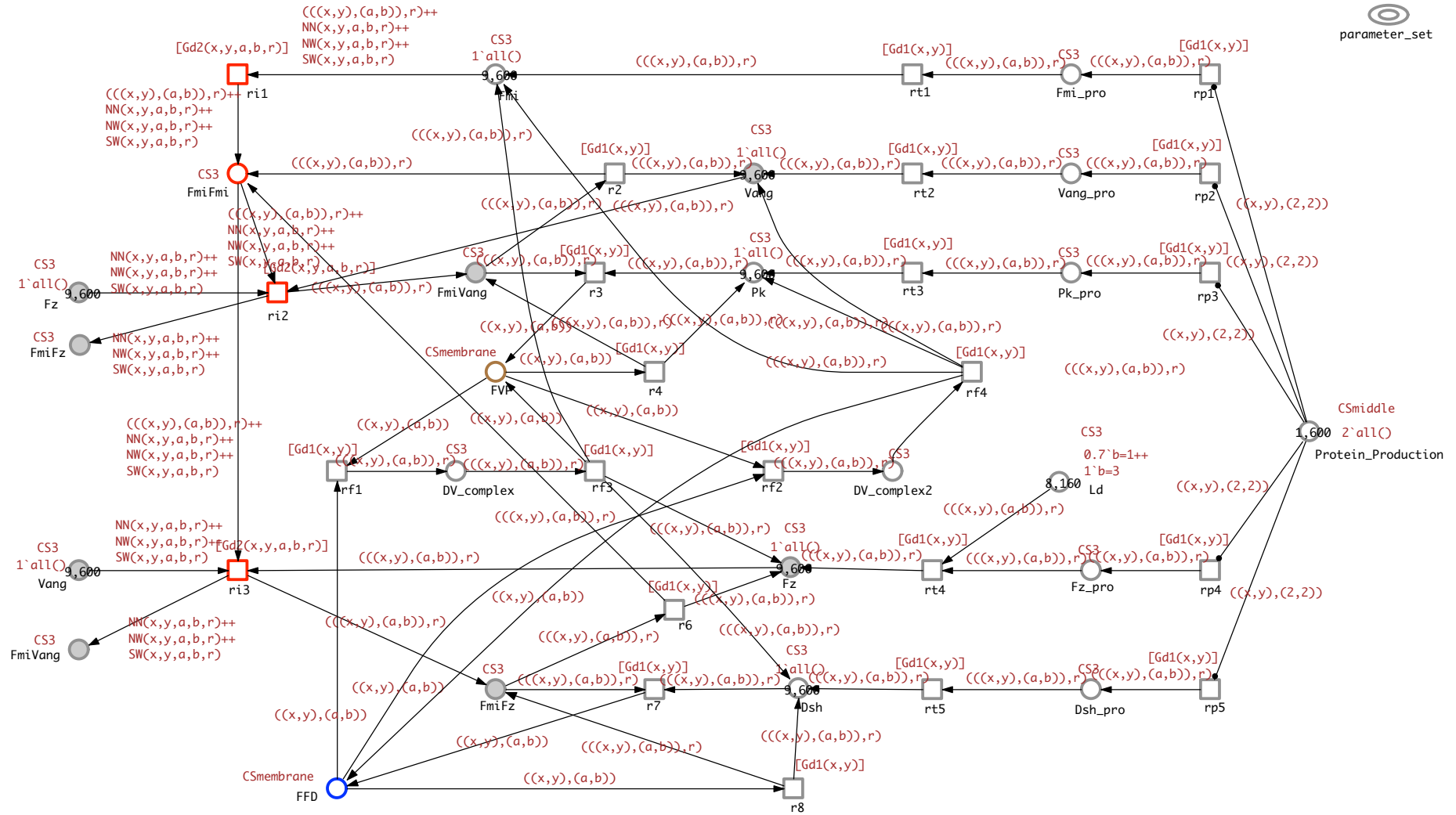


[BioPPN 2011]  
[CMSB 2011]



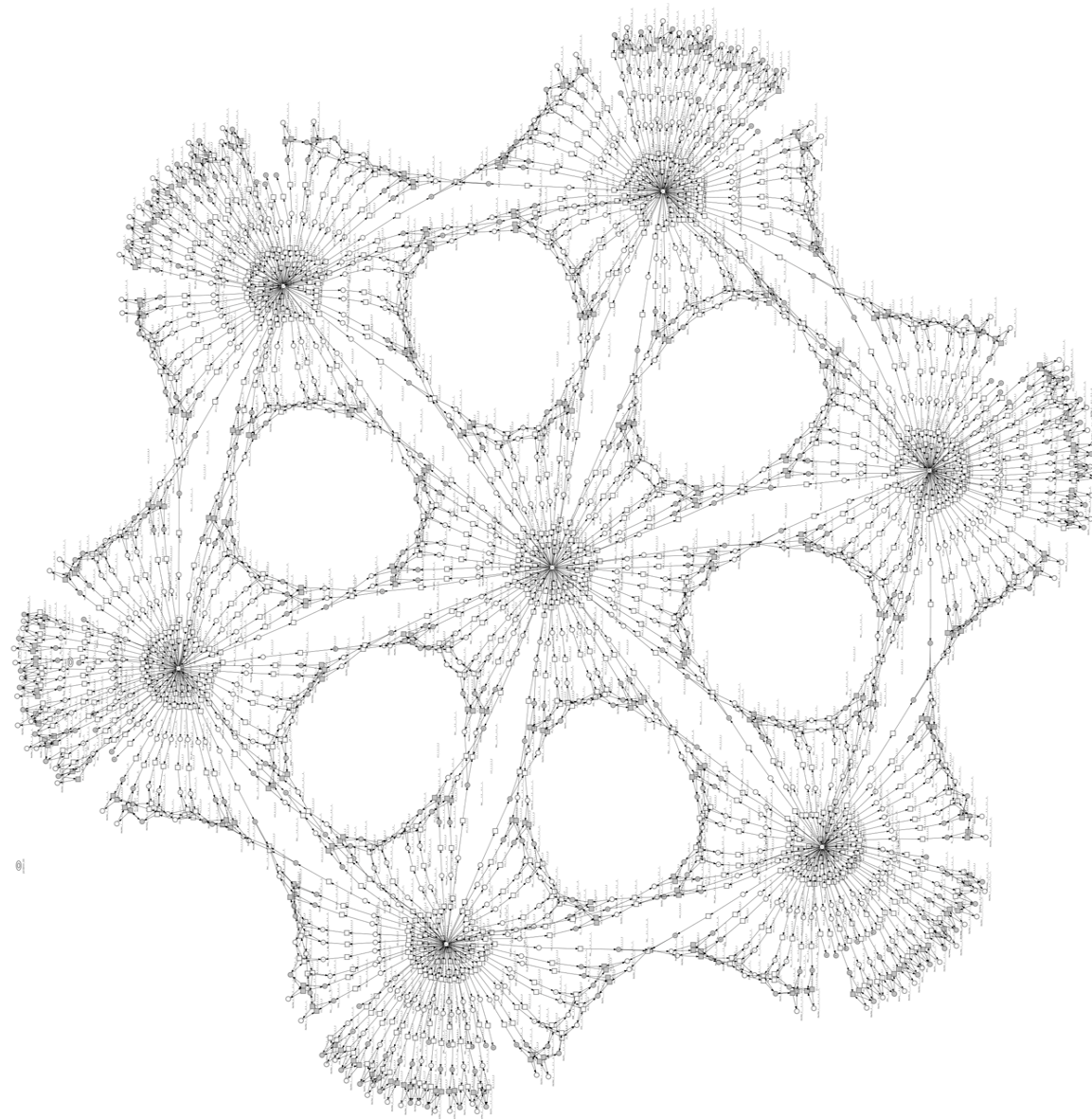


# Ex4: PLANAR CELL POLARITY

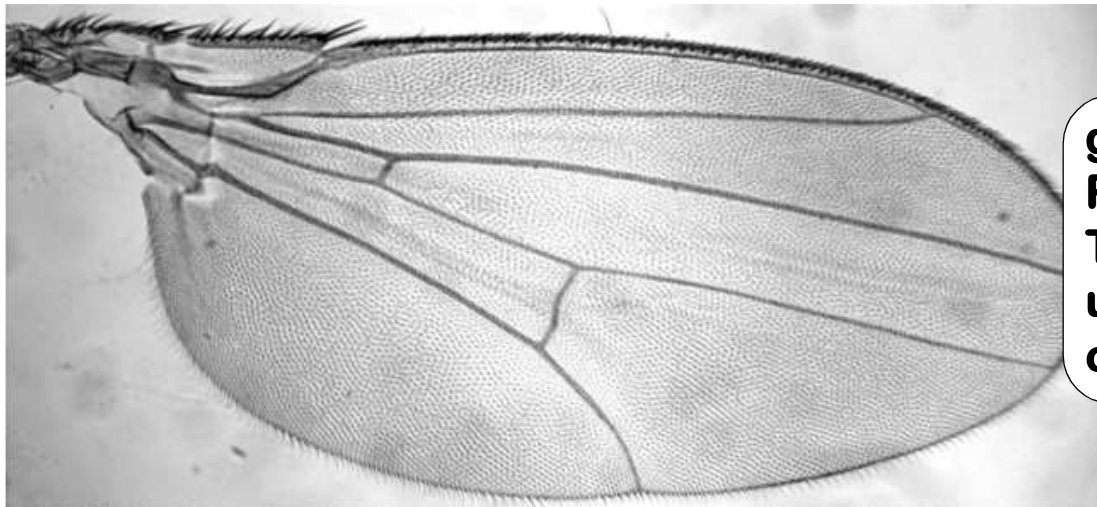


[QIAN GAO, PHD THESIS 2013]

# Ex4: PLANAR CELL POLARITY, PLAIN MODEL (7 CELLS)

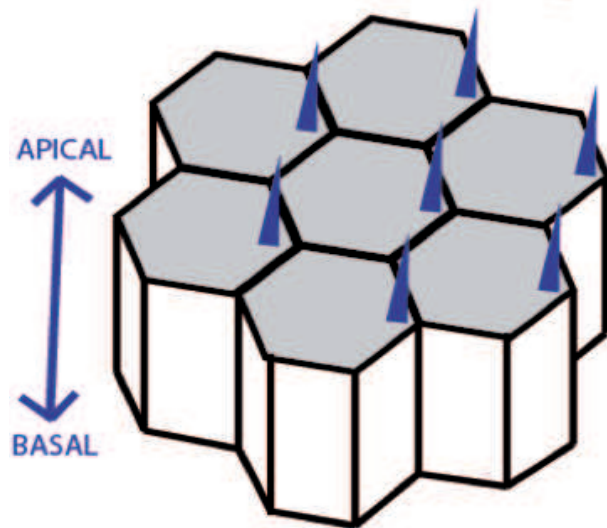


# EX4 - PLANAR CELL POLARITY



<b>grid size:</b>	<b>40 x 40</b>
<b>PLACES:</b>	<b>164,000</b>
<b>TRANSITIONS:</b>	<b>229,686</b>
<b>unfolding:</b>	<b>4 min</b>
<b>cont. simulation:</b>	<b>2 h</b>

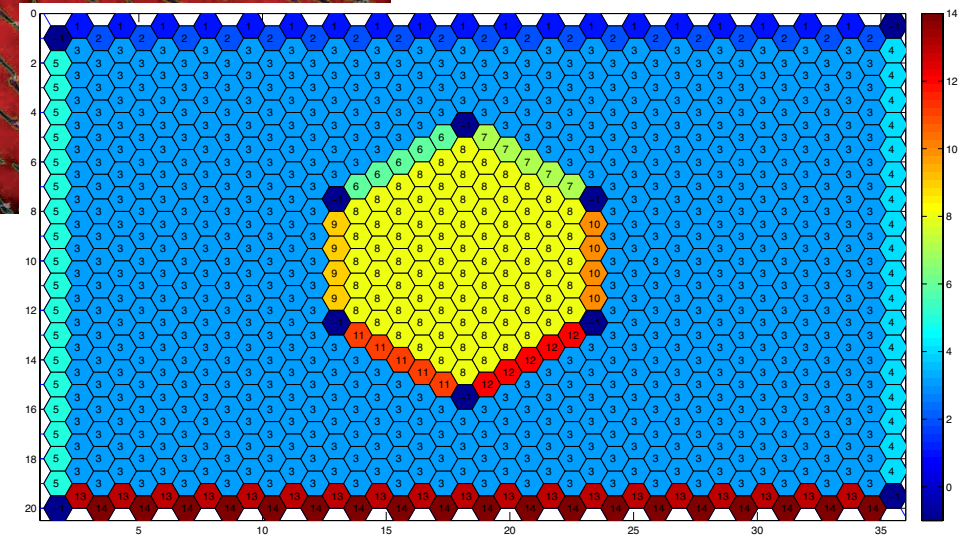
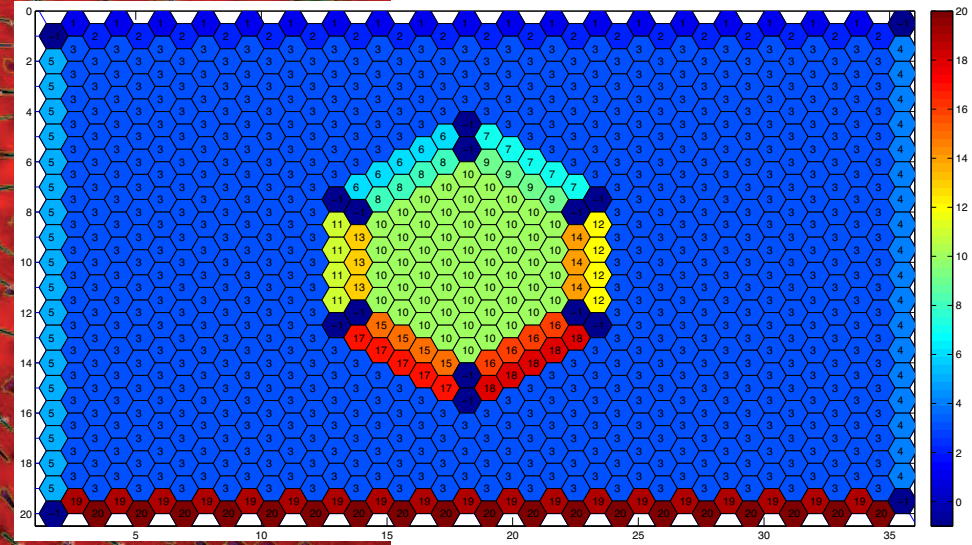
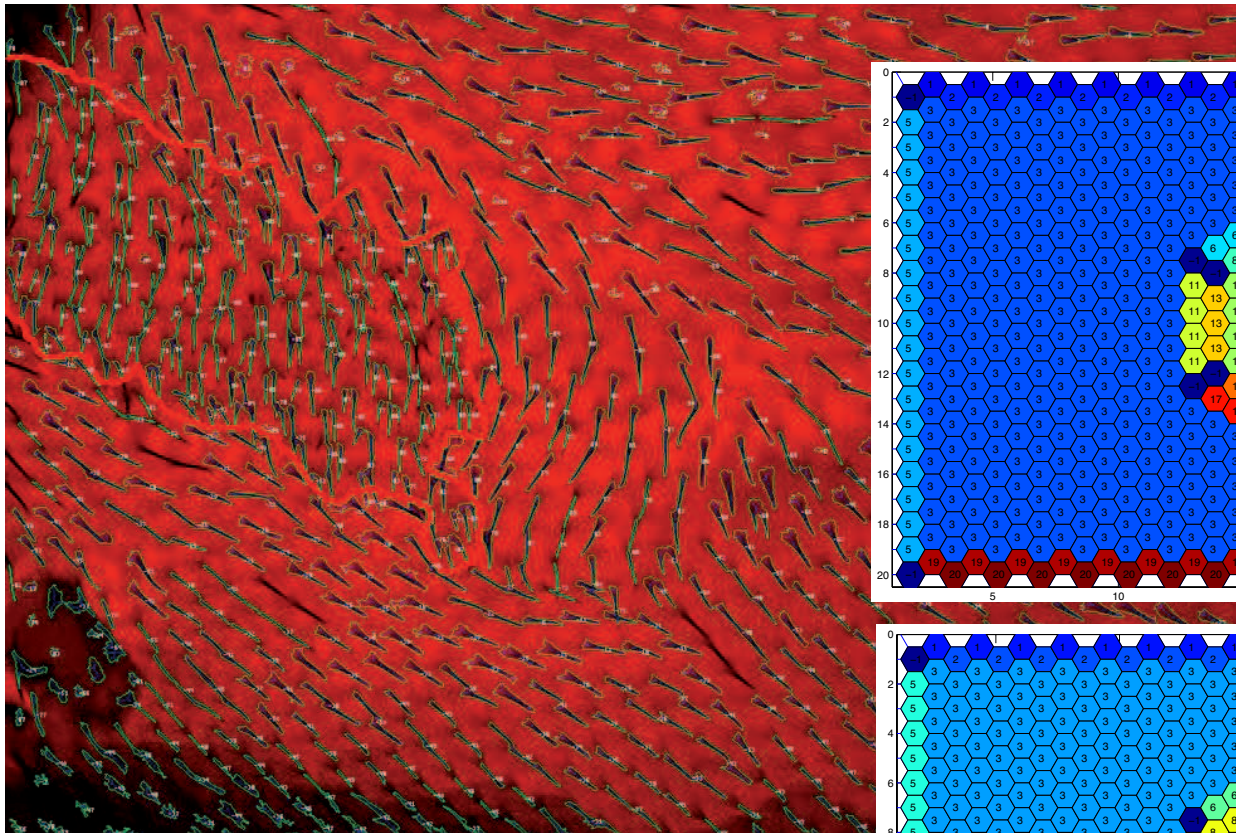
PROXIMAL ← → DISTAL



[BioPPN 2011]  
[CMSB 2011]



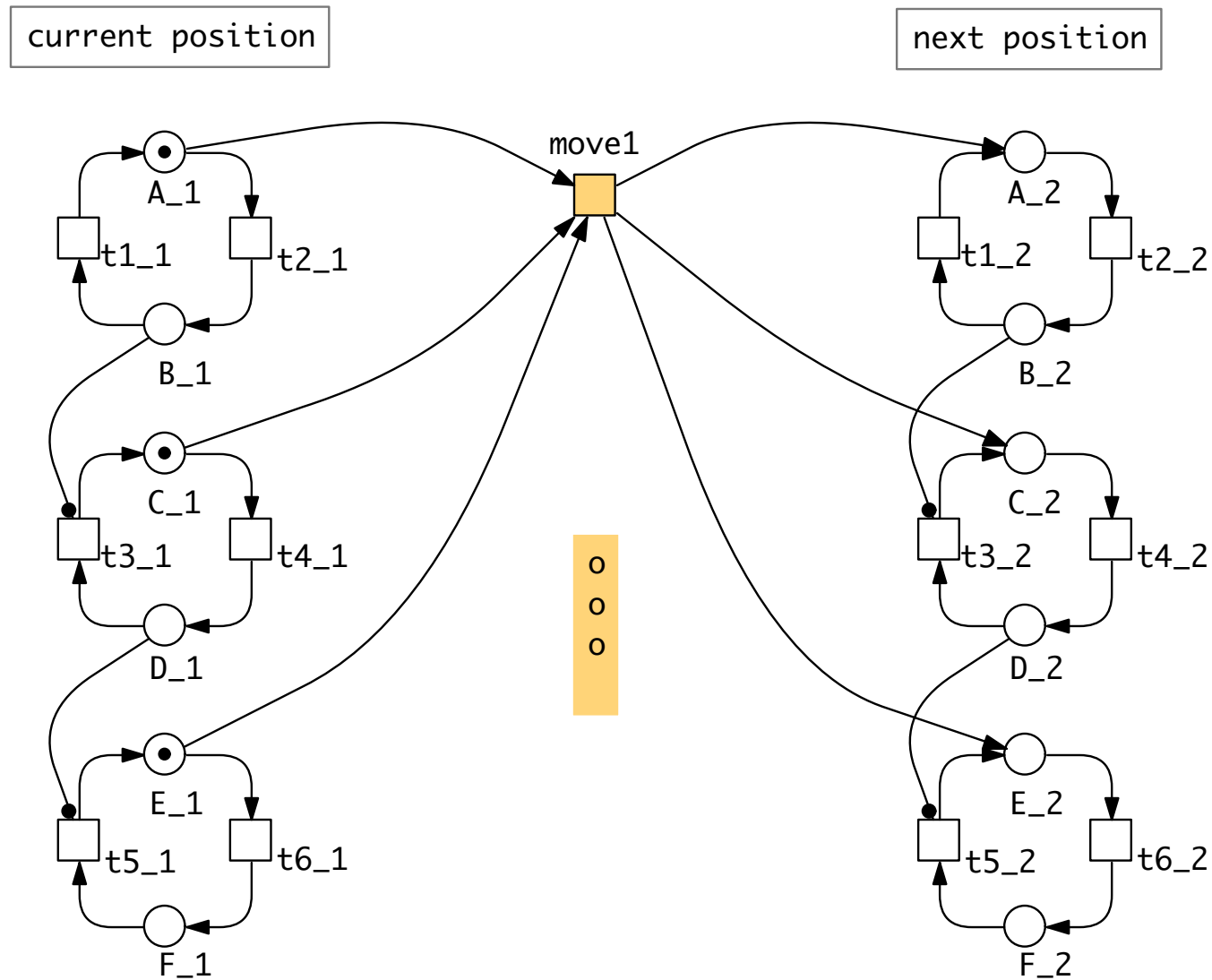
# Ex4 - PLANAR CELL POLARITY



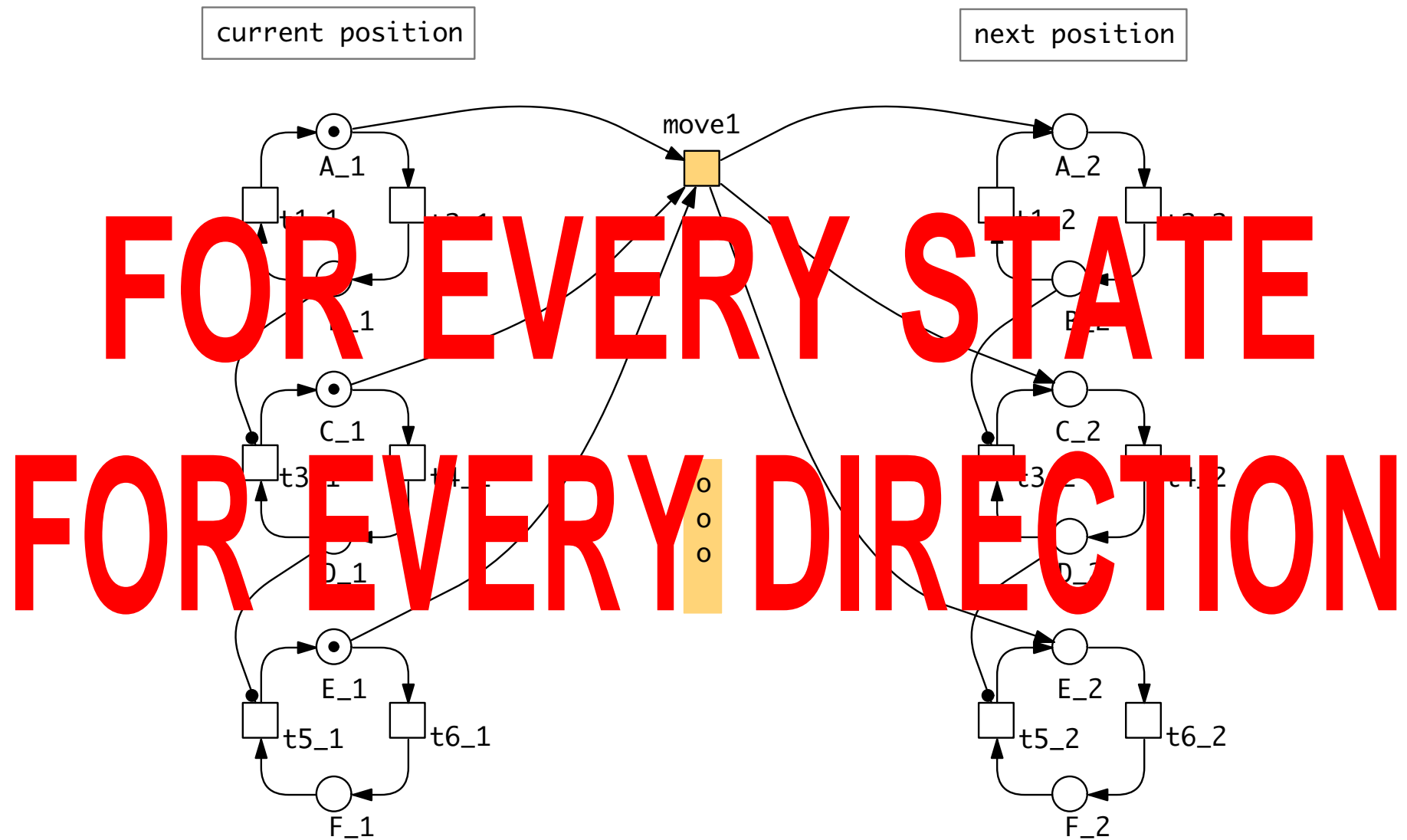
[TCBB 2012]

- ❑ **the spatial modelling principle can be equally applied to all paradigms**
  - > *qualitative, stochastic, continuous, and hybrid*
  - > *model transformations preserve all spatial attributes*
  
- ❑ **all space-related information is encoded in colour**
  - > *reuse in other models*
  
- ❑ **changing the notion of space**
  - > *adapt colour-related definitions*
  - > *net structure itself needs not to be touched.*
  
- ❑ **use of a priori finitely discretised space preserves model analysibility**  
  
----
  
- ❑ **automatic unfolding**
  - > *reuse of all analysis and simulation techniques of uncoloured Petri nets*

# HOW TO ENCODE SPACE, VERSION 1 - THE BIG CONS

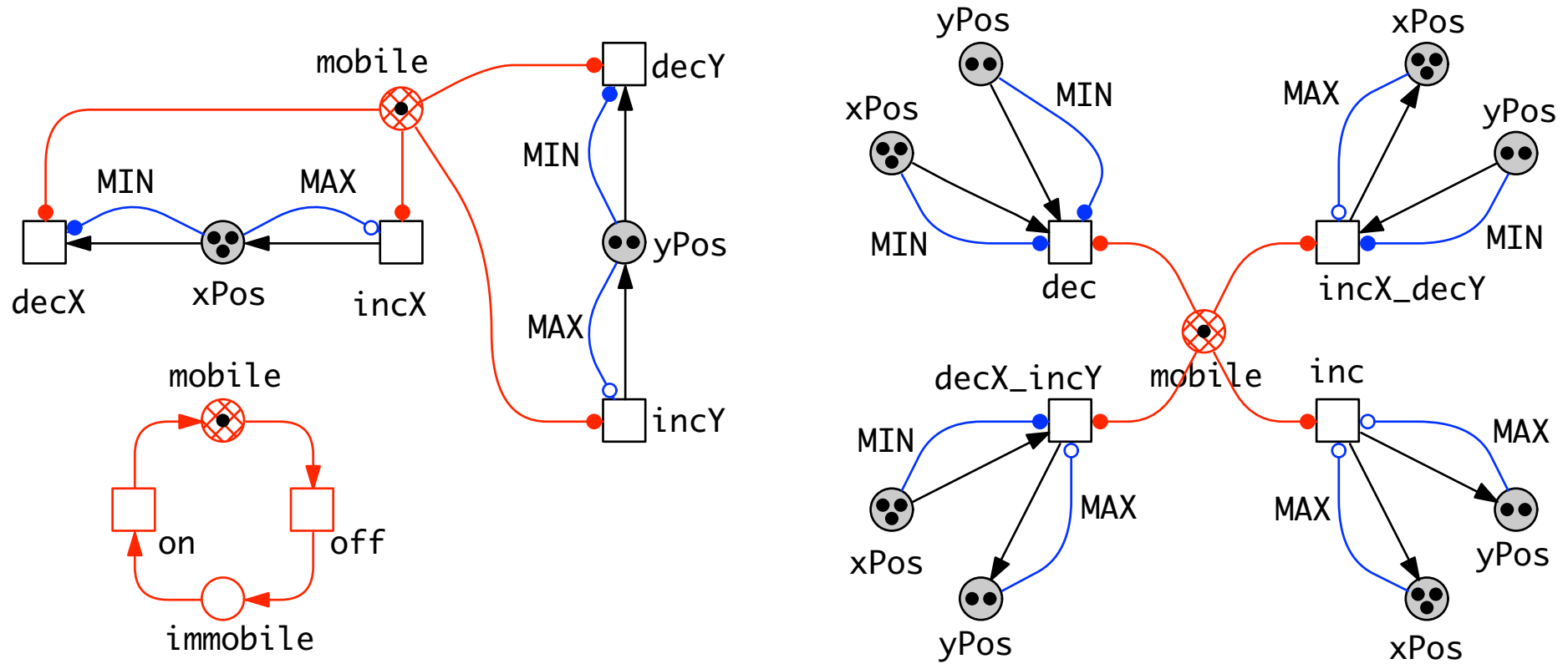






# HOW TO ENCODE SPACE ? VERSION 2

## *fence in*



## *permit movement on/off*

## VERSION1

- ❑ a priori finite space
- ❑ unfolding generates PN for the whole finite universe  
*-> many places might be empty*
- ❑ requires atomic moving objects

## PRO

- ❑ state-dependent rates as usual

## VERSION2

- ❑ potentially infinite space
- ❑ size of the unfolded PN does not depend on size of the universe
- ❑ local states in moving objects possible

## CON

- ❑ **state-dependent rates require special tool support**  
*-> observer variables*

# SUMMARY & OUTLOOK

### ❑ SNOOPY

- > *modelling and animation/simulation of hierarchical graphs, e.g. (extended) fault trees, various Petri net classes, e.g. QPN, XQPN, SPN, XSPN, CPN, TPN, . . . , free style graphs*

### ❑ CHARLIE

- > *QPN, XQPN, Time/Timed Petri nets (TPN)*
- > *mostly standard analysis techniques of Petri net theory*

### ❑ MARCIE

- > *XQPN, SPN, XSPN, SRN*
- > *symbolic and simulative model checking*

### ❑ Patty

- > *animation via web browser*

### □ SNOOPY

- > modelling and animation/simulation of hierarchical graphs, e.g. (extended) fault trees, various Petri net classes, e.g. QPN, XQPN, SPN, XSPN, CPN, TPN,



### □ CHAPLAIN **SBML import/export**

- > QPN, XQPN, Time/Timed Petri nets (TPN)
- > mostly standard analysis techniques of Petri net theory

### □ MARCIE

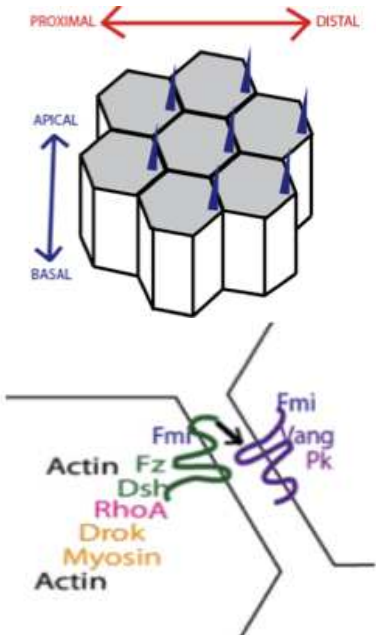
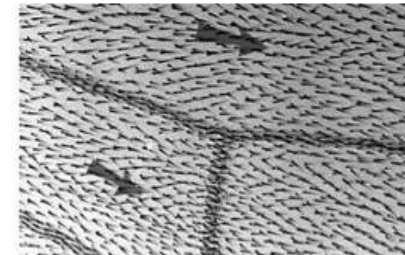
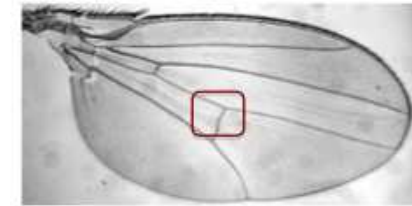
- > XQPN, SPN, XSPN, SRN
- > symbolic and simulative model checking

### □ Pattr

- > animation via web browser

**EXPORT TO MATAB AND  
MANY OTHER TOOLS**

- ❑ **efficient simulation of very large Petri nets**
  - > *stochastic*
  - > *continuous*
  - > *hybrid*
- ❑ **(hierarchical) space**
- ❑ **hierarchical organisation of components**
- ❑ **observables**
- ❑ **dynamic grid size**
- ❑ **shape and volume of components**
- ❑ **biosystem development**



## Multiscale Challenges



❑ **EPSRC Research Grant EP/I036168/1**

❑ **collaborators**

*David Gilbert, Brunel University, London, UK*

*Wolfgang Marwan, Otto-von-Guericke University, Magdeburg, Germany*

❑ **case studies**

*Turing Patterns - Mary Ann Blätke, Fei Liu*

*bacterial colony - Ovidiu Parvu, David Gilbert, Nigel Saunders*

*PCP in fly wing - Pam Gao, David Gilbert, David Tree*

❑ **Snoopy + Charlie + Marcie development**

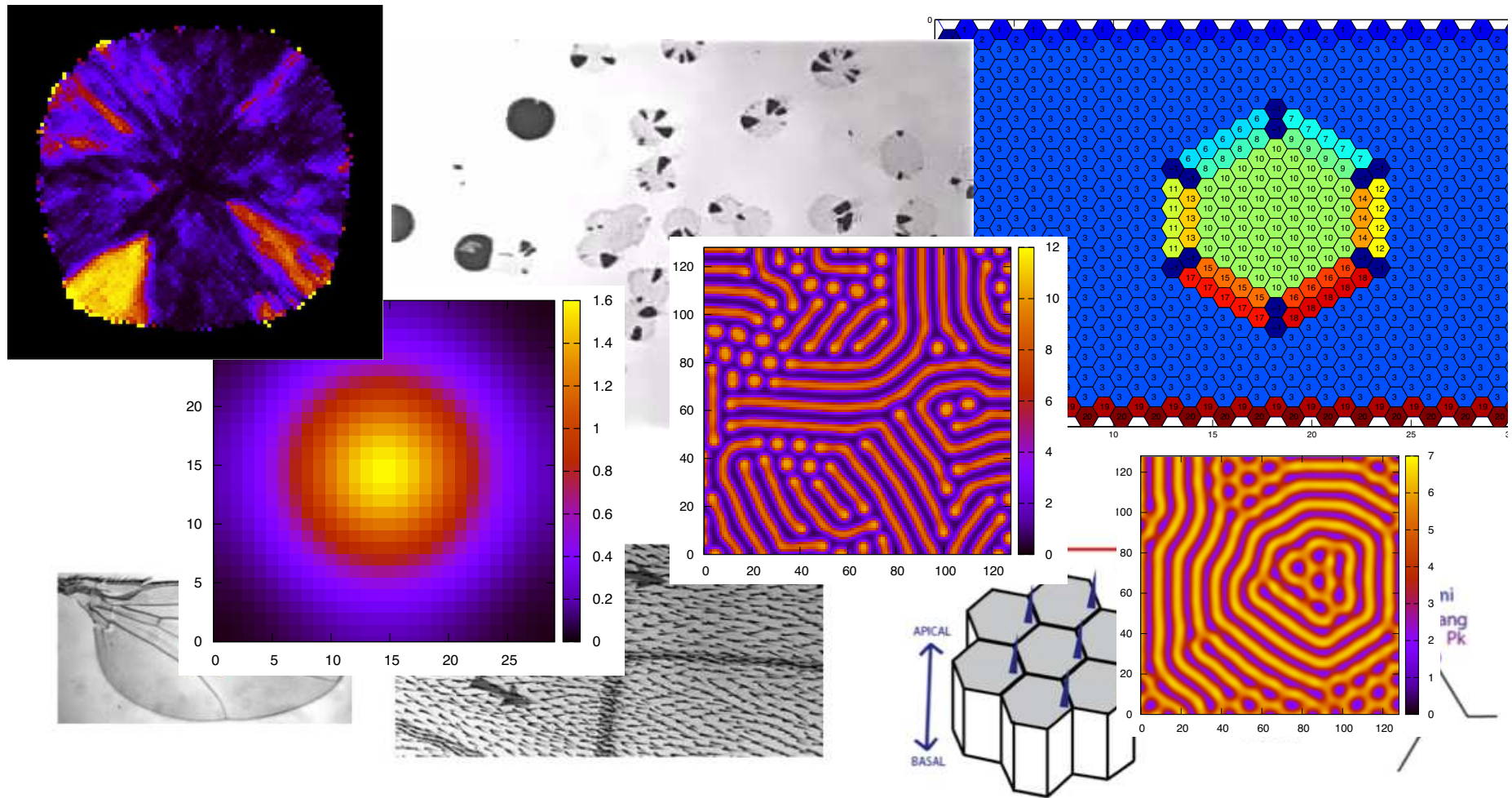
*Christian Rohr, Fei Liu, Mostafa Herayj*

*Martin Schwarick, Jan Wegener*

❑ **plots**

*Mary Ann Blätke, Daniele Maccagnola, Ovidiu Parvu, Christian Rohr, Jan Wegener*

THANKS !



[HTTP://MULTISCALEPN.BRUNEL.AC.UK](http://multiscalepn.brunel.ac.uk)